



Serotype Distribution of Clinical *Streptococcus pneumoniae* Isolates before the Introduction of the 13-Valent Pneumococcal Conjugate Vaccine in Cambodia

Malin Inghammar, Youlet By, Christina Farris, Thong Phe, Laurence Borand, Alexandra Kerleguer, Sophie Goyet, Vonthanak Saphonn, Chanleakhena Phoeung, Sirenda Vong, et al.

► To cite this version:

Malin Inghammar, Youlet By, Christina Farris, Thong Phe, Laurence Borand, et al.. Serotype Distribution of Clinical *Streptococcus pneumoniae* Isolates before the Introduction of the 13-Valent Pneumococcal Conjugate Vaccine in Cambodia. *American Journal of Tropical Medicine and Hygiene*, 2018, 98 (3), pp.791 - 796. 10.4269/ajtmh.17-0692 . pasteur-01739337

HAL Id: pasteur-01739337

<https://pasteur.hal.science/pasteur-01739337>

Submitted on 20 Mar 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Serotype Distribution of Clinical *Streptococcus pneumoniae* Isolates before the Introduction of the 13-Valent Pneumococcal Conjugate Vaccine in Cambodia

Malin Inghammar,^{1,2*} Youlet By,^{3,4} Christina Farris,⁵ Thong Phe,⁶ Laurence Borand,¹ Alexandra Kerleguer,¹ Sophie Goyet,¹ Vonthanak Saphonn,⁴ Chanleakhena Phoeung,⁴ Sirenda Vong,¹ Blandine Rammaert,⁷ Charles Mayaud,¹ Bertrand Guillard,¹ Chadwick Yasuda,⁵ Matthew R. Kasper,⁵ Gavin Ford,⁵ Steven W. Newell,⁵ Ung Sam An,⁸ Buth Sokhal,⁸ Sok Touch,⁹ Paul Turner,^{10,11} Jan Jacobs,^{6,12,13} Mélina Messaoudi,¹⁴ Florence Komurian-Pradel,¹⁴ and Arnaud Tarantola¹

¹Institut Pasteur du Cambodge, Phnom Penh, Cambodia; ²Section for Infection Medicine, Department of Clinical Sciences Lund, Lund University, Skane University Hospital, Lund, Sweden; ³Fondation Mérieux, Phnom Penh, Cambodia; ⁴University of Health Science, Phnom Penh, Cambodia; ⁵Naval Medical Research Unit No. 2, Phnom Penh, Cambodia; ⁶Sihanouk Hospital Center of Hope, Phnom Penh, Cambodia; ⁷CHU de Poitiers, Service de Maladies Infectieuses et Tropicales, INSERM U1070, Université de Poitiers, Poitiers, France; ⁸Cambodian National Laboratory of Public Health, Phnom Penh, Cambodia; ⁹Cambodian Communicable Disease Control Department, Phnom Penh, Cambodia; ¹⁰Cambodia Oxford Medical Research Unit, Siem Reap, Cambodia; ¹¹Nuffield Department of Medicine, Centre for Tropical Medicine and Global Health, University of Oxford, Oxford, United Kingdom; ¹²Department of Clinical Sciences, Institute of Tropical Medicine, Antwerp, Belgium; ¹³Department of Microbiology and Immunology, KU Leuven, Leuven, Belgium; ¹⁴Emerging Pathogens Laboratory, Fondation Mérieux, Centre International de Recherche en Infectiologie, INSERM U1111, Lyon, France

Abstract. Childhood vaccination with the 13-valent pneumococcal conjugate vaccine (PCV13) was introduced in Cambodia in January 2015. Baseline data regarding circulating serotypes are scarce. All microbiology laboratories in Cambodia were contacted for identification of stored isolates of *Streptococcus pneumoniae* from clinical specimens taken before the introduction of PCV13. Available isolates were serotyped using a multiplex polymerase chain reaction method. Among 166 identified isolates available for serotyping from patients with pneumococcal disease, 4% were isolated from upper respiratory samples and 80% were from lower respiratory samples, and 16% were invasive isolates. PCV13 serotypes accounted for 60% (95% confidence interval [CI] 52–67) of all isolates; 56% (95% CI 48–64) of noninvasive and 77% (95% CI 57–89) of invasive isolates. Antibiotic resistance was more common among PCV13 serotypes. This study of clinical *S. pneumoniae* isolates supports the potential for high reduction in pneumococcal disease burden and may serve as baseline data for future monitoring of *S. pneumoniae* serotypes circulation after implementation of PCV13 childhood vaccination in Cambodia.

