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SUMMARY

Hantaviruses are rodent-borne viruses causing serious zoonotic outbreaks worldwide for which no treatment is available. The hantavirus particles are pleomorphic and display a characteristic square surface lattice. The envelope glycoproteins Gn and Gc form heterodimers that further associate into tetrameric spikes, the lattice building blocks. The glycoproteins, which are the sole targets of neutralizing antibodies, drive virus entry via receptor-mediated endocytosis and endosomal membrane fusion. Here we describe the high-resolution X-ray structures of the heterodimer of Gc and the Gn head, and of the homotetrameric Gn base. Docking them into an 11.4 Å resolution cryo-electron tomography map of the hantavirus surface accounted for the complete extramembrane portion of the viral glycoprotein shell and allowed a detailed description of the surface organization of these pleomorphic virions. Our results, which further revealed an in-built mechanism controlling Gc membrane-insertion for fusion, pave the way for immunogen design to protect against pathogenic hantaviruses.

INTRODUCTION

Rodent-borne hantaviruses form a close group of viruses distributed worldwide, classified as Old World and New World hantaviruses (OWHs and NWHs) based on their geographical distribution and natural reservoirs (Jonsson et al., 2010). They cause life-threatening zoonotic outbreaks of severe pulmonary disease (NWHs) and of hemorrhagic fever with renal syndrome (OWHs). Despite the severity of these diseases, no efficient treatment is available. Inhalation of aerosols contaminated with infected rodent excreta is the major route of transmission, although person-to-person transmission of a pulmonary syndrome caused by Andes hantavirus (ANDV) has also been reported (Martinez et al., 2005; Padula et al., 1998; Pizarro et al., 2019). NWHs therefore have the potential to adapt to human-to-human air-borne transmission routes, increasing their epidemic potential.

The *Hantaviridae* constitute one of twelve families of segmented negative-strand RNA viruses in the recently established *Bunyavirales* order (Maes et al., 2019). As in most other members of this order (generically termed bunyaviruses), the viral genome is composed of three negative-polarity, single-stranded RNA segments: small, medium and large. The medium (M) segment encodes a polyprotein precursor that is matured in the ER by signalase cleavage to generate two envelope glycoproteins, Gn and Gc (Figure 1A) (Lober et al., 2001). Gn interacts co-translationally with the membrane fusion protein Gc, maintaining it in a metastable pre-fusion conformation within a Gn/Gc heterodimer. The protomers further associate into $(Gn/Gc)_4$ tetrameric spikes that are transported to the site of particle morphogenesis, the Golgi apparatus or the plasma membrane depending on the virus (Cifuentes-Munoz et al., 2014). The lateral interactions between adjacent spikes are believed to induce the membrane curvature required for budding of nascent virions (Huiskonen et al., 2010). Hantavirus particles are internalized into target cells via receptor-mediated endocytosis, with the glycoprotein shell reacting to the acidic endosomal pH to drive fusion of viral and cellular membranes through a conformational change of Gc (Acuna et al., 2015; Chiang et al., 2016; Cifuentes-Munoz et al., 2011; Jin et al., 2002; Mittler et al., 2019; Ramanathan and Jonsson, 2008; Rissanen et al., 2017).

The X-ray structures of the isolated Gn "head" (Gn^H, comprising the N-terminal 2/3[rds] of the ectodomain) (Li et al., 2016; Rissanen et al., 2017) and of the Gc ectodomain (Guardado-Calvo et al., 2016; Willensky et al., 2016) of Puumala (PUUV) and Hantaan (HNTV) hantaviruses (both OWHs) have been reported. Gc was shown to feature the three characteristic β-sheet rich domains (I, II and III) observed in the class II fusion proteins of the flaviviruses (Rey et al., 1995), alphaviruses (Lescar et al., 2001), phleboviruses (Dessau and Modis, 2013) and rubella virus (DuBois et al., 2013). The available structural snapshots of Gc show a monomer in an extended conformation stabilized by an antibody bound to domain II, and a Gc trimer in the characteristic post-fusion "hairpin" conformation

observed in other viral class II fusion proteins (Bressanelli et al., 2004; Gibbons et al., 2004; Luca et al., 2013; Modis et al., 2004).

The hantavirus square surface lattice was initially observed by negative stain electron microscopy (Martin et al., 1985) and later characterized by cryo-electron tomography (cET) studies on particles of the OWH Tula virus (Huiskonen et al., 2010) and by single particle averaging of the spikes on HTNV virions (Battisti et al., 2011). The cET studies showed pleomorphic particles with locally ordered lattices of tetrameric $(Gn/Gc)_4$ spikes coating the viral membrane. A model for the organization of Gn and Gc in the lattice was proposed (Hepojoki et al., 2010) in which Gn was located at the spike center and Gc at the periphery, respectively making intra- and inter-spike contacts. This model was in line with structural studies combining cET, sub-tomogram averaging (STA) and the X-ray structure of the isolated PUUV Gn^H (Li et al., 2016). It was further supported by functional studies confirming that the crystallographic 2-fold contacts made by HTNV Gc monomers (Guardado-Calvo et al., 2016) recapitulate the lateral interactions between spikes on particles (Bignon et al., 2019). Despite these advances, the actual organization of the spike and the molecular interactions between Gn and Gc have remained elusive.

Here, we report the X-ray structure of the metastable pre-fusion Gn^H/Gc heterodimers of ANDV and Maporal virus (MAPV, an NWH closely related to ANDV) to 3.2 and 2.2 Å resolution, respectively. The ANDV complex required a stabilizing mutation in Gc to avoid a spontaneous conformational change into its post-fusion conformation despite bound Gn^H. We also describe the X-ray structure of the Gn base (Gn^B, the C-terminal third of the ectodomain) to 1.9 Å resolution, showing that it forms a tight tetramer. These structures fitted unambiguously a cET map of the Tula virus Gn/Gc spike refined by STA to 11.4 Å resolution, confirming that the Gn^H/Gc and Gn^B conformations captured in the crystals indeed correspond to their active form in the spikes of infectious hantavirus particles and providing a complete model for the surface glycoprotein lattice. Functional studies by targeted mutagenesis further confirmed the detailed picture of the tetrameric spike organization as well as the lateral interactions between spikes. The Gn^H/Gc structure also revealed that, unlike other class II fusion proteins, Gn^H release at acid pH induces a further conformational change of the domain II tip to expose non-polar side chains for insertion into the endosomal membrane.

RESULTS

Structural characterization of the hantavirus glycoproteins

We produced the soluble ectodomains of the hantavirus Gn^H/Gc heterodimer using a single chain construct in which a soluble linker bypassed the Gn TM and intraviral/cytosolic segments to

connect directly to the Gc N-terminus (Figures 1A and B). Of the various hantavirus orthologs we tested, only constructs from the related New World hantaviruses ANDV and MAPV (accession numbers NP_604472.1 and YP_009362281, respectively) yielded enough protein for structural studies (an aa sequence alignment of Gn and Gc from selected hantaviruses is provided in Figure S1). We obtained monoclinic crystals for MAPV Gn^H/Gc diffracting to 2.2 Å resolution and rhombohedral crystals for ANDV Gn^H/Gc diffracting to 2.6 Å (Table 1) and determined the structures by molecular replacement. Both displayed Gn^H attached to the tip of Gc (Figure 1B), but showed two strikingly different overall conformations, with Gc in the ANDV Gn^H/Gc structure forming the characteristic post-fusion trimer despite having Gn^H bound. Using the same approach, we produced in parallel the C-terminal region of the Gn ectodomain (Gn^B, aa. 375–484) from ANDV. Gn^B crystallized in an orthorhombic and in a tetragonal space group at acidic and neutral pH, respectively. The structure was determined by single wavelength anomalous dispersion (SAD) and refined to 1.9 Å resolution (Table 1). Both crystals showed an identical intertwined Gn^B tetramer containing an N-terminal β-sandwich (aa 379–446) featuring an IgS fold, followed by a C-terminal α-helical hairpin (Figure 1C).

The Gn^H/Gc heterodimer features Gc in the pre-fusion conformation

Examination of the previously reported cET-STA map of the Tula hantavirus Gn/Gc *in situ* tetramer at 16 Å resolution (EMDB: EMD-3364) (Li et al., 2016), indicated that the X-ray structure of the Gn^H/Gc heterodimer could not be satisfactorily docked, whereas a good fit was obtained using the mirror image of this map. Although the hand of tomographic reconstructions is uniquely defined (unlike the reconstructions obtained by single particle cryo-EM), any incompatibility in the convention for the sign of the rotation angles in the various software used for data collection and processing – or the exchange of X and Y axes of the detector - can result in the mirror image of the actual structure. We therefore experimentally established the correct handedness by including human papillomavirus capsids – which have a chiral structure of known handedness - in the sample as reference (see Methods and Figure S2A-D), demonstrating that the hand of the previously reported map was indeed inverted. We also increased the resolution of the map by using a larger dataset of imaged particles combined with strategies for removing noise and accounting for conformational flexibility of the spike itself. In addition, because purified hantavirus particles display a broken surface lattice, we restricted the dataset to include only regular patches on the virion surface for 3D classification. This strategy therefore allowed us to use for averaging a sub-volume of the lattice containing a central spike and the contacts with its four immediate neighbors. At 11.4 Å resolution, the resulting cET map of Tula virus is representative of the hantavirus prefusion spike due to high sequence identity (see Figure S1) of the glycoproteins across hantaviruses (e.g. Tula virus Gn and Gc display 63% and 73% amino acid sequence identity with their ANDV counterparts,

respectively). Docking the 2.2 Å resolution MAPV Gn^H/Gc structure (Figure 1B, left panel) into the cET map as a rigid-body (Figures S3A and Table 2) resulted in a satisfying overall fit, although domain III was partially out of cET density. The 2.6 Å resolution structure of ANDV Gn^H/Gc (Figure S3B) did not fit the cET map, as expected since it has Gc in the post-fusion conformation.

In order to trap the ANDV heterodimer in its pre-fusion form, we inspected the structure of post-fusion Gc in order to find potential mutations interfering with trimer formation. A similar approach has been used to stabilize the pre-fusion conformation of several class I viral fusion proteins (Krarup et al., 2015; Pallesen et al., 2017; Rutten et al., 2020). We noted that the His953 side chain is involved in a tight network of polar interactions with the side chains of Asp679, Lys833 and Asn955 around the molecular 3-fold axis at the center of the Gc trimer (Figure S4A). We thus introduced the H953F mutation, as the bulkier phenylalanine side chain cannot make the same interactions. Furthermore, the H953F mutant had been shown not to interfere with budding of ANDV VLPs (Bignon et al., 2019), indicating that it should not affect the Gc pre-fusion conformation. The ANDV Gn^H/Gc H953F mutant yielded hexagonal crystals diffracting to 3.2 Å resolution (Table 1), in which Gn^H/Gc adopted an organization very close to that of wild-type MAPV Gn^H/Gc (Figure S3C). The two complexes could be aligned with a root mean square deviation (rmsd) of 5.7 Å between all pairs of equivalent Cα atoms (764 in total), although the individual domains superposed to about 0.5 Å rmsd. The structures differed essentially at the level of the hinge of Gc domain II (in the *klaefg β*-sheet, Figure S3D), at its domain I proximal end. This region has been shown to adopt variable conformations in other class II fusion proteins, for instance in flaviviruses (Zhang et al., 2004) and in phleboviruses (Halldorsson et al., 2018). When we docked the atomic model of ANDV Gn^H/Gc H953F into the cET map as a single rigid body - resulting in the statistics displayed in Table 2 - the altered conformation at the hinge brought domain III to fit well within cET density (Figure S3C, lower panel; compare with S3A).

