



**HAL**  
open science

## **New Bacteriophages against Emerging Lineages ST23 and ST258 of *Klebsiella pneumoniae* and Efficacy Assessment in *Galleria mellonella* Larvae**

Damien Thiry, Virginie Passet, Katarzyna Danis-Wlodarczyk, Cédric Lood, Jeroen Wagemans, Luisa de Sordi, Vera van Noort, Nicolas Dufour, Laurent Debarbieux, Jacques G Mainil, et al.

### ► To cite this version:

Damien Thiry, Virginie Passet, Katarzyna Danis-Wlodarczyk, Cédric Lood, Jeroen Wagemans, et al.. New Bacteriophages against Emerging Lineages ST23 and ST258 of *Klebsiella pneumoniae* and Efficacy Assessment in *Galleria mellonella* Larvae. *Viruses*, 2019, 11 (5), pp.411. 10.3390/v11050411 . pasteur-03164319

**HAL Id: pasteur-03164319**

**<https://pasteur.hal.science/pasteur-03164319>**

Submitted on 9 Mar 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

Communication

# New Bacteriophages against Emerging Lineages ST23 and ST258 of *Klebsiella pneumoniae* and Efficacy Assessment in *Galleria mellonella* Larvae

Damien Thiry <sup>1,2,3,\*</sup>, Virginie Passet <sup>2</sup>, Katarzyna Danis-Wlodarczyk <sup>3,‡</sup>, Cédric Lood <sup>3,4</sup>, Jeroen Wagemans <sup>3</sup>, Luisa De Sordi <sup>5,6</sup>, Vera van Noort <sup>4,7</sup>, Nicolas Dufour <sup>5,8</sup>, Laurent Debarbieux <sup>5</sup>, Jacques G. Mainil <sup>1</sup>, Sylvain Brisse <sup>2,†</sup> and Rob Lavigne <sup>3,†</sup>

<sup>1</sup> Bacteriology, Department of Infectious and Parasitic Diseases, FARAH and Faculty of Veterinary Medicine, ULiège, 4000 Liège, Belgium; jg.mainil@uliege.be

<sup>2</sup> Biodiversity and Epidemiology of Bacterial Pathogens, Institut Pasteur, 75015 Paris, France; virginie.passet@pasteur.fr (V.P.); sylvain.brisse@pasteur.fr (S.B.)

<sup>3</sup> Laboratory of Gene Technology, Department of Biosystems, KU Leuven, 3001 Heverlee, Belgium; danis@wp.pl (K.D.-W.); cedric.lood@kuleuven.be (C.L.); jeroen.wagemans@kuleuven.be (J.W.); rob.lavigne@kuleuven.be (R.L.)

<sup>4</sup> Centre of Microbial and Plant Genetics, Department of Microbial and Molecular Systems, KU Leuven, 3001 Heverlee, Belgium; vera.vannoort@kuleuven.be

<sup>5</sup> Department of Microbiology, Institut Pasteur, 75015 Paris, France; luisa.de\_sordi@sorbonne-universite.fr (L.D.S.); nicolas.dufour@ght-novo.fr (N.D.); laurent.debarbieux@pasteur.fr (L.D.)

<sup>6</sup> Laboratoire des Biomolécules, Hôpital Saint-Antoine, Sorbonne Université, 75012 Paris, France

<sup>7</sup> Institute of Biology Leiden, Leiden University, 2311 Leiden, The Netherlands

<sup>8</sup> Service de Réanimation Médico-Chirurgicale, Centre Hospitalier René Dubos, 95300 Pontoise, France

\* Correspondence: damien.thiry@uliege.be; Tel.: +32-4-3669388

† These authors contributed equally to this work.

‡ Current Address: Department of Microbiology and Microbial Infection and Immunity, The Ohio State University, 43210 Columbus, OH, USA.

Received: 14 February 2019; Accepted: 29 April 2019; Published: 3 May 2019



**Abstract:** *Klebsiella pneumoniae* is a bacterial pathogen of high public health importance. Its polysaccharide capsule is highly variable but only a few capsular types are associated with emerging pathogenic sublineages. The aim of this work is to isolate and characterize new lytic bacteriophages and assess their potential to control infections by the ST23 and ST258 *K. pneumoniae* sublineages using a *Galleria mellonella* larvae model. Three selected bacteriophages, targeting lineages ST258 (bacteriophages vB\_KpnP\_KL106-ULIP47 and vB\_KpnP\_KL106-ULIP54) and ST23 (bacteriophage vB\_KpnP\_K1-ULIP33), display specificity for capsular types KL106 and K1, respectively. These podoviruses belong to the *Autographivirinae* subfamily and their genomes are devoid of lysogeny or toxin-associated genes. In a *G. mellonella* larvae model, a mortality rate of 70% was observed upon infection by *K. pneumoniae* ST258 and ST23. This number was reduced to 20% upon treatment with bacteriophages at a multiplicity of infection of 10. This work increases the number of characterized bacteriophages infecting *K. pneumoniae* and provides information regarding genome sequence and efficacy during preclinical phage therapy against two prominent sublineages of this bacterial species.

**Keywords:** antimicrobial resistance; capsule; *Galleria mellonella*; *Klebsiella pneumoniae*; phage therapy

---

*Klebsiella pneumoniae*, a member of the Enterobacteriaceae family, causes a variety of human and animal infections including pneumonia, infections of the urinary tract, bacteremia, and liver abscess.