INTRODUCTION

Streptococcus pneumoniae (pneumococci) cause a wide spectrum of infections, ranging from invasive disease with a high case-fatality rate to asymptomatic colonization. Despite available antibiotics, it is estimated that around 800,000 children die every year due to pneumococcal disease, especially in developing countries where timely access to adequate health care is limited.¹

Pneumococci can be divided into more than 90 different serotypes, based on differences in their capsular polysaccharides, with varying ability to cause severe disease.^{2,3} The distribution of serotypes varies with age and geographical area, whereas the potential coverage rates of vaccines may differ by target population.^{4,5} In high-income countries, the incidence of invasive pneumococcal disease by included vaccine types has declined significantly with the introduction of the pneumococcal conjugated vaccine (PCV).^{6,7} Data from low-income countries are less robust.⁸

In Cambodia, lower respiratory infection is estimated to be the second leading cause of morbidity and mortality.⁹ A new vaccination program of newborns including PCV13 (including serotypes 1, 3, 4, 5, 6A, 6B, 7F, 9V, 14, 18C, 19A, 19F, 23F) was launched in 2015.¹⁰ Data on the serotype distribution in Cambodia before the vaccine introduction are scarce. Two recent studies from a single center in Siem Reap have assessed the serotype distribution in colonizing and invasive

strains in children.^{11,12} Data from other parts of Cambodia and data in adults, however, are lacking. The aim of the present study was to document the serotype distribution of *S. pneumoniae* in Cambodia before the introduction of PCV13 in the national childhood immunization program.

METHODS

All principal microbiological laboratories in Cambodia were contacted for identification of stored isolates of *S. pneumoniae* from specimens taken before January 2015 (i.e., before the introduction of PCV13). Stored strains were identified at 1) Institut Pasteur du Cambodge (IPC), Phnom Penh, as part of the Surveillance and investigation of Epidemics in South-East Asia (SISEA) project, a prospective study of lower respiratory infections in two provincial hospitals (Takeo and Kampong Cham provinces),¹³ as well as from routine cultures performed from 2006 to 2014; 2) Sihanouk Hospital Center of Hope (SHCH), Phnom Penh, systematically collected as part of a microbiological surveillance program “Surveillance of antimicrobial resistance among consecutive blood culture isolates in tropical settings,” 2008 through 2014; 3) Naval Medical Research Unit No. 2 (NAMRU 2), Phnom Penh, as part of a prospective surveillance study “Surveillance and Etiology of Acute Undifferentiated Febrile Illnesses in Cambodia” (Kandal, Kampong Speu, Kratie, Ratanakiri, Stung Treng, and Svay Rieng), 2005–2014. An overview of the origin of isolates and the participating microbiological laboratories are listed in Table 1. The laboratories at the following hospitals were contacted but none of them had any stored pneumococcal isolates from the study time period: National Pediatric Hospital; Kampong Cham; Takeo; Kampot; Battambang;

* Address correspondence to Malin Inghammar, Institut Pasteur du Cambodge, Phnom Penh, Cambodia, and Section for Infection Medicine, Department of Clinical Sciences Lund, Lund University, Lund, Sweden. E-mail: malin.inghammar@med.lu.se

TABLE 1
Origin of isolates and participating microbiological laboratories

Name of institution	Details	Specimen	Time period	Number of samples
Institut Pasteur du Cambodge, Phnom Penh	Surveillance and investigation of Epidemics in South-East Asia project. Prospective study of low respiratory infections in two provincial hospitals (Takeo and Kampong Cham) Ethical approval No. 048-NECHR. ¹³	Blood, pleural fluid, sputum	2007–2009	75
	Routine cultures	Blood, bronchoalveolar lavage, sputum	2006–2014	58
Sihanouk Hospital Center of Hope, Phnom Penh	Microbiological Surveillance Study “Surveillance of antimicrobial resistance among consecutive blood culture isolates in tropical settings.” Ethical approvals No: Ethical approval Nos. 009-NECHR and 0313-NECHR.	Blood, cerebrospinal fluid	2008–2014	15
Naval Medical Research Unit No. 2, Phnom Penh	Surveillance and Etiology of Acute Undifferentiated Febrile Illnesses in Cambodia (Kandal, Kampong Speu, Kratie, Ratanakiri, Stung Treng and Svay Rieng). Ethical approval No: NAMRU2.2005.0004.	Sputum, blood, cerebrospinal fluid	2005–2014	18

NECHR = National Ethics Committee in Cambodia.