To further validate that the interface observed in the crystals indeed corresponded to that of Gn/Gc heterodimers at the viral surface, we used the DbD2 server (Craig and Dombkowski, 2013) to identify pairs of residues interfacing each other at the Gn^H/Gc interface with favorable geometry to form inter-chain disulfide bonds when mutated to cysteine (Figure 1D, left panel). We found that mutants H294C/T734C, T99C/P774C and N94C/V776C (the numbering corresponds to the polyprotein precursor, Figure S1) did not interfere with efficient VLP formation as compared to the wild type construct. SDS-PAGE analysis (Figure 1D, right panel) showed that while Gn and Gc from wild type VLPs migrated as monomers (at ~70 and ~55 kDa, respectively) under both reducing and non-reducing conditions, Gn and Gc from VLPs formed by the double mutants migrated similarly only upon reduction. In contrast, in a non-reducing gel these mutants migrated as a single band, corresponding to a ~130 kDa form, which was recognized by anti-Gn and by anti-Gc antibodies. The

presence of this species, which is absent in wild type VLPs and which dissociates into Gn and Gc monomers by addition of a reducing agent, unambiguously indicated the presence of covalently linked heterodimers on the particles. In conclusion, the match to the cET density and the mutagenesis results strongly support the interpretation that the Gn^H/Gc complex observed in the crystals reflects interactions present on the viral particles.

An overall model of the spike and the surface glycoprotein lattice

We generated a quasi-atomic model for the reference tetrameric spike by applying 4-fold symmetry operations to the ANDV Gn^H/Gc H953F atomic model docked as rigid-body into the asymmetric unit of the cET map using Chimera (Pettersen et al., 2004). Likewise, we docked one protomer into the asymmetric unit of an adjacent spike, and used the 4-fold axis of the reference spike to generate the asymmetric unit of the remaining neighboring spikes (Figures 2A and B), which allowed the examination of spike-spike contacts. The quasi atomic model for the resulting $(Gn^H/Gc)_4$ tetramer filled most of the spike, but left an unaccounted density volume at the membrane-proximal side, where most of the four-fold contacts appear to take place (Figure S5). In the spike model, the C-terminal end of the crystallized Gn^H construct pointed into this unoccupied density (Figure 2C), strongly suggesting that it should correspond to Gn^B (Figure 1A). Indeed, the X-ray structure of the Gn^B tetramer fitted very well as a rigid body within this volume of the map (Figure S5, right panels and Table 2), confirming that Gn^B contributes the most to the intra-spike tetrameric contacts. The separate docking of the Gn^H/Gc heterodimer and the Gn^B tetramer thus yielded a complete model for the external region of the hantavirus spike (Movie S1). The location of the last residue of Gn^H (Glu374) and the first resolved residue of Gn^B (Cys379) are within the connectivity range of the intervening four residues in an extended conformation (Figure 2C). The C-terminal end of Gn^B (Gly484) points towards the viral membrane, at a location where there is density for a bundle of trans-membrane helices resolved in the 11.4 Å resolution cET map (Supplementary Figure S5C).

The model for the spike tetramer further revealed that the *N*-glycans attached to Asn402 and to Asn930 project into internal cavities of the spike (Figures 2B and G) and appear to play a stabilizing structural role, despite the mobility of the sugar chains. Mutations knocking out glycosylation at these sites were indeed shown to result in Gn/Gc spike complexes that reached the Golgi apparatus efficiently, but were not recognized by certain conformational antibodies (Shi and Elliott, 2004). These observations indicate that the integrity of the $(Gn/Gc)_4$ tetramer may be compromised by the absence of the internal glycan chains, which appear to form a scaffold underneath the spike surface. The arrangement of Gn and Gc in the spike is also in line with the observation that in HTNV the *N*-linked glycans are of the high-mannose type (Antic et al., 1992; Schmaljohn et al., 1986; Shi and Elliott, 2004), as they appear sterically protected from further modification in the Golgi. Some

hantaviruses, such as Dobrava and Thottapalayam viruses, display additional *N*-glycosylation motifs in Gn. These other sites are however predicted to be exposed at the top of the spike and would project the sugar chains to solvent at the very top, where they would be susceptible to modification to the complex form in the Golgi apparatus.

The quasi atomic model generated for the hantavirus envelope glycoprotein layer confirmed that the (Gn/Gc)$_4$ spikes are formed essentially by Gn:Gn contacts, and that the surface lattice is formed by inter-spike Gc:Gc contacts. The inferred homodimeric Gc:Gc interface is similar to that observed previously in a crystallographic dimer of HNTV Gc crystallized at neutral pH (Figure S4B-D), (Guardado-Calvo et al., 2016), with an additional contribution from the Gc N-terminal segment, which augments the inner sheet by contributing an additional β-strand (A'$_0$ strand, Figure S1). The relatively small buried surface area (550 Å^2 per protomer) suggests that a certain degree of rocking about the inter-spike contacts is allowed, consistent with the pleomorphic nature of the particles. Imposing the exact interactions observed in the crystallographic Gc dimer to the inter-spike contacts resulted in slightly reduced fitting scores in comparison to fitting the individual Gn/Gc protomers (Table 2), possibly because there are more degrees of freedom that could artificially increase the score at this resolution. Overall, our results are in line with the previously reported data showing that mutations guided by the crystallographic HTNV Gc dimer interface affect spike-spike interactions and particle assembly (Bignon et al., 2019).

The model derived previously by docking the X-ray structure of PUUV Gn^H into the 16-Å resolution Tula virus cET map (Li et al., 2016) with inverted hand was overall consistent with the new model, with Gn^H making 4-fold contacts in the spike. The reason is that at 16 Å resolution the density at the top of the individual spikes was essentially centrosymmetric, and the correction of the hand did not substantially affect the placement of Gn^H. In contrast, the chirality of the lateral contacts between spikes is quite pronounced, and there was no attempt to dock Gc in that study (Li et al., 2016). A detailed comparison showed that Gn^H is indeed oriented differently in the new spike model, which leads to different exposed surfaces and inter-protomer contacts (Figure S2, compare panels E and F). We used our new model of the hantavirus surface glycoprotein lattice to map the available data – most often obtained by Pepscan (Heiskanen et al., 1999; Koch et al., 2003) - on sites recognized by monoclonal antibodies (Mab) targeting different human pathogenic hantaviruses. Most of these data consist of peptides mapping to different regions of the glycoprotein, some of them corresponding to segments distant from each other in the primary sequence. We found that such segments are brought together in the atomic model to form a continuous patch at the particle surface, indicating the location of the epitope for each Mab. Some of the peptides belong to different protomers, but cluster at Gn:Gn and Gc:Gc contact regions at the lattice surface (Figure S2G).

Organization of the Gn^H/Gc protomer in the pre-fusion form.

As in a typical class II viral fusion protein, the three structured domains of Gc are organized around domain I (red in Figure 3), which is a β-sandwich composed of an inner ($B_0I_0H_0G_0$) and an outer ($C_0D_0E_0F_0$) β-sheet, where "inner" and "outer" refer to their orientation in the postfusion trimer (Guardado-Calvo et al., 2016). The D_0E_0 and H_0I_0 connections between adjacent domain I β-strands form two long excursions (amino acids 713–804 and 840–949) projecting from one end of the domain I β-sandwich to form the elongated domain II (yellow in Figure 3). A flexible linker connects the opposite end of the domain I β-sandwich to the immunoglobulin superfamily (IgS) domain III (aa 973–1,061, blue in Figure 3), which in turn connects to the C-terminal transmembrane (TM) anchor (aa 1,108–1,130) via an extended segment termed "stem" (cyan). Domain II is composed of a domain I-proximal "base" sub-domain made of the highly twisted *klaefg* β-sheet (labeled in Figure 1B, right panel) and a couple of short α-helices (Figure 2B), and a distal "tip" subdomain composed of the elongated *bdc* β-sheet packing against the *ij* β-hairpin (Figure 2B). The tip exposes the target membrane insertion surface (TMIS) at the distal end of the protein. Unlike the class II fusion proteins from the flaviviruses and alphaviruses, in which the TMIS is composed of a single loop (the *cd* loop, also termed the "fusion loop" and highlighted in orange in Figure 3B-C), hantavirus Gc has a tripartite TMIS composed of residues in loops *bc* (Tyr739), *cd* (Trp766) and *ij* (Phe900) (Guardado-Calvo et al., 2016; Guardado-Calvo and Rey, 2017). In the heterodimer, Gn^H makes extensive interactions with Gc domain II (Figure 3B, top half of the panel). This is the case for the three Gn^H/Gc structures presented here (Figure 1B), including that with Gc in the post-fusion conformation. The interface between Gn^H and Gc involves about 1,950–2,050 $Å^2$ of buried surface area (BSA) per polypeptide chain, of which roughly 20% are contacts with *N*-linked glycans (Figure 3C). The contact regions include conserved surface patches of both glycoproteins (Figure S3E). The Gc prefusion conformation is also stabilized by intra-chain interactions between domain III and the domain I outer sheet, which is augmented by an additional β-strand (K_0, aa 1,077–1,082) formed by residues belonging to the stem (Figure 3B, cyan), and a Gc N-terminal segment (the "N-tail", aa 652–668, highlighted in light brown in Figure 1B), which is an extension with respect to other viral class II fusion proteins. Three conserved aromatic side chains, (W660 and H665 in the N-tail, and Y1080 in the stem, labeled in Figure 3B) form a platform interfacing with domain III. The N-tail makes an arch that surrounds domain I, contributing an additional short β-strand (A'_0) to the domain I inner sheet. The contribution of the stem and the N-tail to the pre-fusion conformation of domain I are features that have not been observed in other viral class II fusion proteins. The previously reported structure of monomeric HTNV Gc at neutral pH, interpreted as displaying a putative "pre-fusion" conformation, had been obtained after controlled proteolysis that cleaved off the stem, and therefore lacked the K_0 strand (Guardado-Calvo et al., 2016). The N-tail was in turn disordered upstream strand A_0 (aa 669–672). The absence of these

contact sites gave rise to a displaced domain III that resulted in a rod-like Gc molecule in which domains II and III lie at the opposite ends of domain I (Figure 3D). This conformation would be compatible with an extended intermediate form of Gc adopted early during the membrane fusion reaction, as observed in other bunyaviruses (Halldorsson et al., 2018).