*K. pneumoniae* infections are becoming increasingly difficult, and sometimes impossible [1], to treat due to the continuous emergence of multidrug-resistant strains [2–4]. Cells of *K. pneumoniae* are characteristically surrounded by a thick capsule of variable chemical composition, which translates into a large number of classically defined capsular serotypes [5] and an even larger number of in silico-defined *wzi*, *wzc*, or KL-types [6,7]. These three molecular classifications denote the diversity of the capsular polysaccharide synthesis gene cluster and serve as proxies of capsular antigen variation. *K. pneumoniae* isolates can be roughly classified into two pathotypes: opportunistic *K. pneumoniae*, which are often multidrug-resistant (mdrKp), and hypervirulent *K. pneumoniae* (hvKp) [8,9], which are able to infect healthy individuals and cause invasive infections including pyogenic liver abscess. The majority of clinical mdrKp and hvKp isolates are part of a small number of genetic lineages (also called clonal groups). Prominent lineages include mdrKpST258, which is frequently associated with specific carbapenemases (i.e., those of the KPC family) and resistant to multiple other antimicrobials, and the ST23 lineage, which is the most frequent cause of liver abscess [1] and can also acquire clinically significant antibiotic resistance genes [10]. Recently, there has been a sharp increase in the clinical significance of mdrKp and hvKp infections [4,9,11].

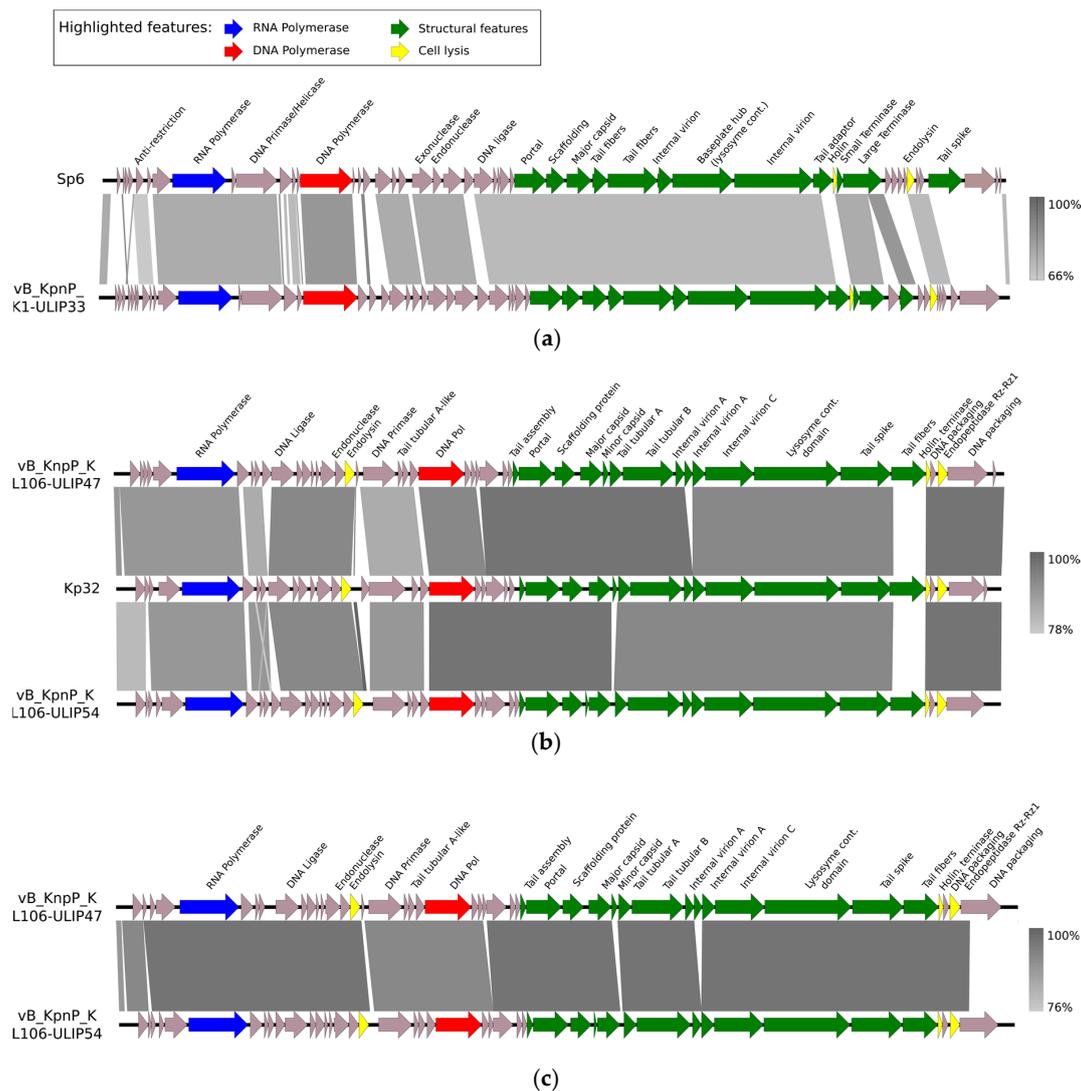
New therapeutic strategies are critically needed against *K. pneumoniae* infections. Phage therapy is increasingly recognized as an attractive approach [12]. Previous work has shown that bacteriophages (phages) against *Klebsiella* can be readily isolated from diverse sources and are a promising tool against *K. pneumoniae* infections in *Galleria mellonella* models [13,14].

The aim of this study was to contribute to developments of the phage therapy approach against *K. pneumoniae* and, more specifically, against its two prominent lineages ST23 and ST258. Specifically, our objectives were (i) to isolate and characterize phages against bacteria in these lineages and to sequence the genome of these phages; (ii) to implement an infection model of *G. mellonella* larvae with *K. pneumoniae* strains of interest; and (iii) to test phages against *K. pneumoniae* in this model.