Siem Reap; Calmette; Khmer Soviet; Kossamak; or Kantha Bopha.

Information was collected for each contributed sample on the following: date of culture; type of specimen; patient's date of birth; and on clinical diagnosis and antibiotic susceptibility, if available. All individual information was anonymized before being analyzed. The study was approved by the National Ethics Committee in Cambodia (No. 460-NECHR).

Serotyping method. The isolates from the respective participating sites had been processed using standard microbiological procedures at the contributing sites and stored at -80°C . Antimicrobial susceptibility had been determined by disk diffusion at the participating laboratories according to the successive versions of Clinical and Laboratory Standards Institute (CLSI) guideline “Performance Standards for Antimicrobial Susceptibility Testing – Supplement,” CLSI M100-S21 (SHCH, NAMRU 2) and the Antibiogram Committee of the French Society for Microbiology, CA-SFM/EUCAST (IPC).¹⁴ Results for penicillin, ceftriaxone, cotrimoxazole, and erythromycin were reported as susceptible (S), intermediate (I), or resistant (R). The isolates were typed in the Rodolphe Mérieux Laboratory of the University of Health Sciences, using a multiplex real-time polymerase chain reaction (PCR) method as described previously.¹⁵ Briefly, DNA was extracted directly from 100 μL stored isolates using an easyMAG automate (bioMérieux, Lyon, France) according to the manufacturer's recommendation and then typed using a panel of multiplex PCR, enabling the detection of the 40 most prevalent *S. pneumoniae* serotypes worldwide and including an internal positive control targeting the *lytA* gene, a gene conserved among pneumococci. A subset of the isolates from the SHCH were serotyped using latex agglutination method with Quellung confirmation of ambiguous results at Angkor Hospital for Children/Cambodia Oxford Medical Research Unit microbiology laboratory.^{16,17}

Data analyses. The pneumococcal serotypes were grouped into PCV13 vaccine types (1, 3, 4, 5, 6A, 6B, 7F, 9V, 14, 18C, 19A, 19F, 23F) and nonvaccine types (all others, including non-typeable). Groups were compared using a Wilcoxon Rank Sum test, χ^2 , or Fisher's exact test, as

appropriate. The distribution of serotype groups in invasive isolates (blood, cerebrospinal fluid [CSF], and pleural fluid) and noninvasive isolates (all other isolates) were compared. In case of multiple samples per patient and index date, only the first isolate was included. A sensitivity analysis was performed, including only isolates originating from prospective studies or prospective systematic sample collection (IPC-SISEA, NAMRU 2 and SHCH). Analyses were made using Stata SE, version 13.1 (StataCorp, College Station, TX).

RESULTS

In total, 249 isolates were identified at the participating institutions: 215 from IPC, 16 from SHCH, and 18 from NAMRU 2. Of these, we were unable to determine the serotype of 79 (32%) isolates because of sample contamination or degradation, four isolates were excluded as they stem from the same patient and index date, leaving 166 (67%) isolates in the analysis. Of these, 133 (80%) came from IPC, 15 (9%) from SHCH, and 18 (11%) from NAMRU 2. The basic characteristics of the isolates are shown in Table 2. Information on sex was available for 106/166 isolates.

Twenty-six (16%) were invasive isolates (blood, CSF, and joint fluid); six (4%) were isolated in upper respiratory samples (ear, eye, and nasopharyngeal swab) and 133 (80%) were from lower respiratory samples (bronchoalveolar lavage fluid or

TABLE 2
Basic characteristics of the patients

Total number	166 (100)
Age in years, median (range)	39 (1–84)
Age groups, <i>N</i> (%)	
0–15 years	36 (22)
16–65 years	101 (61)
> 65 years	29 (17)
Male sex, * <i>N</i> (%)	55 (52)
Time of sampling, <i>N</i> (%)	
Rainy season (May–October)	79 (48)
Dry season (November–April)	87 (52)

* Information on sex was available for 106/166 isolates.

sputum). Table 3 shows the serotype distribution per sample type.