The Gn^H conformation in the complex is essentially the same as that observed in the structures of isolated Gn^H from PUUV and HTNV reported previously (Li et al., 2016; Rissanen et al., 2017), except for three loops, disordered in those structures, which are involved in interactions with Gc in the heterodimer. Many features of the Gn/Gc complex are reminiscent of the alphavirus p62/E1 glycoprotein heterodimer (Voss et al., 2010). Both p62 and Gn present three IgS domains (termed domains A, B, and C) disposed around a central β-ribbon structure (Figure S6). The β-ribbon consists of a long, twisted β-hairpin encircled by several "arches", which in Gn are more elaborate than in p62, displaying various insertions with additional secondary structure elements. Despite the overall similarity, Gn domains A and B are disposed differently with respect to each other compared to p62 and have developed different insertions into the core domains to interact within the heterodimer. These differences notwithstanding, in both cases they engage domain II of their partner fusion protein by interacting with the *b* strand and with the *de* loop, and both feature an IgS domain C immediately downstream the *β*-ribbon. Furthermore, the alphavirus icosahedral surface glycoprotein lattice is maintained by intra-spike contacts made exclusively by p62, whereas the inter-spike 2-fold contacts are mediated by E1 only, just as in the hantavirus Gn/Gc square surface lattice. These similarities indicate that not only E1 and Gc are homologs but also that Gn is evolutionary related to p62 despite having diverged further in structure.

One of the three main Gn regions contacting Gc is the "α1 loop", centered on a 3/10 helical turn (α1 helix, aa 289–291) in the MAPV structure. This loop encompasses a polypeptide segment (aa 277–299) featuring the highly conserved stretch 281-GEDHD-285 across the *Orthohantavirus* genus (Figure S1). The C-terminal end of the α1 loop forms β-strand A_B (Figure S3F-H) running parallel to β-strand *b* of Gc domain II. A second contact site with Gc involves the Gn arch surrounding the β-ribbon (arch1, aa 202–208, labeled in Figure 3B), which interacts with the *fg* and *kl* loops in the hinge region of Gc domain II (Figure 3B). The third set of important interactions is made by the tip of a long β-hairpin (i1-i2 hairpin, Figure 3B, top; Figure 3C), which stands out as an insertion into Gn domain A (Figure S6). The i1-i2 loop at the tip of this β-hairpin caps the *cd* and *bc* loops of Gc, and we accordingly termed it "capping loop" (aa 88–99). This loop adopts several conformations in the absence of Gc, as shown by the structures of isolated Gn^H crystallized at different pH, none of which corresponds to the conformation seen here in the complex with Gc (Figure S3I). A striking additional inter-chain interaction is mediated by the two Gn conserved glycan chains attached at positions N138 and N350, which embrace the tip of domain II (Figure 3C), respectively interacting with the side

chains of Phe900 and Tyr745, both residues forming part of the Gc TMIS. Removal of either of these glycosylation sites in HTNV has been shown to give rise to defects in the intracellular transport of the glycoprotein complex (Shi and Elliott, 2004).

The intra-spike interfaces

The quasi atomic model for the spike shows that the Gn^H:Gn^H contacts at the membrane distal side of the spike are limited to 200 $Å^2$ BSA per protomer, which are mediated by the conserved GFC β-sheet of domain A and the arch-1 region from the clockwise protomer seen from the top (Figures 4A and 4C). In contrast, the Gn^B moiety, at the membrane-proximal side, makes most of the Gn:Gn interactions stabilizing the tetrameric spike (Figures 4B, D, and E). Gn^B is formed by domain C (aa 379–446), a seven-stranded IgS β-sandwich followed by a viral membrane proximal external region (MPER) (Figure 4B) composed of two amphipathic α-helices that form a disulfide-linked α-helical hairpin. The Gn MPER wraps around domain C from an adjacent protomer (Figure 4B) to make an intertwined, compact Gn^B tetramer with 1,000 A^2 of BSA per protomer, distributed equally between domain C and the MPER. Accordingly, a construct containing only domain C, i.e. lacking the Gn MPER, behaved as monomeric in solution (data not shown). Most of the residues that shape the tetrameric interface are conserved across hantaviruses (Figure S1). The base of the tetramer exposes a hydrophobic surface composed of 28 conserved hydrophobic residues (seven per protomer, Figure 4B), facing the viral membrane.

The Gn^H/Gc crystals displayed clear density for the Gc stem except for its N-terminal segment (aa 1,063–1,076) between domain III and the K_0 strand (Figure 3B). It is possible that this segment of the stem requires Gn^B to become structured. We note that the crystals of the Gn^B tetramer included a ligand – which was present in both crystal forms and which we interpreted as a short RNA oligonucleotide – running within a polar groove between two Gn^B protomers. This ligand - the density for which is displayed as a purple surface in Fig. 1C, top panel - copurified with the protein and is not likely to be biologically relevant. Yet it revealed the existence of a reactive, positively charged groove along the sides of the Gn^B tetramer (Figures 1C and 4D), running radially and well placed to accommodate the Glu1066 side chain in the polar N-terminal segment of the Gc stem in the context of the spike, further strengthening the interactions within the tetrameric spike. In the alphavirus p62/E1 complex, the corresponding region of the E1 stem was also shown to be structured in interaction with p62 domain C (Voss et al., 2010).

The C-terminal part of the stem (aa. 1,092–1,107, between the K_0 strand and the TM segment, Figure S1), is also disordered in the Gn^H/Gc crystals. The Psipred server (Buchan and Jones, 2019) predicts this Gc MPER as containing an amphipathic α-helix. This segment of 15 amino acid can easily span the 25-Å distance intervening between the last ordered stem residue in the Gc model and

the cET density for the Gc TM segment (Figure 4A, see also the supplementary Movie). The Gc MPER would lie parallel to - and partially embedded in - the viral membrane outer leaflet. At 11 Å resolution it is not resolved in the cET map. Similar MPERs have been described for other class II fusion proteins just upstream the TM segment, for instance the envelope protein of dengue virus (Zhang et al., 2013), but also for the class I envelope trimer of HIV (Fu et al., 2018) and the class III fusion protein gB of herpes simplex virus (Cooper et al., 2018). We note that mutations in this region, such as S1094L, which would increase the Gc MPER amphipathic character, were reported to increase the infectivity of recombinant vesicular stomatitis virus particles bearing HNTV Gn/Gc glycoproteins (Slough et al., 2019), suggesting that the interaction of the Gc MPER with membranes may affect viral infectivity. Together, the Gn and Gc MPERs appear to form a platform underneath the spike, interfacing with the viral membrane (Figures 4A and E). The compound effect of embedding all or most of these amphipathic helices in the outer leaflet of the lipid bilayer would be expected to significantly affect the curvature of the viral membrane (Martyna et al., 2017).

The fusogenic conformational change of Gc

In order to better understand the structural reorganization that takes place to drive membrane fusion for entry, we also made constructs of the isolated MAPV and ANDV Gc ectodomains, and obtained crystals diffracting to 2.7 and 2.4 Å resolution, respectively (Table 1). The resulting structures revealed Gc in its post-fusion conformation (Figure 5A, central and right panel) even when crystallized at neutral pH, as was the case with the HTNV and PUUV post-fusion Gc trimers reported earlier (Guardado-Calvo et al., 2016; Willensky et al., 2016). Comparison of the pre- and post-fusion conformations of Gc confirmed that the most conspicuous changes are the drastic relocation of domain III and the stem, as well as an altered topology of the β-strands composing the outer and inner sheets of domain I (Figures S7A and E). Similar changes had been reported for the class II fusion proteins from this and other virus families in the pre- and post-fusion forms of domain I (Bressanelli et al., 2004; Gibbons et al., 2004; Halldorsson et al., 2016; Modis et al., 2004).

Unlike the other class II fusion proteins studied until now, the domain II tip in the pre-fusion form adopts an alternative conformation, stabilized by Gn, in which the TMIS is not formed (Figure 5B). The side chains of three key residues of the TMIS - Trp766, Tyr745 and Phe900 – which were shown previously to be crucial for Gc binding to target membranes (Guardado-Calvo et al., 2016), are not exposed. The observed reorganization of the tip mainly affects the *cd* loop, which in the post-fusion form makes an α-helical turn exposing the side chain of Trp766 and burying that of Asn769. By contrast, in the pre-fusion form, the Trp766 side chain is buried and that of Asn769 is exposed to solvent (Figures 5B and S7D). This rearrangement results in a drop of the surface hydrophobicity at this critical membrane-interacting region. The remaining non-polar side chains are covered by the

capping loop of Gn, which cloaks part of the *cd* loop and the *bc* loop, and the N138 glycan, which stacks against Phe900 and covers the *ij* loop.

What controls the observed reorganization of the tip is unknown, but the unusual carboxylate/carboxylic-acid hydrogen bond between the side chains of the strictly conserved Glu757 and Asp759 (Guardado-Calvo et al., 2016) is likely to play a role. We observed that this acid hydrogen bond is required for structuring the tip of domain II in the post-fusion conformation. The presence of Gn reshapes the *bc* loop such that the neighboring Lys744 makes a hydrogen bond / salt bridge with Asp759, preventing formation of the acidic hydrogen bond and therefore precluding the *cd* loop from exposing Trp766 (Figure S7F). How the changes in the *bc* loop are transmitted to the *cd* and *ij* loops remains an open question.

DISCUSSION

Our study revealed the organization of the hantavirus surface glycoprotein lattice by describing the X-ray structures of ANDV and MAPV Gn^H/Gc heterodimers featuring Gc in the prefusion conformation, the conserved tetrameric Gn base of ANDV, and their rigid-body fitting into an 11.4 Å resolution cET map of Tula hantavirus. Given the high amino acid sequence similarity between the envelope glycoproteins, the model presented here is expected to be valid for all hantaviruses. The Gn^H/Gc structures also revealed the role played by the N-terminal tail and stem of Gc in stabilizing the interaction between domains I and III in the metastable prefusion form. In addition, the effect of Gn in the conformation of the Gc tip revealed the way Gn protects from premature TMIS formation. These features have not been previously observed in other class II fusion proteins. Because the domain II tip displays clear amino acid sequence conservation with its counterpart in nairoviruses and in orthobunyaviruses, the observed protection mechanism can potentially be extrapolated to the fusion machinery of those more distant human pathogenic bunyaviruses.

The evolutionary history of the class II fusion proteins remains enigmatic. They have been identified in positive- and in negative-sense ssRNA viruses as well as in multiple and varied eukaryotic organisms where they drive the fusion of gametes and of somatic cells (Fedry et al., 2017; Perez-Vargas et al., 2014). Their complex yet highly conserved 3D fold is strong evidence for common ancestry despite the absence of any remnants of sequence conservation (Modis, 2014). We show here that the similarity in the viral proteins extends beyond the actual fusion effector protein to include the accompanying protein, as in the case of the alphaviruses and hantaviruses. Indeed, despite major differences, we found that the overall organization of the hantavirus spike showed striking parallels with that of the alphavirus spike. These include the roles of domains A and C of Gn,

which like their counterparts in alphavirus p62 make the intra-spike contacts while Gc, like its alphavirus E1 counterpart, provides lateral contacts between spikes. The flavivirus example also shows that in some cases the accompanying protein has been substituted during evolution, as seem to be the case with the flavivirus prM/E heterodimer (Li et al., 2008). Yet even in this case, the accompanying protein prM binds to the domain II tip by interacting with β-strand *b*, in an apparently universal fashion.