Two clinical *K. pneumoniae* strains were selected for phage isolation [6,15,16]. The first was the 2198 (SB4551) strain, a *K. pneumoniae* carbapenemase-producing isolate from an outbreak in Ireland [15]. This strain, characterized by *wzc*-921 and *wzi*-29 alleles, belongs to ST258 clade 1 [17,18] or ST258a [19] associated with the production of a newly described capsular polysaccharide [20]. It carries *bla*<sub>KPC-2</sub> and *bla*<sub>TEM-1</sub> genes, as well as a chromosomal *bla*<sub>SHV-11</sub> gene; aminoglycoside resistance genes *aac6-Ib* and *aadA2*; mutations in the QRDR region of quinolone targets (ParC-80I, GyrA-83I); genes conferring resistance to phenicols, sulfonamide, tetracycline, and trimethoprim (*catA1*, *sullI*, *tetB*, *dfrA12*); and has no virulence genes. The second strain was SA12 (SB4385), an ST23, K1 capsular-type isolate from a human liver abscess infection in France [9]. It carried virulence genes for yersiniabactin (*ybt 1*; *ICEKp10*), colibactin (*clb 2*), aerobactin (*iuc 1*), salmochelin (*iro 1*), and the regulator of mucoid phenotype genes *rmpA* and *rmpA2*; it has no resistance genes except for the chromosomal gene *bla*<sub>SHV-11</sub>. Phages vB\_KpnP\_KL106-ULIP47 and vB\_KpnP\_KL106-ULIP54 were isolated against 2198 and phage vB\_KpnP\_K1-ULIP33 was isolated against SA12; all three from wastewater collected in France (Clichy, Saint-Denis, and Rueil-Malmaison, respectively) in 2015 using standard procedures [21]. Briefly, the wastewater samples were centrifuged at 4000 rpm for 10 min to remove large particles, then filtered and sterilized (0.45 µm). A first enrichment step was performed at 37 °C for 24 h with gentle agitation (50 rpm). When a clarification of the medium was observed, it was then centrifuged at 5000 g for 10 min and 20 µL of supernatant was spread on the surface of LB agar and then covered by a liquid culture of the target bacteria. After incubation for 18 h at 37 °C, individual plaques were selected and purified three times following the same procedure. These three phages produced large, clear plaques surrounded by a halo zone (Figure S1) reflecting the potential presence of an exopolysaccharide depolymerase [22]. The pH, temperature, storage stability, and the lysis kinetic curves were assessed (Figures S2–S5). The host range of the isolated phages was determined using a set of 23 *Klebsiella* spp. strains representative of diverse species and capsular serotypes (Table S1). Based on standard spot assays [23], the three phages showed specificity for the capsular type of their original bacterial host. vB\_KpnP\_K1-ULIP33 showed clear lysis specifically against the K1 strains,

whereas vB\_KpnP\_KL106-ULIP47 and vB\_KpnP\_KL106-ULIP54 were specific for the “undefined” capsular type of their parental strain (KL106, *wzi* 29) (Table S1). This capsular specificity probably reflects the need for phages to first adsorb to and depolymerize the thick capsule. The depolymerases allowing the disruption of the polysaccharide capsule are generally K-type specific in *Klebsiella* [24–26].

To analyze the genome of these phages, polyethylene glycol (PEG) precipitation was performed, followed by CsCl density gradient (layers of 1.33, 1.45, 1.50, and 1.70 g/cm<sup>3</sup>) ultracentrifugation (28,000 g; 3 h; 4 °C), dialysis using Slide-A-Lyzer dialysis cassettes G2 (Thermo Fisher Scientific Inc., Merelbeke, Belgium) and, finally, DNA extraction [27,28]. A sequencing library was obtained using the NEBNext Ultra DNA kit (New England Biolabs, Ipswich, MA, USA) and sequenced using an Illumina MiSeq instrument equipped with a nanoFlowcell (Illumina MiSeq Reagent Nano Kit v2, Brussels, Belgium, paired-end 2\*250 bp reads). After correction of reads (Trimmomatic v0.38) [29], assembly (SPAdes v3.9) [30], and analysis of the genome ends (PhageTerm v1.0.11) [31], the average read coverage depths of the assemblies were 550×, 423×, and 815× for phages vB\_KpnP\_K1-ULIP33, vB\_KpnP\_KL106-ULIP47, and vB\_KpnP\_KL106-ULIP54, respectively. Annotation was performed with the RAST server using the virus domain option [32] followed by manual curation. All genomic data related to this project, including raw Illumina read and GenBank annotation, are available via the NCBI BioProject PRJNA488998. GenBank accession numbers are MK380014 (vB\_KpnP\_K1-ULIP33), MK380015 (vB\_KpnP\_KL106-ULIP47), and MK380016 (vB\_KpnP\_KL106-ULIP54). All three phages carry a linear dsDNA genome with predicted direct repeats, totaling 44,122 bp (vB\_KpnP\_K1-ULIP33), 41,397 bp (vB\_KpnP\_KL106-ULIP47), and 41,109 bp (vB\_KpnP\_KL106-ULIP54). Phage vB\_KpnP\_K1-ULIP33 has direct repeats of length 163 nt, whereas phages vB\_KpnP\_KL106-ULIP47 and vB\_KpnP\_KL106-ULIP54 have direct repeats of 180 nt. Comparative genomics of vB\_KpnP\_K1-ULIP33 with Enterobacteria phage Sp6, and of vB\_KpnP\_KL106-ULIP47 and vB\_KpnP\_KL106-ULIP54 with *Klebsiella* phage KP32, illustrate their genetic relatedness to reference phages [33,34] and the conserved genome organization of the *Autographivirinae* subfamily (Figure 1a–c). Distinguishing features of this phage subfamily include a unidirectional and progressive transcriptional scheme, regulated by the presence of a single subunit RNAP driving the middle/late expression. Analysis of the tailspike proteins with HMMER and HHPRED suggested the presence of tailspike-associated depolymerases present in phages vB\_KpnP\_KL106-ULIP47 (locus D3A56\_0040) and vB\_KpnP\_KL106-ULIP54 (locus D3A57\_0040), consistent with the presence of expanding halos in the plaques [35]. These proteins typically show a conserved (T7-related, gp17) N-terminal connector (aa1–154 pfam03906) and diverse C-terminal domains, associated with predicted pectate lyase domains. Pectate lyase domains were previously shown to have depolymerase activity against *Acinetobacter baumannii* polysaccharide capsules and against extracted exopolysaccharides [36,37]. These domains are likely associated with the capsular specificity of these phages [38,39]. Although vB\_KpnP\_K1-ULIP33 also induced a halo zone around the clear region of plaque lysis, suggestive of a putative depolymerase activity, no depolymerase domain was predicted. However, a tailspike protein (locus D3A55\_0041) was found to have a conserved N-terminal phage\_T7 connector domain (aa3–171 pfam03906). No gene related to phage lysogeny was predicted, suggesting that these phages are strictly lytic, which is an important prerequisite for phage therapy [40]. The location of the lysis cassette genes in vB\_KpnP\_KL106-ULIP47 and vB\_KpnP\_KL106-ULIP54 suggests a typical T7-related genome organization in which the endolysin is located among the middle genes, presumably having a secondary function as a regulator of the phage-encoded RNAP.