The serotype distribution is shown in Figure 1. The three most common serotypes were 19F, 29/166 (17%), 23F, 26/166 (16%), and 34, 22/166 (13%).

Overall, PCV13 serotypes accounted for 60% (95% confidence interval [CI] 52–67) of the isolates (Table 3). Among non-invasive isolates, the proportion of PCV13 serotypes was 56% (95% CI 48–64) and among invasive isolates the proportion was 77% (95% CI 57–89) ($P = 0.05$). The proportion of PCV13 serotypes was similar between age groups: 25 (70%) among 36 isolates from patients ≤ 15 years of age; 59 (58%) among 101 isolates from patients 16–64 years old; and 15 (52%) among 29 isolates from patients aged 65 years or greater ($P = 0.32$).

In the sensitivity analyses among 108 isolates systematically collected as part of surveillance studies (NAMRU 2, SHCH, IPC-SISEA), the proportion of PCV13 serotypes was 50% (95% CI 41–60) overall; 42% (95% CI 32–53) and 76% (95% CI 55–89) among noninvasive and invasive isolates, respectively.

Data on antibiotic susceptibility were available for 127–165 of the isolates. Of these, 95/165 (58%) were reported susceptible for penicillin G and 70/165 (42%) were reported nonsusceptible (intermediate or resistant); 127/137 (77%) were reported susceptible to amoxicillin, 149/157 (95%) were reported susceptible to ceftriaxone; 15/164 (9%) were reported susceptible to cotrimoxazole; and 81/165 (49%) were reported susceptible to erythromycin (Table 4). Thirty-one percent of the isolates were reported nonsusceptible to three or more of these antibiotics. Multidrug antibiotic resistance was more common among PCV13-covered serotypes than among nonvaccine serotypes, the proportion of PCV13 coverage was 23% among the isolates with no reported resistance to these antibiotics; 59% among isolates with resistance to one or two of the antibiotics; and 71% among isolates with resistance to at least three of the antibiotics ($P = 0.006$).

DISCUSSION

Data from this retrospective study of clinical pneumococcal isolates in Cambodia, collected before the introduction of PCV13 in 2015, suggest that this vaccine potentially covers around 80% of the invasive isolates and around 60% of the

noninvasive isolates. The range of serotypes and predicted vaccine coverage is consistent with the two studies previously published. A survey (2013–2014) reported 63% PCV13 coverage in colonizing isolates in pediatric outpatients ($N = 601$) in Cambodia,¹² and 88% among invasive isolates ($N = 40$). A second study from the same center based on invasive isolates ($N = 50$) 2008–2012 predicted 92% PCV13 coverage.¹¹ Furthermore, a review of studies from neighboring countries in Southeast Asia, including both pediatric and adult data, estimated that PCV13 provides 46–72% coverage for the circulating isolates.¹⁸

Interestingly, serotype 34, which is not included in PCV13, was the third most common serotype. This result contrasts with that of a colonizing survey of children from Siem Reap, Cambodia, where the prevalence of serotype 34 was found low.¹² Serotype 34 was not found to play a major role in any of the 25 studies reviewed of pneumococcal disease in Southeast Asia. Notably, serotype 34 is believed to be less invasive,^{3,19} and the studies reviewed focused mainly on invasive disease. This could have affected the differences in importance attributed to serotype 34. Because of the limited sample size, it is not possible to infer whether the difference is due to a true increased prevalence compared with neighboring countries or due to sampling variation. Nevertheless, this serotype may become important in the post-PCV introduction era.

The frequency of antibiotic resistance was high in our sample set; overall 60% of the isolates were penicillin-susceptible. PCV13-covered serotypes were significantly less susceptible to penicillin, cotrimoxazole and erythromycin, as compared with the nonvaccine serotypes, whereas there was no significant difference in the frequency of susceptibility to amoxicillin or ceftriaxone. Furthermore, PCV13-covered serotypes were more likely to express resistance to more than one antibiotic class than non-PCV13 serotypes. This finding further supports a potential benefit of PCV13, reducing the incidence of infections caused by penicillin-resistant pneumococci or multidrug-resistant pneumococci.