The quasi-atomic model of the spike presented here provides a framework to understand an accumulating body of data on hantavirus biology. The conserved *N*-linked glycans, which were observed to be buried in the spike and which remain high-mannose in released viral particles, are a prime exemplar. Our structures support the hypothesis that the tetrameric spikes form early in the ER, before they are transported to the Golgi apparatus. The sugar chains are then not accessible to the glycosyl transferases that modify the glycan chains, buried within the tetramer. As the sugar chains are buried, they are not used as a shield to evade the immune system as in other viruses, nor as cellular attachment factors as in alphaviruses, flaviviruses or phleboviruses (Watanabe et al., 2019). Rather, the glycans appear to play an intrinsic structural role by forming an internal scaffold.

Previous studies have reported a dynamic behavior of the hantavirus spikes, which exhibit a temperature-dependent equilibrium between two conformational states that co-exist at physiological temperatures (Bignon et al., 2019) - a process termed "breathing". The closed form, which does not bind membranes at neutral pH but which is functional in acid-pH-induced membrane fusion, and the open form, which binds membranes even at neutral pH but is unable to drive membrane fusion upon exposure to acid pH. The structure of the Gn^H/Gc heterodimer presented in this manuscript appears to reflect the closed form, since the capping loop partially shields the TMIS and the Trp766 side chain is buried. The open form probably results from a partial dissociation of the heterodimer, leading to a change in the conformation of the *cd* loop in Gc and its interaction with membranes. It has been hypothesized that spike breathing may provide advantages to establish persistent infections in rodents by exposing decoy epitopes that elicit a poorly neutralizing response (Bignon et al., 2019). In line with this hypothesis, neutralizing antibodies isolated from mice (Xu et al., 2002) and bank voles (Lundkvist and Niklasson, 1992), which were respectively immunized with inactivated HNTV and PUUV infected tissues, recognize epitopes occluded in the closed form (Figure S2E-G) but which can be transiently exposed as the spike breaths.

The atomic model of the hantavirus spike now paves the way for the development of novel immunogens. For a certain category of enveloped viruses, designing subunit vaccines using the recombinant fusion glycoprotein ectodomain for immunization has been complicated by the spontaneous and irreversible switch to the post-fusion form, which does not expose the relevant epitopes targeted by neutralizing antibodies. One example is the respiratory syncytial virus (RSV)

fusion glycoprotein, which required mutagenesis to stabilize its pre-fusion conformation (Krarup et al., 2015) and expose the right epitopes to elicit strongly neutralizing antibodies. We found a similar situation with the ANDV Gn^H/Gc complex, and we describe one way of interfering with the adoption of the post-fusion form by introducing the H953F mutation. We have furthermore validated our model for the spike by designing disulfide bridges crosslinking the Gn/Gc protomer, providing a proof-of-principle that such additional stabilization is feasible. Our results make it now possible to further investigate inter-protomer disulfide bonds to stabilize the overall spike tetramer as a potential next-generation immunogen. As spike "breathing" has been demonstrated in Andes virus and other hantaviruses that cause severe disease in humans, the envelope glycoproteins engineered as proposed here should provide superior immunogens for eliciting highly potent neutralizing antibodies targeting the closed form of the spike.

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AUTOR CONTRIBUTIONS

N.D.T., J.T.H., F.A.R., and P.G.C. designed the experiments. AS expressed, purified, and crystallized the proteins. A.S. and P.G.C. processed the crystallographic data. E.A.B., N.A.M. and N.D.T. prepared and analyzed the VLPs. R.S., S.L., and J.T.H. prepared samples for cryo-ET, collected and processed

the data. J.C.M., R.K.J., K.C., N.D.T., J.T.H analyzed the data; P.G.C and F.A.R. prepared the manuscript with input from all authors.

DECLARATION OF INTERESTS

E.A.B., N.D.T., F.A.R. and P.G.C. are coinventors in a pending patent application for the development of stabilized immunogens for an Andes virus vaccine. K.C. is a member of the scientific advisory board of *Integrum Scientific, LLC*, and F.A.R is in the Board of *Virtexx, SAS*. The other authors declare no competing financial interest.

MAIN FIGURES TITLES AND LEGENDS

Figure 1. X-ray structures of the hantavirus glycoproteins. A) Domain organization of the hantavirus M segment. The upper panel shows linear diagram colored according to protein domains, with the signalase cleavage site (Gc N-terminus) marked by an arrow. Glycosylated asparagines are labeled in green above the ORF.. The lower panel shows a schematic representation of the Gn/Gc heterodimer with the viral membrane as a dashed box and TM segments indicated. The ectodomains of Gn and Gc are colored as in (A) with *N*-linked glycans indicated with "Y" symbols. Dashed ovals outline the two separate fragments whose structures were determined here. **B)** Structures of the hantavirus Gn^H/Gc heterodimer. First row: diagram of the single-chain Gn^H/Gc construct used for crystallization upon thrombin treatment (engineered thrombin sites are in cyan). The second and third rows show two orthogonal views of MAPV Gn^H/Gc (left), ANDV (Gn^H/Gc)$_3$, (middle) and the ANDV Gn^H/Gc-H953F mutant (right). In the middle panel, the (Gn^H/Gc)$_3$ trimer is shown with Gc in yellow, grey and light-cyan surfaces, and Gn as red ribbons. In the side view (second row), the front Gn chain was omitted for clarity and the matching Gc chain is shown as yellow ribbons, as in the left and right panels. All three panels in this row display Gc domain I in the same orientation. The top view (third row) also shows the glycan chains as green sticks and labeled. **C)** Structure of the ANDV Gn^B tetramer in side view (top) and bottom view (lower panel), with one Gn^B chain highlighted in orange. The glycan attached to N402 in Gn^B is indicated as green sticks. The density corresponding to an omit map contoured at 1.5σ is shown in blue, marking a reacting groove which binds a ligand from the supernatant (see Methods). We postulate that in the spike the Gc stem may run within this groove. **D)** Engineering of inter-chain disulfide bonds. Left panel: closeup of the Gn^H/Gc interface marking the location of pairs of residues mutated to cysteine (connected by green bars and labeled). The right panel shows an SDS-PAGE analysis of VLPs released into the supernatant of cells expressing wild type ANDV GnGc and the indicated Cys double mutants, under reducing and non-reducing conditions (as indicated).

Figure 2. Organization of hantavirus Gn/Gc spikes on virions. A) The 11.4 Å resolution cET-STA map of the Tula virus spike plus the asymmetric unit its four neighbors viewed from the top, along the 4-fold axis and with the docked x-ray structures of the ANDV Gn^H/Gc-H953F mutant and of. One protomer of the spike is colored as in Fig. 1. A protomer of the adjacent spike, related by a local 2-fold axis, is highlighted in salmon/blue (respectively for Gn^H and Gc). B) Orthogonal view of the spike with the 4-fold axis in the plane of the Figure. The conserved *N*-linked are shown as green sticks and labeled. OL and IL indicate the outer and inner leaflets of the viral membrane, respectively. C) Closeup of the spike showing the Gn^B tetramer (for clarity, the Gc moiety in the front was removed). The N- and C-termini of Gn^H and Gn^B are indicated and labeled in blue and black, respectively. D) Closeups of selected regions corresponding to the dashed boxes in panels A to C to show the quality of the fit, color-framed to match the corresponding boxed areas (in dashed lines) in the panels just above (see also the supplementary Movie). E) Reconstruction of a Tula virus particle generated by back-projecting the 11.4 Å cET/STA density of the spike to the original spike locations on a particle. The resulting surface protein lattice is shown in cyan with the viral membrane in gray. F-G) The spike in a top view (F) with one of the four protomers colored as in Figure 1 and the sugar residues in green, and in side view (G). As in C), the Gc subunit at the forefront (marked by a dashed oval in panel F) was omitted in (G) to visualize the Gn^B tetramer at the spike center. The Gn^H N- and C-termini are marked as in (C) (only the N-termini in (G), for clarity).

Figure 3. The hantavirus Gn/Gc prefusion heterodimer. A) The single open reading frame of the hantavirus genomic RNA segment M colored by domains according to the key to the right. N-glycosylation sites are shown as "Y" symbols (ANDV numbering, Figure S1). B) The 2.2 Å resolution X-ray structure of the MAPV Gn^H/Gc heterodimer colored as in A, with the disulfide bonds in green and the *N*-glycans displayed as sticks colored according to atom type C) Orthogonal view of the Gn^H/Gc interface seen from domain II tip. D) Ribbon representation of HNTV Gc monomer (PDB:5LJY) centered on domain I, showing the extended conformation adopted by Gc in the absence of the stem. Domain I is in the same orientation in panels B and D. The Gc N-tail in the left panel is highlighted in light brown and displayed as thicker tubes, and the stem in cyan.

Figure 4. Interchain contacts stabilizing the hantavirus spike. A) Side view of the spike, omitting the front Gc subunit for clarity. The four protomers are colored as in Figure 3, with Gn^B in orange. The Gc-MPER, not resolved in the structure, was modeled as an α-helix (colored cyan) to indicate its putative location in the lipid-head region of the membrane's outer layer (schematically indicated by a thick black line). B) X-ray structure of the Gn^B. Disulfide bonds are indicated in the front protomer as

green sticks and numbered. The side chains of non-polar residues of the MPER exposed to the viral membrane are shown as sticks and labelled in the left whereas the right panel shows the interactions between two protomers with interfacial residues highlighted. C, D, and E) Slices of the spike model normal to the 4-fold axis, corresponding to the regions indicated in the vertical bar at the left of panel A. A dashed outline follows the contour of one spike protomer along the three radial sections displayed. C) Contacts in the membrane distal region (the spike's "crown"). The Asn350 glycan is shown as blue sticks. D) The spike's "mid" region, displaying two glycans attached to Asn402 and Asn930, which fill the internal cavities underneath the spike crown. The segment of the Gc stem that is disordered in the X-ray structure (dotted line in Figure 3B) projects into a cET density volume in close proximity with the reactive groove (red arrowheads) at the Gn^B tetramer interface (indicated in blue in Figure 1C. E) The membrane proximal region. The two amphipathic helices of the Gn MPER (at the Gn^B C-terminus) and the predicted Gc MPER, which precede the TM segments, form a supporting platform embedded in the outer leaflet of the viral membrane. The approximate location of the three TM segments per protomer are indicated with full blue circles. The directionality of the helices is indicated with arrows. A thin dashed rectangle surrounding the Gc MPER helix was added to indicate that this segment was not resolved in the X-ray structure.