**Figure 1.** Comparative genomics (nucleic acid sequence) of (a) vB\_KpnP\_K1-ULIP33 with Enterobacteria phage Sp6 (Genus Sp6virus, AY288927), (b) vB\_KpnP\_KL106-ULIP47, vB\_KpnP\_KL106-ULIP54 with *Klebsiella* phage *Klebsiella pneumoniae* 32 (Genus *K. pneumoniae* 32virus, MH172262); and (c) vB\_KpnP\_KL106-ULIP47 with vB\_KpnP\_KL106-ULIP54.

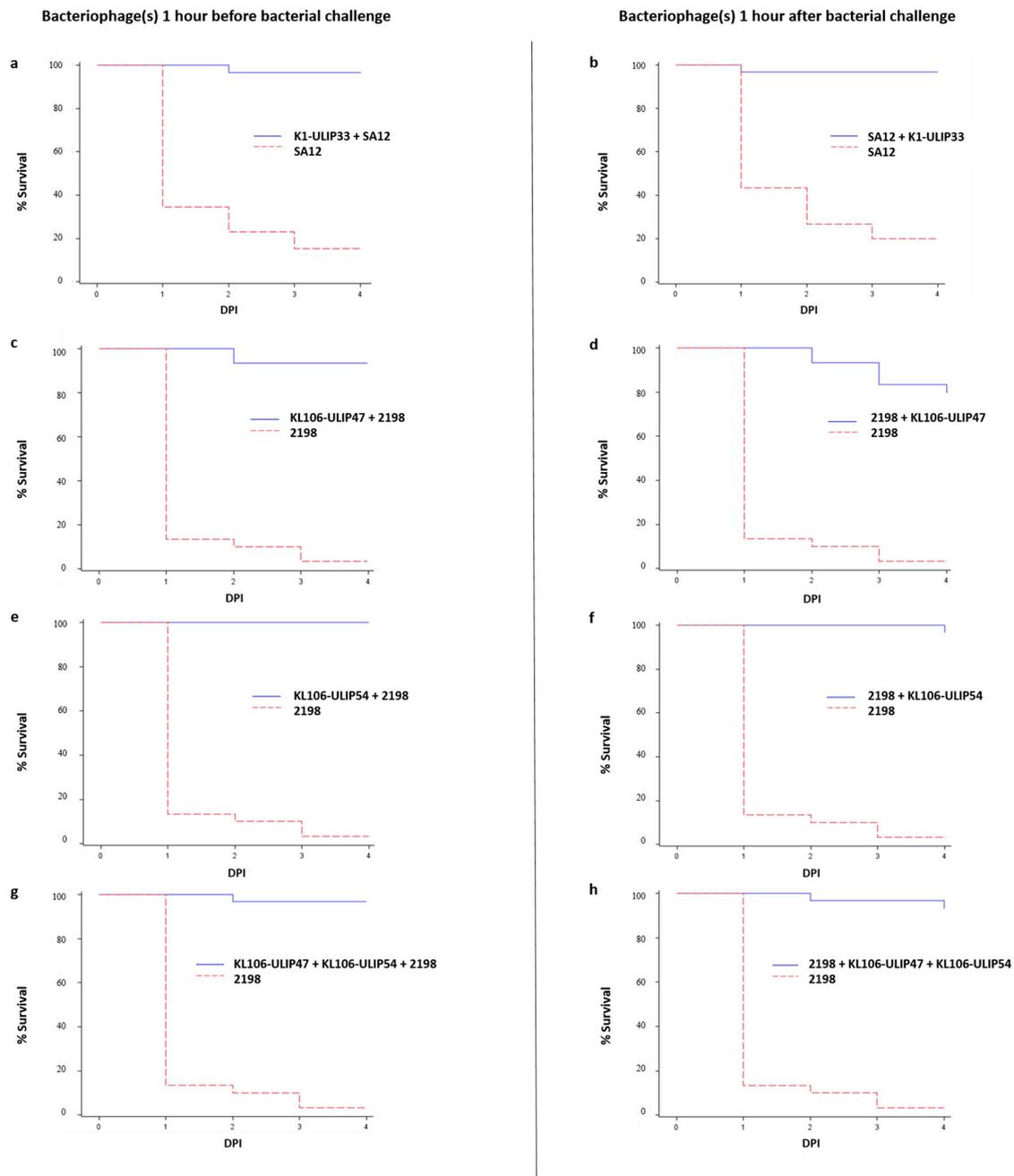
To assess the potential in vivo efficacy of phages against *K. pneumoniae* in a preclinical setting with an emphasis on the prevention of infection, a *G. mellonella* larvae model was used. This model allows testing phages within a more complex system than Petri dishes and has interesting features, including similarities between the systemic cellular and humoral immune responses of these larvae and the inflammatory responses of the mammalian innate immune system [41]. Previous reports have found this model to be useful for studies of the virulence of *K. pneumoniae* and for therapeutic approaches [14,42–44]. We first determined that the optimal inoculum concentration for *K. pneumoniae* infection was  $10^4$  CFU/10  $\mu$ L, as this dose induced a mortality rate of 70–90% in 4 days, both for strain 2198 and for strain SA12. We confirmed (data not shown) that the mortality of larvae infected with *K. pneumoniae* was dose-dependent [43]. We next assessed phage efficacy against *K. pneumoniae* infection in two independent experimental setups.