This study has many limitations. The pre-hospitalization use of antibiotics is widespread in Cambodia, which explains why microbiological yield in cultures is generally low.¹² Furthermore, we relied on retrospective data on antimicrobial susceptibility testing collected over a long timeframe and at

TABLE 3
Serotype distribution per specimen

	Total number of samples	Samples included in PCV13* number (%)	Serotypes (n)†
Blood	21	18 (86)	1 (4), 14 (3), 19A (3), 19F (3), 23A (1), 23F (2), 34 (1), 38 (1), 6AB (1), 7F (1), 9V (1)
Cerebrospinal fluid	4	1 (25)	18C (1), 24F (2), 34 (1)
Joint fluid	1	1 (100)	23F (1)
Upper respiratory samples‡	6	4 (67)	11A (1), 14 (1), 19A (2), 19F (1), 23F (1), 35B (1)
Lower respiratory samples§	133	74 (56)	10A (1), 11A (2), 12F (1), 13 (4), 14 (4), 15A (2), 15BC (4), 17F (1), 18 (1), 19A (5), 19F (25), 20 (1), 21 (1), 22F (2), 23F (22), 3 (1), 34 (20), 35A (4), 35B (2), 35F (4), 38 (1), 6AB (16), 6C (2), 9V (1), non-typeable (6)
Total	166	99 (60)	1 (4), 10A (1), 11A (3), 12F (1), 13 (4), 14 (8), 15A (2), 15BC (4), 17F (1), 18 (1), 18C (1), 19A (10), 19F (29), 20 (1), 21 (1), 22F (2), 23A (1), 23F (26), 24F (2), 3 (1), 34 (22), 35A (4), 35B (3), 35F (4), 38 (2), 6AB (17), 6C (2), 7F (1), 9V (6), non-typeable (6)

* Serotypes included in PCV13—13-valent pneumococcal conjugate vaccine.

† Serotypes included in PCV13 are marked in bold.

‡ Upper respiratory samples: ear, eye, and nasopharyngeal swab.

§ Lower respiratory samples: sputum or bronchoalveolar fluid.

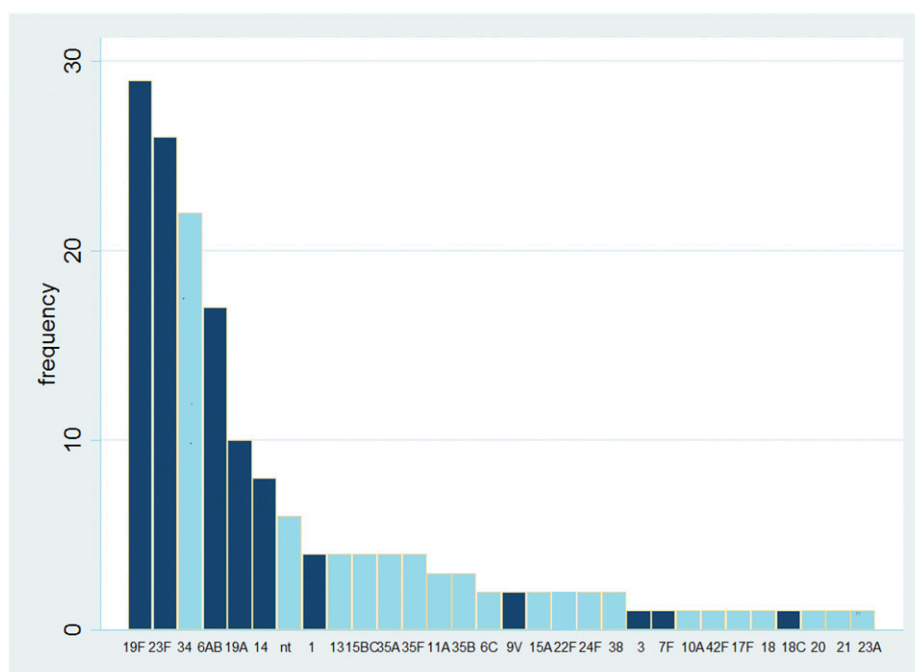


FIGURE 1. Serotype distribution of 166 isolates cultured from samples collected from 2006 to 2014 in Cambodia. Serotypes included in 13-valent pneumococcal conjugate vaccine are shaded in dark. Non-typeable strains are denoted "nt." This figure appears in color at www.ajtmh.org.

different centers; despite internal and external quality management at the participating laboratories, we cannot exclude errors at the time of assessment. Pre-hospitalization antimicrobial use effective against susceptible nonvaccine serotypes could potentially have biased our results toward a higher proportion of antimicrobial nonsusceptible serotypes. Despite researchers' efforts to contact all available microbiological laboratories in Cambodia, the study size remained small, with a limited number of invasive isolates. None of the public microbiology laboratories store cultured isolates. We were, however, able to identify isolates from patients with severe pneumococcal infections from five different geographical regions in Cambodia, prospectively and systematically collected as part of well-conducted epidemiological studies on lower and/or severe respiratory infections. We, therefore, are confident that our results may be still generalizable to the general population. Furthermore, results were very similar to the two previously published studies from Cambodia, despite

differences in the targeted age group and geographical areas. Importantly, the present study is the first to include adult data.