Figure 5. Conformational change of the Gc domain II tip for TMIS formation. A) The post-fusion conformation. The left and central panels show a top (along the 3-fold axis) and a side view of the ANDV (Gn/Gc)$_3$ trimer with Gc in the characteristic post-fusion hairpin conformation displayed in surface representation and with Gn shown as green ribbons. One Gc subunit is colored by domains, with Tyr739, Trp766, and Phe900 of the TMIS displayed and labeled. For clarity, the Gn polypeptide chain in the front was omitted in the side view. N-linked glycans are shown as thick sticks. The right panel shows the structure of ANDV Gc post-fusion trimer crystallized at pH 7.5 and depicted as in the central panel. The horizontal bar indicates the target membrane into which the TMIS would insert. B) The tip of MAPV Gc domain II showing the organization of the *bc*, *cd*, and *ij* loops in the pre- (top panel) and post- (bottom panel) fusion conformations, from the 2.2 and 2.4 Å resolution structures, respectively (Table 1). Left and right panels are two orthogonal views, as indicated. Side chains important for target membrane insertion and for forming the polar network around Glu757-Asp759 are highlighted. In the prefusion conformation, the *bc* and *ij* loops are respectively buried by the Gn N138 glycan and the capping loop, as indicated.

SUPPLEMENTAL FIGURE TITLES AND LEGENDS

Figure Supplementary 1. Amino acid sequence analysis of the hantavirus glycoprotein precursor polyprotein. Related to Figure 1**.** A) Hydrophobicity diagram and amino acid sequence conservation plot along the open reading frame of the M segment coding for the polyprotein precursor, with Gn and Gc colored by domain as depicted in the bar above the sequence alignment in panels B and C, and TM segments in black (including the signal sequence). The upper panel plots the posterior probability for each residue of being part of a TM region, as calculated by Phobius (Kall et al., 2004). The second row plots the sequence conservation scores per amino acid (blue line) calculated from the sequences of 64 hantavirus M segments (listed in Methods) with Consurf (Ashkenazy et al., 2016). The score for each aa was normalized so that the average for all residues is zero. Negative values represent positions more conserved than average. The red line plots the scores averaged in a sliding window of 10 residues. As expected, the sequence of Gn^H, which is the most exposed region of Gn, displays mainly positive scores, i.e. lower conservation than average, whereas for Gn^B and most of Gc it is the opposite. The right panel is a key to decode the contacts made by residues indicated below the amino acid sequence alignment in B and C.

B and C) Multiple sequence alignment of Gn (panel B) and Gc (panel C) from seven representative hantaviruses. Secondary structure elements are displayed and labeled above the sequences on a background colored according to domains when experimentally determined (Gn^H: domain A green, domain B blue, β-ribbon magenta, insertions in white; Gn^B in orange. Gc: N-tail light brown, domain I red, domain II yellow, domain III dark blue, stem cyan), or in a gray background when the corresponding segement was not resolved in the structure. Instead, the secondary structures predicted by Psipred (Buchan and Jones, 2019) are indicated. Strictly conserved residues are highlighted in a red background, and asparagines within *N*-linked glycosylation motifs on a green background. Disulfide bonds are indicated with a green number under the paired cysteines in the alignment. For MAPV and ANDV, the sequence of segments not resolved in the crystals are framed in green (375-378 in Gn and two segments between aa 1063 and 1108 in Gc). The *Hantaviridae* family is divided in four subfamilies, with the *Orthohantavirus* genus being one of four genera in the *Mammantavirinae* subfamily. The displayed alignment includes four members (out of 35 species) of the *Orthohantavirus* genus: two New-World hantaviruses: Andes virus (ANDV, NP_604472.1) and Maporal virus (MAPV, YP_009362281) and two Old-World hantaviruses: Hantaan virus (HNTV, CAA68456.1) and Puumala virus (PUUV, CAB43026.1). The three remaining sequences belong one member each of the three other genera in the subfamily: Laibin virus (LIBV, A0A0D5W3U2) (*Mobatvirus* genus); Longquan virus (LQNV, AGI62344.1) (*Loanvirus* genus), and Thottapalayam virus (TPMV, A0A075IFP0) (*Thottimvirus* genus).

Figure supplementary 2. Tula virus map: handedness confirmation and antigenic sites. Related to Figure 2. A) Slice of a tomogram recorded on a cryo-EM grid prepared with TULV particles (black arrow) mixed with HPV16 pseudovirus particles (white arrow) (scale bar = 100nm). B-D) STA performed using the initial reference displayed on the left (in grey) to produce the reconstructions displayed in blue on the right. The isosurfaces are displayed as a full top view (left) and a magnified cutaway side view (right). The hand is represented by the stripes (tilted red bars) below. B) Reference = TULV particle (EMD-3364). C) Reference = centrosymmetric object resulting from averaging the TULV particle (EMD-3364) with its mirror image. D) Reference = mirror image of EMD-3364. Using the same procedures in which the HPV16 particles refine to give a cryo-ET map with the correct T=7 dextro hand (Lee et al., 2015) when starting from a centrosymmetric reference, the results of the TULV refinements match the reference displayed in D (the reversed EMDB: EMD-3364 map) regardless of the handedness of the reference used. This result confirms that the true handedness of the TULV particle is flipped relative to that reported in EMD-3364, consistent with finding that our Gc/Gn^H heterodimer X-ray structure didn't fit that map, but did fit its mirror image. E-G) The antigenic surfaces of Gn and Gc. E) Top view of the quasi-atomic model for the PUUV spike (PDB: 5FYN) obtained by docking the X-ray structure of PUUV Gn^H (PDB: 5FXU) into the 16 Å resolution cET map of Tula virus (EMDB: EMD-3364) reported previously (Li et al., 2016), which had the inverted hand. The Gn molecules are in surface representation (top panel) with the two discontinuous peptides identified as part of the epitope of the PUUV neutralizing mAb 5A2 (Heiskanen et al., 1999) (peptides A and B, colored green and red, as indicated). The middle and bottom panels are the corresponding ribbon representations shown in two orthogonal orientations and ramp-colored from N- to C-termini as indicated in the colored bar underneath. The *N*-linked glycans are represented as sticks and the capping loop labelled. F) The same representation of panel E but using the spike model generated in this work. The Gc molecules are not shown for clarity. G) The antigenic surface of Gc is shown in the context of the spike. As in F, top and side views of the spike are shown, with Gn depicted as in the left panels and Gc shown as a white (central spike) or yellow surface (adjacent spikes), with the epitopes for neutralizing Mabs 3D8, 3G1, 1C9, and Y5 colored as indicated (Heiskanen et al., 1999; Koch et al., 2003; Xu et al., 2002). Note the orange and pale blue epitopes spanning across spikes.

Figure supplementary 3. Various Gc conformations in the Gn^H/Gc heterodimer. Related to Figure 2. A-C) Comparison of the X-ray structures of MAPV-Gn^H/Gc (A), ANDV-Gn^H/Gc (B), and ANDV-Gn^H/Gc-H953F mutant (C) from this work. The three structures were aligned on Gn and colored as in Figure 3. The domains are labeled and the main axes of inertia of Gc domains I and II are drawn with the angle between them quoted, highlighting the flexion about the hinge at the *klfeg β*-sheet of domain II. The

lower panels in A and C show the rigid body docking of the atomic structures of MAPV-Gn^H/Gc and ANDV- Gn^H/Gc-H953F, respectively, into the cET/STA map of the Tula virus spike. The different flexion at the hinge results in domain III partially out of cET density in (A). The conformation of ANDV-Gn^H/Gc, shown in (B), does not fit the map. D) Alignment on Gc domain I from the various structures (colored-coded as indicated) to compare the different flexion about the domain II hinge. The conformation of ANDV-GnGc-H953F, which fits best the cET map (Table 2), is also very similar to that observed in the previously reported structure of HNTV-Gc/scFv-A5 (PDB: 5LJY), which formed a crystallographic Gc dimer recapitulating the contacts between spikes on the viral particle (Bignon et al., 2019; Guardado-Calvo et al., 2016). E) Sequence conservation at the Gn^H and Gc surfaces shown as an "open book" of the MAPV Gn^H/Gc interface. The conservation score, obtained from an alignment of 65 sequences (listed in Methods) using ESPript (Robert and Gouet, 2014) mapped at the surface of ANDV Gn^H and Gc using a color ramp from white (conserved) to dark purple (variable). The interface is outlined in yellow and the conserved residues are labeled. F to H) Comparison of the conformation of the η1 loop in the ANDV-Gn^H/Gc (G), ANDV-Gn^H/Gc-H953F (H), and MAPV-Gn^H/Gc (I) heterodimers. Gn^H is shown as ribbons, Gc in surface representation and the glycan linked to Asn138 in sticks. Both subunits are colored according to domains as in Figure 3. The conserved Tyr325 and the ^{283}EDH^{285} motif are represented as sticks and labelled. I) Closeup showing the conformational variability of the capping loop in the various structures available for isolated Gn^H compared with Gn^H in the heterodimer.

Figure supplementary 4. Gc:Gc interfaces. Related to Figure 2. A) Two orthogonal views of the ANDV Gc post-fusion trimer showing a network of polar interactions about the 3-fold molecular axis at the level of domain I, involving the side chain of H953. Other residues involved in the polar network are also labelled. The H953F mutation breaks the ring of polar interactions. B-D) Comparison of the Gc:Gc interfaces observed in the X-ray structure of HNTV-Gc in an extended conformation. B) The HTNV Gc crystallographic dimer and C) the ANDV Gn^H/Gc aligned on domain I of the HTNV Gc crystallographic dimer, to show the predicted contacts and highlighting the different conformation of domain III in the extended structure. The curved blue arrows show the movement of domain III from its conformation in the Gc monomer to reach that in the Gc/Gn^H heterodimer. D) The outcome of individually docking the ANDV-Gn^H/Gc-H953F structures into two adjacent, 2-fold related spikes in the cET map. The resulting 2-fold interactions are similar but not identical, reflecting in part the resolution of the cET map, but also a certain variability of the spike-spike contacts on the pleomorphic particle. Two orthogonal views are shown, across (left panel) and down (right panel) the 2-fold axis.

Figure supplementary 5. The unaccounted density left by the Gc/Gn^H heterodimer at the center of the spike. Related to Figure 2. A) Two orthogonal views of the cET map of the hantavirus spike with the ANDV Gn^H/Gc-H953F protomer docked. The unaccounted density is labelled. B) The Gn^B tetramer nicely fills the space left vacant by Gn^H/Gc. The four-fold axis of the spike is indicated. C) View of the TM density within the region boxed in red in panel B, looking down the empty red arrow. The Gn and Gc MPER are superposed underneath this density (in the orientation of the red arrow in panel B) to show that the C-terminal end of Gn, as well as the C-terminal end of the modelled Gc MPER (both labeled with "C"), point to the TM density, as expected. See also the supplementary Movie for the overall fit of the X-ray structures into the cET density of the spike.