In the first experiment, we assessed the efficacy of phage vB\_KpnP\_K1-ULIP33 against infection by strain SA12. A total of 150 larvae were divided into five groups of 10 larvae with technical triplicates (Table S2a). In a second experiment, we analyzed the individual or combined effect of phages vB\_KpnP\_KL106-ULIP47 and vB\_KpnP\_KL106-ULIP54 on strain 2198. Here, a total of 330

larvae were divided into five groups and 11 subgroups of 10 larvae in technical triplicates (Table S2b). In both experiments, phages were administered either 1 h prior to bacterial infection (group A) or 1 h post-bacterial inoculation (group B). The timing of phage inoculation was selected in order to allow the spread of bacteria within the larvae but without allowing enough time for the infection to develop. Groups C, D, and E corresponded to assays of phage toxicity, infectivity control, and injection safety, respectively. Phages were inoculated with a multiplicity of infection (MOI) close to 10 on the left last proleg and the bacterial inoculation was performed on the right last proleg. The concentrations of the inoculated *K. pneumoniae* SA12 and 2198 were verified and were, respectively,  $2 \times 10^4$  CFU/10  $\mu$ L and  $7 \times 10^3$  CFU/10  $\mu$ L. The titers of the phage inoculums were also verified after inoculation and were  $2 \times 10^5$  PFU/10  $\mu$ L for vB\_KpnP\_K1-ULIP33,  $2 \times 10^5$  PFU/10  $\mu$ L for vB\_KpnP\_KL106-ULIP47, and  $7 \times 10^4$  PFU/10  $\mu$ L for vB\_KpnP\_KL106-ULIP54. Data from each independent experiment were pooled and the protection of the *G. mellonella* larvae by the phages was assessed with the log-rank test ( $p$ -values  $< 0.005$  were considered as statistically significant). The Kaplan–Meier analyses were performed with the LIFETEST procedure of SAS version 9.4 for Windows and graphs were designed with SAS<sup>®</sup> ODS Graphics Editor.

Considering the different technical replicates, the first experiment, which tested the in vivo efficacy of vB\_KpnP\_K1-ULIP33 against SA12, showed that only 0–30% of the larvae survived in the infected groups at 4 days post-inoculation (DPI), whereas the survival rates of prophylactic and treatment groups ranged from 90% to 100%. In the second experiment, which tested the in vivo efficacy of vB\_KpnP\_KL106-ULIP47 and vB\_KpnP\_KL106-ULIP54 against strain 2198, 0–10% of the larvae survived in the infected groups at 4 DPI, whereas the survival rates of prophylactic and treatment groups ranged from 80% to 100%. In both experiments, groups of larvae inoculated with phage (but not bacteria) showed comparable survival rates as the PBS control groups, ranging from between 70% and 100% (Figure S1a,b). The survival curves are presented in Figure 2; data from the triplicate experiments were pooled. Protection of the *G. mellonella* larvae by the phages was found to be statistically significant ( $p$ -values  $< 0.0001$  for each experiment). No significant difference was observed between the cocktail and the monophage groups. Note that despite their different stabilities (Figures S2–S5), these phages have high genetic relatedness and similar host ranges, and may therefore not be the best candidates for a phage cocktail.

These data show that the three studied phages could efficiently prevent a *K. pneumoniae* infection induced by their host strains. Both phage only and PBS control groups showed similar survival rates, demonstrating the safety of the phages in this model. A recent report indicated protection against *K. pneumoniae* ST258 infection in *G. mellonella* with another phage [14]. The present study confirms that strains belonging to this ST can be targeted by phages and reports, for the first time, on phage efficacy against ST23 *K. pneumoniae* in *G. mellonella*. The very low MOI used in this study allowed for assessment of the efficacy of the phages while avoiding the phenomenon of “lysis from without”. Overall, this study confirms that the *G. mellonella* is a flexible and rapid tool to assess phage efficacy. Indeed, it accommodates many human pathogenic strains in contrast to rodent models and it allows a quick (less than 48 h in this study) evaluation of the killing activity of phages in vivo. However, the relevance of the *G. mellonella* model to predict the phage efficacy in higher animals including humans, and in particular, with higher MOIs and timings of phage administration, remains to be determined [44].



**Figure 2.** Kaplan–Meier survival curves of the *Galleria mellonella* larvae inoculated with *K. pneumoniae* SA12 (ST23) (a,b) and *K. pneumoniae* 2198 (ST258) (c–h) with, respectively, phage vB\_KpnP\_K1-ULIP33 (K1-ULIP33), and phages vB\_KpnP\_KL106-ULIP47 (KL106-ULIP47) and vB\_KpnP\_KL106-ULIP54 (KL106-ULIP54), one hour before or one hour after bacterial inoculation.

**Supplementary Materials:** The following are available online at <http://www.mdpi.com/1999-4915/11/5/411/s1>, Figure S1: Picture of halo zones of Phages (a) vB\_KpnP\_K1-ULIP33, (b) vB\_KpnP\_KL106-ULIP47 and (c) vB\_KpnP\_KL106-ULIP54. Figure S2: The temperature stability of phages vB\_KpnP\_K1-ULIP33 (A), vB\_KpnP\_KL106-ULIP47 (B), and vB\_KpnP\_KL106-ULIP54 (C). Figure S3: The pH stability of phages vB\_KpnP\_K1-ULIP33 (A), vB\_KpnP\_KL106-ULIP47 (B), and vB\_KpnP\_KL106-ULIP54 (C). Figure S4: The storage stability of phages vB\_KpnP\_K1-ULIP33 (A), vB\_KpnP\_KL106-ULIP47 (B), and vB\_KpnP\_KL106-ULIP54 (C) at 4 °C. Figure S5: Lysis kinetic curves of vB\_KpnP\_K1-ULIP33 lysis on the SB4385 strain (A), vB\_KpnP\_KL106-ULIP47 (B), and vB\_KpnP\_KL106-ULIP54 (C) on the SB4551 strain. Table S1: Bacterial strains characteristics and phages spot assays results. Table S2: Experimental designs of the main *Galleria mellonella* experiments with (a) *K. pneumoniae* SA12 (ST23) and phage vB\_KpnP\_K1-ULIP33 and (b) *K. pneumoniae* 2198 (ST258), phage vB\_KpnP\_KL106-ULIP47,

and vB\_KpnP\_KL106-ULIP54. Each group contains 10 larvae and each experiment condition was reproduced in technical triplicates.