It has clearly been demonstrated in high-income countries that the incidence of invasive pneumococcal disease due to serotypes included in PCV significantly decreased with the introduction of the vaccine into the childhood vaccination schedule due to a direct effect in vaccinated children as well as an indirect herd effect among older age groups.⁶ However, after the introduction of the vaccine, an increase of non-vaccine serotypes by either selection or replacement has been observed in many countries.³ If nonvaccine serotypes carry antibiotic resistance genes, serotype shifting may potentially lead to an increase of the prevalence of antibiotic-resistant pneumococcal clones.²⁰ For these reasons, pre-vaccination data are needed to monitor effects of the introduction of PCV. In low-income settings, data are scarce and accurate prediction is often hampered by low-quality epidemiological, biological, or clinical data. Despite the limited study size, our study based on isolates

TABLE 4
Antimicrobial susceptibility data of the isolates according to vaccine coverage of serotypes

	Overall N (%)	PCV13 isolates N (%)	Non-vaccine isolates N (%)	P value
Penicillin G*				0.02
Susceptible	95 (58)	50 (51)	45 (68)	
Amoxicillin†				0.31
Susceptible	127 (93)	68 (91)	59 (95)	
Ceftriaxone‡				0.10
Susceptible	149 (94)	87 (93)	62 (98)	
Cotrimoxazole§				0.005
Susceptible	15 (9)	4 (4)	11 (17)	
Erythromycin§				0.006
Susceptible	81 (49)	40 (40)	41 (62)	

PCV13 = 13-valent pneumococcal conjugate vaccine.

* Antibiotic susceptibility testing data were available for 165/166 isolates.

† 157/166 isolates tested.

‡ 164/166 isolates tested.

§ 165/166 isolates tested.

prospectively collected as part of either epidemiological studies or routine care, from patients with severe pneumococcal infections of all ages and from five geographical regions, provides a good baseline pre-vaccination assessment of the epidemiology of pneumococcal strains in Cambodia.

In conclusion, our study supports the potential for high reduction in pneumococcal disease burden with the introduction of PCV13 in childhood vaccination program. Multidrug resistance was higher among strains included in PCV13, which further supports likely vaccine impact of the now implemented vaccine program.

Received September 1, 2017. Accepted for publication December 2, 2017.

Published online January 8, 2018.

Acknowledgments: We acknowledge patients who agreed to participate in the studies. We also acknowledge the medical staff at the hospitals and microbiological laboratories for taking care of the patients, and for collecting and processing the samples.

Financial support: The SISEA study was funded through a grant from the Agence Française de Développement, Paris, France. The surveillance program at SHCH was funded through the Belgian Directorate of Development Cooperation. The studies at NAMRU-2 were funded by Global Emerging Infectious Surveillance section of the Armed Forces Health Surveillance Branch, work unit #D1310 and #D1409. The serotyping was supported by grants from Fondation Mérieux, the Swedish Government Funds for Clinical Research, and The Royal Physiographic Society in Lund. Serotyping work at COMRU was supported by Li Ka Shing University of Oxford Global Health Programme (Grant No. LG25 to P. T.) and by the Wellcome Trust as part of the Wellcome Trust-Mahidol University-Oxford Tropical Medicine Research Program. The funders played no role in the design of the study, data collection or analysis, decision to publish, or preparation of the manuscript.

Disclaimer: The opinions and assertions contained herein are those of the authors and are not to be construed as official or reflecting the views of the Department of the Navy, Department of Defense, or the U.S. Government. C. F., C. Y., M. R. K. G. F., and S. W. N. are a military service members. This work was prepared as part of their official duties. Title 17 U.S.C. §105 provides that "Copyright protection under this title is not available for any work of the United States Government." Title 17 U.S.C. §101 defines a U.S. Government work as a work prepared by a military service member or employee of the U.S. government as part of that person's official duties. The study protocol was approved by the Naval Medical Research Center Institutional Review Board (Protocol NMRCA 2016.0003-0009) in compliance with all applicable Federal regulations governing the protection of human subjects.