Figure supplementary 6. Topological comparison of the class II "accompanying" protein. Related to Figure 3. Secondary structure topology diagrams of hantavirus Gn (upper panel), alphavirus p62 (middle panel, based on 3N40 (Voss et al., 2010)), and phlebovirus Gn (lower panel, based on PDB:5Y0W) (Wu et al., 2017) colored according to domains: domain A (green), domain B (blue), domain C (gray), and the β-ribbon connector (magenta). The regions contacting E1 (alphavirus) and Gc (hantavirus) are indicated and labelled. The diagrams show that the topological arrangement in hantavirus Gn and alphavirus p62 is the same, in spite of the presence of various insertions (shown in white). In alphaviruses, P62 is the precursor to mature E3 (N-terminal part, labeled) and E2, which are processed by furin in the TGN upon transport of the spike to the cell surface. The E3 extension (light blue) is absent in bunyaviruses, which do not undergo a maturation cleavage of the accompanying protein. The lower panel indicates that phlebovirus Gc is more distant, but still has the same domain arrangement about a central β-ribbon, even when the topological connectivity within these domains has evolved further. Domain C was not resolved in the available studies of phlebovirus Gc

Figure supplementary 7. The membrane fusogenic reorganization of Gc. Related to Figure 5. A) MAPV-Gc in pre-fusion conformation, crystallized at pH 8.5 as a heterodimer with Gn^H (left panel) shown next to HNTV-Gc in an extended conformation, crystallized at pH 7.5 in complex with a human antibody fragment (not shown) (middle panel), and MAPV-Gc in post-fusion conformation, crystallized at pH 4.6 (right panel). Only one protomer of the trimeric post-fusion trimer is shown, for clarity. The three panels show Gc as ribbons in the same orientation upon alignment on domain I, ramp-colored from the N- to C-terminus as indicated. A putative target membrane is schematically represented by a horizontal line. Gn^H is displayed as a gray surface (left panel). B) Postfusion forms of Andes virus Gc in absence (orange) and presence (magenta) of Gn are superposed on domain I. Both structures are virtually identical except at the domain II tip. C) Closeup of the *cd* loop showing that

Trp766, which is a key residue required for the interaction with target membranes, is exposed to solvent in the absence of Gn (upper panel) but is buried in the hydrophobic core of the tip β-sandwich in its presence (lower panel). D) Zoom on the domain I reorganization during the fusogenic conformational change. Ribbon representation ramp-colored as in panel A, showing the reorganization of the secondary structure elements of domain I. Residues stabilizing the different conformations, and which drastically change environment during the transition, are shown as sticks. The left panel shows domain-I in prefusion conformation, with β-strand A'_0 running parallel to B_0 in the inner ($G_0H_0I_0B_0$) sheet (augmenting the Gc:Gc homo-dimeric interface at the inter-spike contact); β-strand K_0 from the stem interacting with C_0 to in the outer ($F_0E_0D_0C_0$) sheet, and a small α-helix formed by residues of the linker between domains I and III, plugging the distal end of the domain I β-sandwich. The middle panel shows domain I in the intermediate, extended conformation of Gc, showing that the absence of the stem does not affect its conformation but drastically alters the location of domain-III. The right panel shows domain I in postfusion conformation, with β-strand A_0 forming part of an antiparallel β-sheet with I_0 and B_0 from a neighboring protomer. The residues that compose β-strand K_0 move with the rest of the stem away from domain I and towards the tip of domain II, as shown in panel A. E) Closeup of the domain II tip, comparing the structures of MAPV Gc in presence (prefusion, top-left panel) and absence (postfusion, top-right panel) of Gn^H, and HNTV in presence (extended, bottom-left panel), and absence (postfusion, bottom-right panel) of the A5 antibody targeting the *bc* loop (PDB:5LJY). In addition to the changes in the *cd* loop, both Gn and A5 capture conformations of the *bc* loop that differ from those observed in the post-fusion form, illustrating the high malleability of this loop. It is noteworthy that the reorganization induced by Gn, but not the one induced by A5, results in the interaction between conserved Lys744 and Asp759. In the post-fusion forms, Lys744 points outward, allowing Glu757 and Asp759 to form an acidic hydrogen bond that stabilizes the domain II tip in the post-fusion form, as described earlier (Guardado-Calvo et al., 2016).

STAR METHODS TEXT

LEAD CONTACT

Further information and request for resources and reagents should be directed to and will be fulfilled by the Lead Contact Felix A. Rey (rey@pasteur.fr).

MATERIALS AVAILABILITY

The unique/stable reagents generated in this study are available from the Lead Contact with a completed Materials Transfer Agreement.

DATA AND CODE AVAILABILITY

The atomic coordinates and structure factors of the X-ray structures have been deposited in the Protein Data Bank (www.pdb.org) with accession codes 6Y5W, 6Y5F, 6Y62, 6Y68, 6Y6Q, 6Y6P, 6YRB, and 6YRQ. The cET map has been deposited in the Electron Microscopy Data Bank (www.ebi.ac.uk/pdbe/emdb/) with accession number EMD-11236. Atomic coordinates of the tetrameric spike quasi-atomic model have been deposited in the Protein Data Bank with the accession code 6ZJM. The previously reported "inverted" map EMD-3364 has been redeposited with accession code EMD-4867. The scripts used to process the cryo-ET data are available at https://github.com/OPIC-Oxford/TomoPreprocess (TomoPreprocess) and https://github.com/OPIC-Oxford/PatchFinder (PatchFinder).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Cell lines.

Drosophila S2 cells (ThermoFisher) line was derived from a primary culture of late stage Drosophila melanogaster embryos. They were grown at 28C in serum-free insect cell medium (GE Healthcare HyClone). 293FT cells (Thermo Fisher Scientific) is a female human embryonic kidney cell line. They were grown at 37C with 8% CO_2 in DMEM medium (ThermoFisher) supplemented with 10% FCS. Vero E6 cells (CRL-1587) were purchased from ATCC and culture in DMEM media (Sigma) supplemented with 3% fetal calf serum (FCS).

METHODS DETAILS

Protein expression and purification

In order to obtain a soluble Gn/Gc complex amenable to crystallize, we followed the approach used to crystallize the alphavirus p62/E1 glycoprotein heterodimer ectodomains (Li et al., 2010; Voss et al., 2010). The strategy involved making single-chain constructs spanning the ectodomains of both glycoproteins, in this case the hantavirus Gn and Gc, bypassing the TM

segments in the precursor polyprotein by a soluble, flexible linker. The linker we used was 43-amino acids long in total with aa sequence GGSG<u>LVPRGS</u>GGGSGGGS**WSHPQFEK**GGGTGGGT<u>LVPRGS</u>GTGG. It contained two thrombin cleavage sites (underlined) at either end and a strep-tag sequence (in bold) in the middle, separated by flexible GGGS or GGGT repeats. To optimize protein production and help in the purification, we cloned synthetic codon-optimized genes for expression in Drosophila cells (Genscript) into a modified pMT/BiP plasmid (Invitrogen, hereafter termed pT350), which translates the protein in frame with an enterokinase cleavage site and a double strep-tag at the C-terminal end of the Gc sequence.

The plasmids generated were used to obtain stable transfectants of Drosophila S2 cells together with the pCoPuro plasmid (ratio 1:20) for puromycin selection. The stable cell lines were selected and maintained in serum-free insect cell medium (GE Healthcare HyClone) containing 7 μg/ml puromycin. Cultures of 1 liter were grown in spinner flasks in serum-free insect cell medium (GE Healthcare HyClone) supplemented with 1% penicillin/streptomycin antibiotics to about 1×10^7 cells/mL, and the protein expression was induced with 4 μM $CdCl_2$. After 5 days, the S2 media supernatant was concentrated to 40 ml and supplemented with 10 μg/mL avidin and 0.1M Tris-HCl pH 8.0, centrifuged 30 minutes at 20,000 g and purified by strep-tactin affinity chromatography followed size exclusion chromatography using a Superdex 200 10/30 column (GE Healthcare) in 10 mM Tris-HCl 8.5, 150 mM NaCl, 1mM EDTA (hereafter buffer TN).

Initially, we produced a single chain containing the complete ectodomains of Gn (aa 21–484) and Gc (aa 652–1107) of Andes virus (strain Chile-9717869, NCBI code NP_604472.1). The majority of the protein obtained with this construct formed heterogeneous aggregates from which we could not obtain crystals. We therefore generated a different construct with the linker described above joining the Gn head (Gn^H, aa 21–374, Figure S1A) (Li et al., 2016; Rissanen et al., 2017) to the complete Gc ectodomain. Drosophila Schneider 2 cells stably transfected with the Gn^H/Gc construct secreted large amounts of protein, which were purified using affinity and size exclusion chromatography (SEC). The proteins behaved mostly as monomers in solution as assessed by multi-angle light scattering (MALS) (data not shown). Using the same approach, we produced Gn^H/Gc heterodimers from two hantavirus orthologs, Maporal virus (MAPV, strain HV-97021050, NCBI code YP_009362281.1) and from Longquan virus (LQNV, strain Longquan-Ra-90, NCBI code AGI62346.1). We also introduced the H953F mutation in Gc of the Andes virus construct to destabilize its transition to the post-fusion form (as explained in the text). The aa sequence identity of the polyprotein precursor of Andes viruses with MAPV and LQNV is 85% and 40%, respectively. All constructs yielded large amount of soluble Gn^H/Gc secreted into the cell's supernatant. The LQNV construct led to the secretion of large molecular weight aggregates only, whose characterization we did not pursue.

We used the same plasmid for S2 cells expression to produce the isolated HNTV Gn^H (strain 76-118, UniprotKB code P08668.1), the complete ANDV and MAPV Gc ectodomains, ANDV Gn^B (aa. 375-484) and the isolated ANDV Gn domain C (aa 375–450). All these constructs secreted large amounts of protein that could be purified to homogeneity by affinity and size exclusion chromatography.

Crystallization and structure determination

Gn^H/Gc heterodimers: The fractions eluted from the SEC run corresponding to monomeric ANDV-Gn^H/Gc, ANDV-Gn^H/Gc-H953F, and MAPV-Gn^H/Gc were concentrated to 7 mg/mL using a Vivaspin centricon in 10 mM Tris-HCl 8.5, 150 mM NaCl (buffer TN). The flexible linker joining Gn^H and Gc was removed by adding 1 unit of biotinylated thrombin (Novagen) per mg of protein, and the mixture was incubated for 5 days at 4°C until the digestion was completed as judged by SDS-PAGE. These samples were used without further purification in crystallization trials at 18°C using the sitting-drop vapor diffusion method. Crystals of ANDV-Gn^H/Gc grew in the presence of 15-16% (w/v) PEG 3350, 125-400 mM potassium acetate, those of ANDV-Gn^H/Gc-H953F in 2.4M (NH$_4$)$_2$HPO$_4$, 0.1 M Tris-HCl pH 8.5, and those of MAPV-Gn^H/Gc in 0.1 M MgCl$_2$, 0.1 M Tris pH 8.5 and 17% (w/v) PEG 20000. Crystals were flash-frozen by immersion into a cryo-protectant containing the crystallization solution supplemented with 20% (v/v) glycerol, followed by rapid transfer into liquid nitrogen.