**Author Contributions:** Conceptualization, S.B., R.L., L.D. and D.T. Methodology, S.B., R.L. and D.T. Manipulations, D.T., V.P., K.D.-W., N.D., J.W. and L.D.S. Genomic analysis, C.L., R.L. and D.T. Writing—Original Draft Preparation, D.T., S.B. and R.L.; Writing—Review & Editing, All authors. Supervision, S.B., R.L., V.v.N. and J.G.M.

**Funding:** We thank the Biermans-Lapôte Foundation for a post-doctoral fellowship that supported D.T. work in Institut Pasteur. L.D.S. is funded by a Roux-Cantarini fellowship from the Institut Pasteur (Paris, France). R.L., K.D.-W. and J.W. are supported by a GOA grant from KU Leuven ('Phage Biosystems'), C.L. holds a SB PhD fellowship from FWO Vlaanderen (1S64718N). K.D.-W. was supported by a PDM mandate from KU Leuven.

**Acknowledgments:** We thank Dearbhaile Morris (NUIG, Galway, Ireland) for providing strain 2198 (SB4551), Dominique Decré (Saint-Antoine hospital, Paris, France) for providing strain SA12 (SB4385) and Carla Rodrigues for assistance with *wzi* and KL typing of the isolates.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Lee, C.R.; Lee, J.H.; Park, K.S.; Jeon, J.H.; Kim, Y.B.; Cha, C.J.; Jeong, B.C.; Lee, S.H. Antimicrobial Resistance of Hypervirulent *Klebsiella pneumoniae*: Epidemiology, Hypervirulence-Associated Determinants, and Resistance Mechanisms. *Front. Cell. Infect. Microbiol.* **2017**, *7*, 483. [[CrossRef](#)]
2. Pendleton, J.N.; Gorman, S.P.; Gilmore, B.F. Clinical relevance of the ESKAPE pathogens. *Expert. Rev. Anti. Infect. Ther.* **2013**, *11*, 297–308. [[CrossRef](#)]
3. Seiffert, S.N.; Marschall, J.; Perreten, V.; Carattoli, A.; Furrer, H.; Endimiani, A. Emergence of *Klebsiella pneumoniae* co-producing NDM-1, OXA-48, CTX-M-15, CMY-16, QnrA and ArmA in Switzerland. *Int. J. Antimicrob. Agents.* **2014**, *44*, 260–262. [[CrossRef](#)]
4. Davis, G.S.; Price, L.B. Recent Research Examining Links Among *Klebsiella pneumoniae* from Food, Food Animals, and Human Extraintestinal Infections. *Curr. Environ. Heal. Reports* **2016**, *3*, 128–135. [[CrossRef](#)] [[PubMed](#)]
5. Wyres, K.L.; Wick, R.R.; Gorrie, C.; Jenney, A.; Follador, R.; Thomson, N.R.; Holt, K.E. Identification of *Klebsiella capsule* synthesis loci from whole genome data. *Microb. Genom.* **2016**, *2*, e000102. [[CrossRef](#)] [[PubMed](#)]
6. Brisse, S.; Passet, V.; Haugaard, A.B.; Babosan, A.; Kassis-Chikhani, N.; Struve, C.; Decré, D. *wzi* Gene Sequencing, a Rapid Method for Determination of Capsular Type for *Klebsiella* Strains. *J. Clin. Microbiol.* **2013**, *51*, 4073–4078. [[CrossRef](#)]
7. Pan, Y.J.; Lin, T.L.; Chen, Y.H.; Hsu, C.R.; Hsieh, P.F.; Wu, M.C.; Wang, J.T. Capsular types of *Klebsiella pneumoniae* revisited by *wzc* sequencing. *PLoS ONE* **2013**, *8*, e80670. [[CrossRef](#)]
8. Shon, A.S.; Bajwa, R.P.S.; Russo, T.A. Hypervirulent (hyper mucoviscous) *Klebsiella pneumoniae*: A new and dangerous breed. *Virulence* **2013**, *4*, 107–118. [[CrossRef](#)] [[PubMed](#)]
9. Bialek-Davenet, S.; Criscuolo, A.; Ailloud, F.; Passet, V.; Jones, L.; Delannoy-Vieillard, A.S.; Garin, B.; Le Hello, S.; Arlet, G.; Nicolas-Chanoine, M.H.; Decré, D.; Brisse, S. Genomic definition of hypervirulent and multidrug-resistant *Klebsiella pneumoniae* clonal groups. *Emerg. Infect. Dis.* **2014**, *20*, 1812–1820. [[CrossRef](#)]
10. Cheong, H.S.; Chung, D.R.; Park, M.; Kim, S.H.; Ko, K.S.; Ha, Y.E.; Kang, C.I.; Peck, K.R.; Song, J.H. Emergence of an extended-spectrum  $\beta$ -lactamase-producing serotype K1 *Klebsiella pneumoniae* ST23 strain from Asian countries. *Epidemiol. Infect.* **2017**, *145*, 990–994. [[CrossRef](#)] [[PubMed](#)]
11. Holt, K.E.; Wertheim, H.; Zadoks, R.N.; Baker, S.; Whitehouse, C.A.; Dance, D.; Jenney, A.; Connor, T.R.; Hsu, L.Y.; Severin, J.; et al. Genomic analysis of diversity, population structure, virulence, and antimicrobial resistance in *Klebsiella pneumoniae*, an urgent threat to public health. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, E3574–E3581. [[CrossRef](#)]
12. Vandenhuevel, D.; Lavigne, R.; Brüssow, H. Bacteriophage Therapy: Advances in Formulation Strategies and Human Clinical Trials. *Annu. Rev. Virol.* **2015**, *2*, 599–618. [[CrossRef](#)]
13. Manohar, P.; Nachimuthu, R.; Lopes, B.S. The therapeutic potential of bacteriophages targeting gram-negative bacteria using *Galleria mellonella* infection model. *BMC Microbiol.* **2018**, *18*, 97. [[CrossRef](#)]