Authors' addresses: Malin Inghammar, Section for Infection Medicine, Department of Clinical Sciences, Lunds Universitet, Skane University Hospital, Lund, Sweden, E-mail: malin.inghammar@med.lu.se. Youlet By, Vonthanak Saphonn, and Chanleakhena Phoeung, University of Health Science, Phnom Penh, Cambodia, E-mails: youlet.by@fondation-merieux.org, vonthanak@uhs.edu.kh, and p_chanleakhena@uhs.edu.kh. Christina Farris, Gavin Ford, and Steven W. Newell, Naval Medical Research Unit No. 2, Phnom Penh, Cambodia, E-mails: christina.farris@namru2.org.kh, gavinford@gmail.com, and steven.newell@nrl.navy.mil. Thong Phe, Sihanouk Hospital Center of Hope, Phnom Penh, Cambodia, E-mail: thongphe@sihosp.org. Laurence Borand, Sophie Goyet, Sirenda Vong, and Charles Mayaud, Epidemiology and Public Health, Institut Pasteur, Phnom Penh, Cambodia, E-mails: lborand@pasteur-kh.org, sgoyet@pasteur-kh.org, svong@pasteur-kh.org, and charlesmayaud@gmail.com. Alexandra Kerleguer, Department of Biology, Institut Pasteur, Phnom Penh, Cambodia, E-mail: akerleguer@pasteur-kh.org. Blandine Rammaert, UFR Medecine et Pharmacie, Service de Maladies Infectieuses et Tropicales, INSERM U1070, Université de Poitiers, Poitiers, France, E-mail: brammaert@yahoo.fr. Bertrand Guillard, Medical Laboratory, Institut Pasteur, Phnom Penh, Cambodia, E-mail: bguillard@pasteur-kh.org. Chadwick Yasuda, US Naval Research Laboratory, CBMSE, Washington, DC,

E-mail: chadwick.yasuda@nrl.navy.mil. Matthew R. Kasper, Naval Medical Research Unit No. 2, Phnom Penh, Cambodia, E-mail: matthew.l.kasper2.mil@mail.mil. Ung Sam An, Cambodian National Laboratory of Public Health, Phnom Penh, Cambodia, E-mail: usa@niph.org.kh. Buth Sokhal and Sok Touch, Cambodian Communicable Disease Control Department, Phnom Penh, Cambodia, E-mails: buthsokhal@gmail.com and touch358@online.com.kh. Paul Turner, Department of Microbiology, Cambodia Oxford Medical Research Unit, Angkor Hospital for Children, Siem Reap, Cambodia, and Centre for Tropical Medicine, Churchill Hospital, University of Oxford, Oxford, United Kingdom, E-mail: paul@tropmedres.ac. Jan Jacobs, Institute of Tropical Medicine, Clinical Sciences, Antwerp, Belgium, E-mail: jjacobs@itg.be. Méline Messaoudi and Florence Komurian-Pradel, Laboratoire des Pathogènes Émergents, Centre International de Recherche en Infectiologie, Lyon, France, E-mails: melina.messaoudi@fondation-merieux.org and florence.pradel@fondation-merieux.org. Arnaud Tarantola, Epidemiology and Public Health Unit, Institut Pasteur du Cambodge, Phnom Penh, Cambodia, E-mail: atarantola@pasteur-kh.org.