The structure of ANDV-Gn^H/Gc was solved by molecular replacement (MR) with Phaser from the suite PHENIX (Liebschner et al., 2019) using PUUV Gn^H (PDB code 5FXU) and HNTV-Gc (5lJY) as search models. The final model was built by combining real space model building in Coot (Emsley et al., 2010) with reciprocal space refinement with phenix.refine. The final model, which was validated with Molprobity (Williams et al., 2018), contained aa 23–374 of Gn and 659–1069 of Gc with two disordered regions: aa 124–126 (loop i$_2$D$_A$) and 697–702 (loop C$_0$D$_0$). The structures of ANDV-Gn^H/Gc-H953F, and MAPV-Gn ^{H}/Gc were obtained by MR using the structure of ANDV-Gn ^{H}/Gc as search model. The final model of ANDV-Gn^H/Gc-H953F contains aa 22–374 in Gn and 661–1092 in Gc, with a break at Gc residues 1063–1076 (the N-terminal part of the stem). The structure of MAPV-Gn^H/Gc contains aa 22–374 in Gn and 653–1083 in Gc; the N-terminal part of the stem (residues 1065–1076) was also disordered.

Gc postfusion trimers: The protein from the SEC factions corresponding to monomers of ANDV-Gc and MAPV-Gc was concentrated to around 10 mg/mL in TN buffer and used in crystallization experiments. Crystals of MAPV-Gc appeared in an acidic condition containing 30% (v/v) PEG 400, 0.1 M CdCl$_2$, 0.1M Na-acetate pH 4.6. The structures were determined by MR using HNTV-Gc (5LJZ) as initial model, showed the characteristic post-fusion trimeric conformation of Gc around a crystallographic 3-fold axis. The final model contains aa 657–1069 without internal breaks.

Crystals of proteolyzed ANDV-Gn^H/Gc were grown in a condition containing 16% (w/v) PEG 4000, 10% (v/v) 2-propanol, 0.2M $(NH_4)_2SO_4$, 0.1M Hepes pH 7.5. The final model contains aa 659–1069 and showed density for almost all the residues but three short breaks: 697–700 (loop C_0D_0), 765–766 and 775–778 (loop *cd*).

ANDV Gn^B crystals: The purified protein was concentrated to around 7 mg/mL in TN buffer for crystallization trials. Crystals appeared after 2-4 months at 18°C in two different conditions: 30% (v/v) MPD, 20 mM CaCl2, 0.1 M Na-acetate pH 4.6 (hereafter termed the acidic condition), and 35% (v/v) MPD, 0.2M NaCl, 0.1 M Hepes pH 7.5 (the neutral pH condition). Crystals growing in the acidic condition belonged to orthorhombic C222$_1$ space group and diffracted to 1.9 Å resolution. The Mathews coefficient (Kantardjieff and Rupp, 2003) indicated 4 molecules in the asymmetric unit, and a self-rotation function clearly indicated the presence of a non-crystallographic 4-fold axis. Crystals grown in the neutral pH condition belonged to the tetragonal space group I4 with two molecules in the asymmetric unit and diffracted to 2.4 Å resolution. We used experimental phasing with sodium iodide to determine their structure, as we found no good search model for MR. We soaked crystals for 20 minutes in mother liquor supplemented with 500 mM NaI before plunging them directly into liquid nitrogen. In order to optimize the anomalous signal, we collected a highly redundant dataset merging three independent datasets collected from two different crystals using a kappa goniometer. The final SAD dataset showed anomalous signal up to 3.6 Å. Substructure determination and phasing were carried out using the SHELX suite (Sheldrick, 2010) and an initial model was built using *Phenix AutoBuild* (Terwilliger et al., 2008). The final model was obtained by combining real space model building in *Coot* with reciprocal space refinement using *phenix.refine*. It contains aa 379–483 without Ramachandran outliers. Unexpectely, we found clear density between protomers in the tetramer that could be assigned a short RNA oligonucleotide (3 pyrimidine nucleotides). We used this model to solve the structure of the crystals grown at acidic pH by molecular replacement. The structure showed 4 molecules in the asymmetric unit forming a tetramer virtually identical to that observed at neutral pH, including the presence of the RNA oligo between protomers.

Crystals of HTNV Gn^H at neutral pH. The reported structures of HNTV Gn^H were derived from crystals grown at acidic pH (Rissanen et al., 2017). We determined the HTNV Gn^H structure at neutral pH by concentrating the purified protein in TN buffer and made crystallization experiments at 18°C. We obtained crystals diffracting to 1.9 Å growing at pH 7.5 in a condition containing 25% (w/v) PEG 6000, 0.1M LiCl, 0.1M Hepes pH 7.5, and 20% (v/v) glycerol. We determined the structure using PUUV Gn^H (5FXU) as search model. The resulting HTNV Gn^H structure resolved aa 20–373 with a single internal break in the η1 loop (aa 281–290).

The X-ray diffraction data reported in this manuscript were recorded at beamlines Proxima I and II at the Synchrotron Soleil (St. Aubin, France). The statistics of all the crystals and refinement are provided in Table 1.

Amino acid sequence conservation analyses

The conservation scores per residue shown in Figures S1 and S3 were obtained by using the program Espript (Robert and Gouet, 2014) and a multiple sequence alignment generated by Clustal omega (Sievers et al., 2011) using a set of 69 hantavirus M segment sequences extracted from (Zhang et al., 2013). The GenBank accession numbers for the sequences used in the analysis are: JX465398, JX465397, JX465402, JX465399, JX465400, JX465401, JF784178, NC_010708, EF641799, EF641806, HQ834696, FJ539167, EU929073, EU929074, HM015219, EF543526, JX465390, JX465391, JX465392, JX465393, JX465394, JX465395, JX465403, Y00386, AF143675, AB620029, AY675353, AB027115, GF796031, AY168577, AJ410616, GQ205412, AY961616, JQ082301, AF288298, GU592827, EF990916, L08756, GQ274938, HM756287, AB297666, AB433850, EF198413, AJ011648, EU072489, AJ011647, EU072488, Z69993, X55129, L36930, DQ177347, L39950, DQ284451, DQ285047, AB620104, AB620107, U26828, AB620101, U36801, U36802, AF030551, L33474, L37903, AF291703, AF028024, AF005728, FJ608550, AY363179, AF307323.

VLP expression and design of Gn/Gc cysteine mutants

For ANDV VLP expression we used the plasmid pI.18/GPC that encodes the full-length glycoprotein precursor of ANDV strain CHI-7913 under the control of the cytomegalovirus promotor (termed pI.18/GPC) (Cifuentes-Munoz et al., 2011). Site-directed mutagenesis was done by DNA synthesis and sub-cloning into pI.18/GPC using intrinsic restriction sites (GenScript). Briefly, 293FT cells (Thermo Fisher Scientific) maintained in DMEM supplemented with 10% FCS were grown in 100 mm plates and calcium-transfected with the pI.18/GPC wild type or the different mutant constructs. 48 h later the cell surface proteins were labeled with biotin in order to separate the biotinylated (surface proteins) from non-biotinylated (intracellular proteins) fractions using a cell surface protein isolation kit (Pierce). For protein detection, two western blots were made separately for each of three independent VLP preparations, using primary anti-Gc monoclonal antibodies 2H4/F6 and 5D11/G7 (Cifuentes-Munoz et al., 2011; Godoy et al., 2009), anti-Gn monoclonal antibody 6B9/F5 (Cifuentes-Munoz et al., 2011) or anti-β-actin Mab (Sigma) were used at 1:2.500 and subsequently detected with an anti-mouse immunoglobulin horseradish peroxidase conjugate (Thermo Fisher Scientific) 1:5.000 and a chemiluminescent substrate (Pierce). All these antibodies were previously characterized (Cifuentes-Munoz et al., 2011). For Gn/Gc VLP production, the supernatant of

transfected 293FT cells were harvested at 48 h post-transfection and concentrated as previously established (Acuna et al., 2015).

Tula virus growth and purification

Tula virus (TULV, strain Moravia) was cultivated on Vero E6 cells (ATCC 94 CRL-1586), as previously described (Huiskonen et al., 2010). Four days post infection (dpi), the growth medium (DMEM, Sigma) was replaced by medium supplemented with 3% fetal calf serum (FCS). At 5, 6 and 7 dpi the virus-containing medium was collected and clarified by centrifugation (3,000 × g for 30 min) to remove large cell debris. The medium was then concentrated 100-fold using a 100-kDa cut-off filter (Amicon) and placed on top of a 25–65% sucrose density gradient (in standard buffer [20 mM Tris and 100 mM NaCl pH 7.0]) in a SW32.1 tube (Beckman Coulter), and the virus was banded by ultracentrifugation (SW32 Ti rotor, 25,000 rpm, 4°C, 12 h). Virus-containing fractions were pooled and diluted 1:1 in 20 mM Tris and 100 mM NaCl before pelleting by ultracentrifugation (Beckman Coulter TLS 55 rotor, 50,000 rpm, 4°C, 2 h). The virus pellet was resuspended in the standard buffer.

An aliquot (3µl) of each sample was applied to 1.2-µm hole carbon grids (C-flat, Protochips) that were then floated on a droplet of standard buffer to remove residual sucrose. An aliquot (3 µl) of 6-nm gold fiducial markers was added to enable tilt series alignment. The sample was then vitrified by plunge freezing to liquid ethane-propane mixture by using a vitrification apparatus (CP3, Leica).

Chirality of the hantavirus spike.

To experimentally assess the hand of the Tula virus cET map, we prepared EM grids as previously described with Tula virus particles premixed with a sample of recombinantly produced human papillomavirus type 16 (HPV16) pseudovirus capsids (a kind gift from Dr. Kay Grünewald). HPV particles are chiral, and their hand is known from previous studies (Conway and Meyers, 2009) and can be used as reference. We collected a total of 7 tilt series from –45 to 45 degrees in 5-degree increments with a dose of 5.5 e⁻/Å² per tilt. For each tilt image, we acquired 8 movie frames at a calibrated magnification of ×37,037, corresponding to a pixel size of 1.35 Å. The defocus ranged from 3 µm to 4 µm between the tilt series. We reconstructed the tomograms as described in the paragraph below. We did STA of both TULV spikes and HPV particles in Dynamo following exactly the same workflow, using initial references low pass filtered at 50 Å to limit model bias. We imposed icosahedral symmetry for HPV averaging using as reference the EMD-5991 map (Lee et al., 2015) and C4 symmetry for TULV averaging using as reference EMD-3364 (Li et al., 2016). Because of potential template bias, we did multiple refinements changing the hand of the reference, or using a centrosymmetric map obtained by averaging the reference map with its mirror image, thereby

eliminating any potential template bias. For HPV, the procedure using the centrosymmetric reference converged to the correct hand, while using the mirror image it did not. For Tula virus, the procedure converged to the same hand in all cases (Figure Supplementary S2).