14. D'Andrea, M.M.; Marmo, P.; Henrici De Angelis, L.; Palmieri, M.; Ciacci, N.; Di Lallo, G.; Demattè, E.; Vannuccini, E.; Lupetti, P.; Rossolini, G.M.; et al.  $\phi$ BO1E, a newly discovered lytic bacteriophage targeting carbapenemase-producing *Klebsiella pneumoniae* of the pandemic Clonal Group 258 clade II lineage. *Sci. Rep.* **2017**, *7*, 2614. [[CrossRef](#)]
15. Morris, D.; Boyle, F.; Morris, C.; Condon, I.; Delannoy-Vieillard, A.S.; Power, L.; Khan, A.; Morris-Downes, M.; Finnegan, C.; Powell, J.; et al. Inter-hospital outbreak of *Klebsiella pneumoniae* producing KPC-2 carbapenemase in Ireland. *J. Antimicrob. Chemother.* **2012**, *67*, 2367–2372. [[CrossRef](#)] [[PubMed](#)]
16. Wick, R.R.; Heinz, E.; Holt, K.E.; Wyres, K.L. Kaptive Web: User-Friendly Capsule and Lipopolysaccharide Serotype Prediction for *Klebsiella* Genomes. *J. Clin. Microbiol.* **2018**, *56*, e00197-18. [[CrossRef](#)]
17. Deleo, F.R.; Chen, L.; Porcella, S.F.; Martens, C.A.; Kobayashi, S.D.; Porter, A.R.; Chavda, K.D.; Jacobs, M.R.; Mathema, B.; Olsen, R.J.; et al. Molecular dissection of the evolution of carbapenem-resistant multilocus sequence type 258 *Klebsiella pneumoniae*. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 4988–4993. [[CrossRef](#)] [[PubMed](#)]
18. Bowers, J.R.; Kitchel, B.; Driebe, E.M.; MacCannell, D.R.; Roe, C.; Lemmer, D.; de Man, T.; Rasheed, J.K.; Engelthaler, D.M.; Keim, P.; et al. Genomic Analysis of the Emergence and Rapid Global Dissemination of the Clonal Group 258 *Klebsiella pneumoniae* Pandemic. *PLoS ONE* **2015**, *10*, e0133727. [[CrossRef](#)] [[PubMed](#)]
19. Wright, M.S.; Perez, F.; Brinkac, L.; Jacobs, M.R.; Kaye, K.; Cober, E.; van Duin, D.; Marshall, S.H.; Hujer, A.M.; Rudin, S.D.; et al. Population structure of KPC-producing *Klebsiella pneumoniae* isolates from midwestern U.S. hospitals. *Antimicrob. Agents Chemother.* **2014**, *58*, 4961–4965. [[CrossRef](#)]
20. Bellich, B.; Ravenscroft, N.; Rizzo, R.; Lagatolla, C.; D'Andrea, M.M.; Rossolini, G.M.; Cescutti, P. Structure of the capsular polysaccharide of the KPC-2-producing *Klebsiella pneumoniae* strain KK207-2 and assignment of the glycosyltransferases functions. *Int. J. Biol. Macromol.* **2019**, *130*, 536–544. [[CrossRef](#)] [[PubMed](#)]
21. Van Twest, R.; Kropinski, A.M. Bacteriophage enrichment from water and soil. *Methods Mol. Biol.* **2009**, *501*, 15–21. [[PubMed](#)]
22. Hughes, K.A.; Sutherland, I.W.; Clark, J.; Jones, M.V. Bacteriophage and associated polysaccharide depolymerases—novel tools for study of bacterial biofilms. *J. Appl. Microbiol.* **1998**, *85*, 583–590. [[CrossRef](#)] [[PubMed](#)]
23. Kutter, E. Phage host range and efficiency of plating. *Methods Mol. Biol.* **2009**, *501*, 141–149.
24. Lin, T.L.; Hsieh, P.F.; Huang, Y.T.; Lee, W.C.; Tsai, Y.T.; Su, P.A.; Pan, Y.J.; Hsu, C.R.; Wu, M.C.; Wang, J.T. Isolation of a bacteriophage and its depolymerase specific for K1 capsule of *Klebsiella pneumoniae*: Implication in typing and treatment. *J. Infect. Dis.* **2014**, *210*, 1734–1744. [[CrossRef](#)]
25. Maciejewska, B.; Olszak, T.; Drulis-Kawa, Z. Applications of bacteriophages versus phage enzymes to combat and cure bacterial infections: an ambitious and also a realistic application? *Appl. Microbiol. Biotechnol.* **2018**, *102*, 2563–2581. [[CrossRef](#)] [[PubMed](#)]
26. Hsu, C.R.; Lin, T.L.; Pan, Y.J.; Hsieh, P.F.; Wang, J.T. Isolation of a bacteriophage specific for a new capsular type of *Klebsiella pneumoniae* and characterization of its polysaccharide depolymerase. *PLoS ONE* **2013**, *8*, e70092. [[CrossRef](#)]
27. Ceyssens, P.J.; Miroshnikov, K.; Mattheus, W.; Krylov, V.; Robben, J.; Noben, J.P.; Vanderschraeghe, S.; Sykilinda, N.; Kropinski, A.M.; Volckaert, G.; et al. Comparative analysis of the widespread and conserved PB1-like viruses infecting *Pseudomonas aeruginosa*. *Environ. Microbiol.* **2009**, *11*, 2874–2883. [[CrossRef](#)]
28. Green, M.R.; Sambrook, J. Preparation of Single-Stranded Bacteriophage M13 DNA by Precipitation with Polyethylene Glycol. *Cold Spring Harb. Protoc.* **2017**, *2017*, pdb.prot093419. [[CrossRef](#)]
29. Bolger, A.M.; Lohse, M.; Usadel, B. Trimmomatic: A flexible trimmer for Illumina sequence data. *Bioinformatics* **2014**, *30*, 2114–2120. [[CrossRef](#)]
30. Bankevich, A.; Nurk, S.; Antipov, D.; Gurevich, A.A.; Dvorkin, M.; Kulikov, A.S.; Lesin, V.M.; Nikolenko, S.I.; Pham, S.; Pribelski, A.D.; et al. SPAdes: A New Genome Assembly Algorithm and Its Applications to Single-Cell Sequencing. *J. Comput. Biol.* **2012**, *19*, 455–477. [[CrossRef](#)]
31. Garneau, J.R.; Depardieu, F.; Fortier, L.-C.; Bikard, D.; Monot, M. PhageTerm: A tool for fast and accurate determination of phage termini and packaging mechanism using next-generation sequencing data. *Sci. Rep.* **2017**, *7*, 8292. [[CrossRef](#)] [[PubMed](#)]
32. Aziz, R.K.; Bartels, D.; Best, A.A.; DeJongh, M.; Disz, T.; Edwards, R.A.; Formsma, K.; Gerdes, S.; Glass, E.M.; Kubal, M.; et al. The RAST Server: Rapid Annotations using Subsystems Technology. *BMC Genomics*. **2008**, *9*, 75. [[CrossRef](#)] [[PubMed](#)]