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

REFERENCES

- O'Brien KL, Wolfson LJ, Watt JP, Henkle E, Deloria-Knoll M, McCall N, Lee E, Mulholland K, Levine OS, Cherian T; Hib and Pneumococcal Global Burden of Disease Study Team, 2009. Burden of disease caused by *Streptococcus pneumoniae* in children younger than 5 years: global estimates. *Lancet* 374: 893–902.
- Brueggemann AB, Griffiths DT, Meats E, Peto T, Crook DW, Spratt BG, 2003. Clonal relationships between invasive and carriage *Streptococcus pneumoniae* and serotype- and clone-specific differences in invasive disease potential. *J Infect Dis* 187: 1424–1432.
- Browall S et al., 2014. Clinical manifestations of invasive pneumococcal disease by vaccine and non-vaccine types. *Eur Respir J* 44: 1646–1657.
- Harboe ZB, Benfield TL, Valentiner-Branth P, Hjuler T, Lambertsen L, Kaltoft M, Krogfelt K, Slotved HC, Christensen JJ, Konradsen HB, 2010. Temporal trends in invasive pneumococcal disease and pneumococcal serotypes over 7 decades. *Clin Infect Dis* 50: 329–337.
- Hausdorff WP, Feikin DR, Klugman KP, 2005. Epidemiological differences among pneumococcal serotypes. *Lancet Infect Dis* 5: 83–93.
- Feikin DR, Kagucia EW, Loo JD, Link-Gelles R, Puhon MA, Cherian T, Levine OS, Whitney CG, O'Brien KL, Moore MR; Serotype Replacement Study Group, 2013. Serotype-specific changes in invasive pneumococcal disease after pneumococcal conjugate vaccine introduction: a pooled analysis of multiple surveillance sites. *PLoS Med* 10: e1001517.
- Huss A, Scott P, Stuck AE, Trotter C, Egger M, 2009. Efficacy of pneumococcal vaccination in adults: a meta-analysis. *CMAJ* 180: 48–58.
- Johnson HL, Deloria-Knoll M, Levine OS, Stoszek SK, Freimanis Hance L, Reithinger R, Muenz LR, O'Brien KL, 2010. Systematic evaluation of serotypes causing invasive pneumococcal disease among children under five: the pneumococcal global serotype project. *PLoS Med* 7: pii: e1000348.
- GBD 2015 Mortality and Causes of Death Collaborators, 2016. Global, regional, and national life expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980–2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet* 388: 1459–1544.
- Gavi The Vaccine Alliance, 2017. *Country Hub: Cambodia*. Available at: <http://www.gavi.org/country/cambodia/>. Accessed June 2, 2017.
- Moore CE et al., 2016. Characterisation of invasive *Streptococcus pneumoniae* isolated from Cambodian children between 2007–2012. *PLoS One* 11: e0159358.

12. Turner P, Turner C, Suy K, Soeng S, Ly S, Miliya T, Goldblatt D, Day NP, 2015. Pneumococcal infection among children before introduction of 13-valent pneumococcal conjugate vaccine, Cambodia. *Emerg Infect Dis* 21: 2080–2083.
13. Vong S et al., 2013. Acute lower respiratory infections in ≥ 5 year-old hospitalized patients in Cambodia, a low-income tropical country: clinical characteristics and pathogenic etiology. *BMC Infect Dis* 13: 97.
14. Société Française de Microbiologie, 2015. *Comité de l'antibiogramme de la Société Française de Microbiologie Recommandations 2015*. Available at: http://www.sfm-microbiologie.org/UserFiles/files/casfm/CASFM_EUCAST_V1_2015.pdf. Accessed June 11, 2017.
15. Messaoudi M et al., 2016. The relevance of a novel quantitative assay to detect up to 40 major *Streptococcus pneumoniae* serotypes directly in clinical nasopharyngeal and blood specimens. *PLoS One* 11: e0151428.
16. Jauneikaite E, Tocheva AS, Jefferies JM, Gladstone RA, Faust SN, Christodoulides M, Hibberd ML, Clarke SC, 2015. Current methods for capsular typing of *Streptococcus pneumoniae*. *J Microbiol Methods* 113: 41–49.
17. Turner P, Hinds J, Turner C, Jankhot A, Gould K, Bentley SD, Nosten F, Goldblatt D, 2011. Improved detection of nasopharyngeal cocolonization by multiple pneumococcal serotypes by use of latex agglutination or molecular serotyping by microarray. *J Clin Microbiol* 49: 1784–1789.
18. Jauneikaite E, Jefferies JM, Hibberd ML, Clarke SC, 2012. Prevalence of *Streptococcus pneumoniae* serotypes causing invasive and non-invasive disease in South East Asia: a review. *Vaccine* 30: 3503–3514.
19. Brueggemann AB, Peto TE, Crook DW, Butler JC, Kristinsson KG, Spratt BG, 2004. Temporal and geographic stability of the serogroup-specific invasive disease potential of *Streptococcus pneumoniae* in children. *J Infect Dis* 190: 1203–1211.
20. Darenberg J, Henriques Normark B, 2009. The epidemiology of pneumococcal infections—the Swedish experience. *Vaccine* 27 (Suppl 6): G27–G32.