EM Data Acquisition and Processing

To improve the resolution of the TULV cET map, we collected additional data sets using a Tecnai F30 "Polara" transmission electron microscope (FEI) operated at 300 kV and at liquid nitrogen temperature. We used SerialEM (Mastronarde, 2005) to acquire images on a direct electron detector (K2 Summit; Gatan) mounted behind an energy filter (QIF Quantum LS; Gatan). The data set included a total of 49 tilt series from –30 to 60 degrees in 3-degree increments with a dose of 4.7 e⁻/Å² per tilt. Each tilt image was composed of 6 movie frames at a calibrated magnification of ×25,000, corresponding to a pixel size of 2 Å. The defocus ranged from 2.5 µm to 4.0 µm between the tilt series. Movie frames were aligned and averaged using motioncor2 to correct for beam induced motion (Zheng et al., 2017). Contrast transfer function (CTF) parameters were estimated using CTFFIND4 (Rohou and Grigorieff, 2015) and a dose-weighting filter was applied to the images according to their accumulated electron dose as described previously (Grant and Grigorieff, 2015). These pre-processing steps were carried out using a custom set of scripts named "tomo_preprocess" (https://github.com/OPIC-Oxford/TomoPreprocess). Tilt images were then aligned using gold fiducial markers in IMOD (Kremer et al., 1996), corrected for the effect of CTF by phase flipping and used to reconstruct the tomograms by weighted back projection in IMOD (Mastronarde and Held, 2017). Subtomogram averaging was performed in Dynamo (Castano-Diez et al., 2012). Initial particles for this refinement were created by modelling the surface of the membrane using the Dynamo's tomoview function and defining their initial orientation normal to this surface. These initial 'seeds' were more densely populated than the true spikes and overlapping spikes were removed based on cross-correlation after each iteration in the refinement. Refinements were carried out with C4 symmetry using the existing Tula virus spike map as a reference (EMD-3364). The map was low-pass filtered to 50 Å resolution to avoid model bias. A custom script (PatchFinder; available at https://github.com/OPIC-Oxford/PatchFinder) was used to restrict the dataset to only those particles that having regular glycoprotein surface patches. The script locates the eight closest spikes for each particle, and these are classified as interacting neighbors only if their lattice constraints are within set tolerances (maximum 49-Å and 25-degree deviation from expected location). Spikes were defined as being part of a lattice if they had at least three interacting neighbors. The final dataset was subjected to 3D classification in RELION (Bharat and Scheres, 2016) before further refinement in Dynamo. The amplitudes of the final maps were weighted to correct for the effect of dose-weighting of the original images using a custom script (available on request). The final reconstruction, comprised of ~18,064

subvolumes, was filtered to 11.4 Å as determined by FSC (0.143 threshold). The threshold value for the rendered isosurface was determined according to the molecular weight of the viral ectodomain proteins assuming an average protein density of 0.81 Da/Å³.

QUANTIFICATION AND STATISTICAL ANALYSIS

Two separate western-blots were performed on each of the three independent preparations (n=3) of each sample as described in the Methods section.

SUPPLEMENTAL VIDEO TITLE AND LEGEND

Supplementary Movie 1. Quality of the docking into the cET map (related to Figure 2). The movie samples various views of the Tula virus cET map with the ANDV models for Gn and Gc docked, showing the quality of the fit.

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Table 1. Crystallographic statistics

	ANDV-Gn^{H}Gc pH 8.5	ANDV-Gn^{H}Gc (H953F) pH 8.5	MAPV-Gn^{H}Gc pH 8.5	ANDV-Gc pH 7.5	MAPV-Gc pH 4.6	HNTV-Gn^H pH 7.5	Gn^B pH 4.6	Gn^B pH 7.5
Data collection								
Space group	H 3	P 6_3 2 2	C 1 2 1	H 32	H 32	P 2_1 2_1 2_1	C 2 2 2_1	I 4
a (Å)	130.3	188.5	120.3	92.6	90.4	53.6	90.6	67.8
b (Å)	130.3	188.5	65.1	92.6	90.4	92.2	123.1	67.8
c (Å)	129.5	117.3	144.9	318.5	506.4	94.0	90.1	121.9
β (°)	90	90	99.7	90	90	90	90	90
Resolution (Å)	37.62-2.55	49.36-3.20	38.07-2.20	39.80-2.70	39.83-2.40	35.34-1.94	45.3-1.90	37.7-2.35
Last resolution bin (Å)	2.66-2.55	3.42-3.20	2.26-2.20	2.83-2.70	2.49-2.40	1.98-1.94	1.94-1.90	2.43-2.35
Total observations	68478 (8692)	335389 (59067)	258945 (13723)	139957 (18675)	482431 (52803)	289204 (18180)	542782(31473)	114803 (10501)
Unique reflections	25995 (3241)	20744 (3657)	55909 (4505)	14898 (1949)	31958 (3310)	35256 (2248)	39873 (2451)	11445 (1080)
Completeness (%)	97.3 (99.8)	99.8 (99.4)	99.5 (98.9)	99.8 (99.9)	100 (100)	99.6 (95.6)	99.8(97.0)	99.6(96.4)
Redundancy	2.6 (2.7)	16.2 (16.2)	4.6 (3.0)	9.4 (9.6)	15.1 (16.0)	8.2 (8.1)	13.6(12.8)	10.0(9.7)
<I/σ>	11.2 (1.4)	13.5 (1.2)	11.0 (0.9)	8.2 (1.2)	11.1 (1.0)	14.6 (1.8)	11.1(0.9)	8.1(1.7)
Rsym (%)	5.7 (73.5)	19.2 (269.5)	8.2 (114.1)	13.6 (120)	16.9 (262)	7.6 (105)	11.2(288)	18.4(121)
CC$_{1/2}$	99.7 (49.0)	99.9 (53.2)	99.7 (33.8)	99.7 (91.8)	99.8 (50.6)	99.9 (65.5)	99.8(53.7)	98.8(45.5)
Refinement								
PDB accesion code	6Y5W	6Y5F	6Y62	6Y6Q	6Y68	6Y6P	6YRQ	6YRB
Resolution (Å)	37.62-2.55	49.63-3.20	35.14-2.20	39.80-2.70	39.83-2.40	35.35-1.94	45.3-1.90	37.7-2.35
Last resolution bin (Å)	2.64-2.55	3.36-3.20	2.24-2.20	2.79-2.70	2.44-2.40	1.99-1.94	1.94-1.90	2.58-2.35
Nb.of reflections	25987 (2465)	20680 (2685)	55841 (2606)	14734 (2625)	31948 (2798)	35189 (2661)	39739 (2560)	11444 (2691)
Nb.of reflections for Rfree	1360 (165)	1032 (157)	2814 (132)	1338 (122)	2914 (128)	1673 (153)	1957(123)	570(126)
Rfactor (%)	21.8 (33.0)	21.4 (34.2)	19.4 (31.4)	23.4 (37.7)	19.6 (32.2)	16.7 (23.0)	18.7(38.2)	22.6(29.8)
Rfree (%)	26.1 (35.7)	26.6 (44.9)	22.6 (35.8)	28.7 (34.2)	23.6 (33.6)	21.0 (25.2)	23.0(42.2)	26.5(34.2)
Nb. of protein+sugar atoms (chain A/B/C/D)	2776/3144	2809 / 3201	2801/3208	3084	3204	2745	767/744/ 772/806	803/744
Nb. of waters/Cd^{+2}/I$^-$	64	1	261	4	83/4	202	137	55/0/6
Nb. of (PO$_4$)$^{-2}$/(SO$_4$)$^{-2}$ atoms		10/0		0/10				
Nb. of RNA/MPD atoms							256/32	130
Mean B value (Å^2)								
Protein+sugar (A/B/C/D)	85.6/57.1	104.8 / 104.0	52.3/63.9	98.0	70.2	46.5	52/54/53/55	46/48
Waters/ Cd^{+2}/I$^-$	50.0	78.6	49.4	66	57.0/125	47.6	52	43/-/115
(PO4)$^{-2}$/(SO$_4$)$^{-2}$ atoms		161		0/152				
RNA/MPD							53/67	52
Root mean square deviations								
Bond lengths (Å)	0.002	0.007	0.006	0.003	0.008	0.018	0.006	0.002
Bond angles (°)	0.56	1.111	0.707	0.600	0.935	1.419	0.923	0.546
Ramachandran favored/outliers (%)	93.6/0.0	91.7/0.5	95.0/0.0	94.3/0.26	95.4/0.5	94.7/0.3	97.1/0.0	97.38/0.0

Table 2. EM fitting statistics

	Unconstrained fitting[1]		Constrained fitting[2]	
	CC	% atoms inside	CC	% atoms inside
ANDV Gn^H/Gc (H953F)				
Central	0.888	88.9	0.861	82.8
Neighbour	0.880	80.0	0.834	69.9
Dimer	0.884	84.5	0.857	76.3
MAPV Gn^H/Gc				
Central	0.873	84.6	0.812	74.8
Neighbour	0.867	76.5	0.814	63.9
Dimer	0.870	80.6	0.816	69.4
ANDV Gn^B				
Central			0.882	87.1

1. Fitting a protomer of ANDV-Gn^H/Gc (H953F) or MAPV Gn^H/Gc
2. Fitting a dimer of ANDV-Gn^H/Gc (H953F) or MAPV Gn^H/Gc and a tetramer of ANDV Gn^B

Hantavirus particle

(Gn/Gc)₄ tetramer docked into the cET map
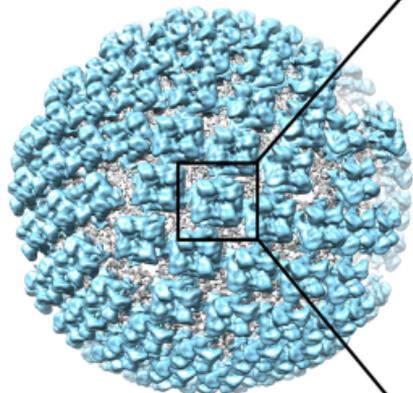
120 Å

Lattice of tetrameric spikes formed by glycoproteins Gn and Gc

20 Å
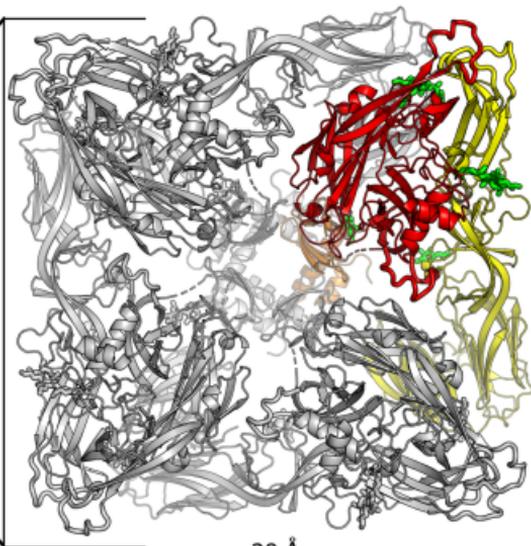
One Gn/Gc promer highlighted in colors:
Gn: red/orange; Gc: yellow