33. Dobbins, A.T.; George, M., Jr.; Basham, D.A.; Ford, M.E.; Houtz, J.M.; Pedulla, M.L.; Lawrence, J.G.; Hatfull, G.F.; Hendrix, R.W. Complete genomic sequence of the virulent Salmonella bacteriophage SP6. *J. Bacteriol.* **2004**, *186*, 1933–1944. [[CrossRef](#)]
34. Kęsik-Szeloch, A.; Drulis-Kawa, Z.; Weber-Dąbrowska, B.; Kassner, J.; Majkowska-Skrobek, G.; Augustyniak, D.; Lusiak-Szelachowska, M.; Zaczek, M.; Górski, A.; Kropinski, A.M. Characterising the biology of novel lytic bacteriophages infecting multidrug resistant *Klebsiella pneumoniae*. *Viol. J.* **2013**, *10*, 100. [[CrossRef](#)] [[PubMed](#)]
35. Cornelissen, A.; Ceysens, P.J.; Krylov, V.N.; Noben, J.P.; Volckaert, G.; Lavigne, R. Identification of EPS-degrading activity within the tail spikes of the novel *Pseudomonas putida* phage AF. *Virology* **2012**, *434*, 251–256. [[CrossRef](#)]
36. Oliveira, H.; Costa, A.R.; Konstantinides, N.; Ferreira, A.; Akturk, E.; Sillankorva, S.; Nemeč, A.; Shneider, M.; Dötsch, A.; Azeredo, J. Ability of phages to infect *Acinetobacter calcoaceticus*-*Acinetobacter baumannii* complex species through acquisition of different pectate lyase depolymerase domains. *Environ. Microbiol.* **2017**, *19*, 5060–5077. [[CrossRef](#)] [[PubMed](#)]
37. Liu, Y.; Mi, Z.; Mi, L.; Huang, Y.; Li, P.; Liu, H.; Yuan, X.; Niu, W.; Jiang, N.; Bai, C.; et al. Identification and characterization of capsule depolymerase Dpo48 from *Acinetobacter baumannii* phage IME200. *PeerJ* **2019**, *7*, e6173. [[CrossRef](#)] [[PubMed](#)]
38. Olszak, T.; Shneider, M.M.; Latka, A.; Maciejewska, B.; Browning, C.; Sycheva, L.V.; Cornelissen, A.; Danis-Włodarczyk, K.; Senchenkova, S.N.; Shashkov, A.S.; et al. The O-specific polysaccharide lyase from the phage LKA1 tailspike reduces *Pseudomonas* virulence. *Sci. Rep.* **2017**, *7*, 16302. [[CrossRef](#)]
39. Oliveira, H.; Costa, A.R.; Ferreira, A.; Konstantinides, N.; Santos, S.B.; Boon, M.; Noben, J.P.; Lavigne, R.; Azeredo, J. Functional Analysis and Antivirulence properties of a new depolymerase from a Myovirus that infects *Acinetobacter baumannii* capsule K45. *J. Virol.* **2019**, *93*, e01163-18. [[CrossRef](#)]
40. Pirnay, J.-P.; Verbeken, G.; Ceysens, P.-J.; Huys, I.; De Vos, D.; Ameloot, C.; Fauconnier, A. The Magistral Phage. *Viruses* **2018**, *10*, 64. [[CrossRef](#)]
41. Desbois, A.P.; Coote, P.J. Utility of Greater Wax Moth Larva (*Galleria mellonella*) for Evaluating the Toxicity and Efficacy of New Antimicrobial Agents. *Adv. Appl. Microbiol.* **2012**, *78*, 25–53. [[PubMed](#)]
42. Wei, W.J.; Yang, H.F.; Ye, Y.; Li, J.B. *Galleria mellonella* as a model system to assess the efficacy of antimicrobial agents against *Klebsiella pneumoniae* infection. *J. Chemother.* **2017**, *29*, 252–256. [[CrossRef](#)] [[PubMed](#)]
43. Insua, J.L.; Llobet, E.; Moranta, D.; Pérez-Gutiérrez, C.; Tomás, A.; Garmendia, J.; Bengoechea, J.A. Modeling *Klebsiella pneumoniae* Pathogenesis by Infection of the Wax Moth *Galleria mellonella*. *Infect. Immun.* **2013**, *81*, 3552–3565. [[CrossRef](#)] [[PubMed](#)]
44. Forti, F.; Roach, D.R.; Cafora, M.; Pasini, M.E.; Horner, D.S.; Fiscarelli, E.V.; Rossitto, M.; Cariani, L.; Briani, F.; Debarbieux, L.; et al. Design of a Broad-Range Bacteriophage Cocktail That Reduces *Pseudomonas aeruginosa* Biofilms and Treats Acute Infections in Two Animal Models. *Antimicrob. Agents Chemother.* **2018**, *62*, e02573-17. [[CrossRef](#)]

