1 **Global phylogeography and evolutionary history of *Shigella dysenteriae* type 1**

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ABSTRACT

Together with plague, small-pox and typhus, epidemics of dysentery have been a major scourge of human populations for centuries¹. A previous genomic study concluded that *Shigella dysenteriae* type 1 (Sd1), the epidemic dysentery bacillus, emerged and spread worldwide after World War (WW) I, with no clear pattern of transmission². This is not consistent with the massive cyclic dysentery epidemics reported in Europe during the 18th and 19th centuries^{1,3,4} and the first isolation of Sd1 in Japan in 1897⁵. We report here a whole-genome analysis of 331 Sd1 isolates from around the world, collected between 1915 and 2011, providing us with unprecedented insight into the historical spread of this pathogen. We show here that Sd1 has existed since at least the 18th century, and that it swept the globe at the end of the 19th century, diversifying into distinct lineages associated with WWI, WWII, and various conflicts or natural disasters across Africa, Asia, and Central America. We also provide a unique historical perspective on the evolution of antibiotic resistance over a 100-year period, beginning decades before the antibiotic era, and identify a prevalent multiple antibiotic-resistant lineage in South Asia that was transmitted in several waves to Africa, where it caused severe outbreaks of disease.

TEXT

January 2016 marks one hundred years since the invasion force from Britain, Australia, New Zealand and France withdrew from the Dardanelles, in the then Ottoman Empire, only eight months after landing. Most of the more than 120,000 casualties evacuated from the Gallipoli Peninsula were suffering from epidemic bacillary dysentery⁶, caused by *Shigella dysenteriae* type 1^{7,8} (Sd1), a bacterium producing the powerful Shiga toxin. This human-adapted clone of *Escherichia coli*⁹ was isolated for the first time by Kiyoshi Shiga during a dysentery outbreak in Japan, during which 90,000 cases and 20,000 deaths occurred in the last six months of 1897 alone⁵. In the second half of the 20th century, large outbreaks of disease due to Sd1 were still being reported in Central America, with estimates of more than 500,000 cases and 20,000 deaths for the 1969-1973 epidemic^{10,11}, Africa, where there were an estimated 100,000 cases and 5-10,000 deaths in the 1979 epidemic¹², and Asia^{13,14}.

Very little is known about the origins, evolution and spread of this important human pathogen, including, in particular, the strains involved in the major outbreaks and the genetic relationships between them. We carried out a whole-genome sequence analysis on a set of Sd1 isolates selected from more than 35 international strain collections, to represent the widest possible temporal and geographic distribution of available isolates, to obtain a phylogenetic framework that was robust over time and space and to infer transmission dynamics. This unique collection included 325 isolates from 66 countries spanning four continents, collected between 1915 and 2011. Sixty-seven historical isolates collected between 1915 and 1960, including 14 isolates obtained

during World War I (WWI)^{15,16}, were included in the collection, together with several isolates from each major outbreak reported since the 1960s. Short-read sequences from six Sd1 published genomes² were also included, with *S. flexneri*, *S. boydii*, *S. sonnei* and *Escherichia coli* genomes used as outgroups.

Single-nucleotide polymorphisms (SNPs) were detected by mapping short-read sequences against Sd1 reference genomes: Sd197¹⁷, which was isolated during an outbreak in China in the 1950s, and Sd1617¹⁸, which was isolated in Guatemala during the 1968-1969 epidemic. Maximum likelihood (ML) phylogenetic analysis was performed on 14,677 (mapping against Sd197) and 15,752 (mapping against Sd1617) chromosomal SNPs, which were randomly distributed over the non-repetitive non-recombinant core genome (85.6% of the Sd197 chromosome, Supplementary Information). Four genetic lineages (Fig. 1a, Supplementary Information) were identified. Lineage I contained only M115, which was isolated from a case in England in 1926. Lineage II contained mostly isolates collected in Europe between 1915 and 1958. Lineage III contained isolates from around the world and could be split into four sublineages with strong geographical affinities: IIIa in eastern and southeastern Asia (with isolates collected between 1927 and 1971), IIIb in Central America (1955-1992), IIIc in West Africa (1954-2006), and IIId in southern Asia and eastern Africa (1956-1977) and then in West Africa (1979-1998). Finally, lineage IV contained most of the Sd1 isolates obtained from the Indian subcontinent and Africa in the last few decades.

Ten of the 14 isolates (71%) amassed by Captain E.G.D. Murray during WWI belonged to the European lineage, lineage II, and most were isolated at the 2nd Western General Hospital, Manchester, which received many of the soldiers evacuated during the

Gallipoli campaign (Supplementary Fig. 1). The other four isolates belonged to three of the four sublineages of the global lineage, lineage III. None of the WWI isolates belonged to sublineage IIIId, which gave rise to the modern lineage, lineage IV.

The two candidate vaccine strains developed to date are derived from lineage III parental isolates (IIIb for parental strain Sd1617 of vaccine strain WRSd1¹⁹ and IIIId for parental strain 7-87 of vaccine strain SC-599²⁰).

ML phylogenetic analysis revealed a strong correlation between root-to-tip branch lengths and the known years of isolation for the sequenced Sd1 isolates, indicating a clock-like evolution (Supplementary Fig. 2). We therefore used a Bayesian phylogenetic approach to provide estimates of the nucleotide substitution rates and divergence times of the different lineages for a spatially and temporally representative subset of 125 isolates (Fig. 2). We estimated the genome-wide substitution rate at 8.7×10^{-7} substitutions site⁻¹ year⁻¹ [95% credible interval (CI) = $7.6 \times 10^{-7} - 9.9 \times 10^{-7}$], giving a most recent common ancestor (MRCA) for all the Sd1 in our collection dating from 1747 (95% CI, 1645 - 1822). This finding is consistent with historical data from the 18th to mid-19th centuries, describing cyclic dysentery epidemics in Western and Northern Europe associated with extraordinarily high mortality rates. For example, the 1738-1742 and 1779 epidemics in France killed more than 200,000 people¹, the 1770-1775 epidemic in Sweden killed almost 35,000 people (12% of all deaths during the period)³, and a large number of deaths from dysentery were also reported during the Irish Great Famine of 1846-1849⁴. The MRCA for all isolates other than M115 was dated to the mid-19th century (1853; 95% CI 1831-1871), whereas the MRCAs for each of the sublineages of global lineage

III were estimated to have existed between 1889 (95% CI 1881-1897) and 1903 (95% CI 1893-1913), indicating that this lineage spread worldwide over a period of less than two decades. This dating is also consistent with Shiga's observation that the dysentery outbreak of 1897 had begun in the late 1880s in the southern part of Japan²¹.

Our findings show that the global spread of Sd1 predates WWI. It therefore occurred earlier than for another *Shigella* serogroup, *S. sonnei*, which has been shown to have spread to other continents from Europe during the second half of the 20th century²². We cannot demonstrate causality between the spread of Sd1 and historical events on the basis of the results presented here, but the late 1800s coincided with a period of intense European emigration, the colonisation of various territories in Africa and Asia by European powers, facilitated by the opening of the Suez canal (1869) and the development of steamships.

Geographic and temporal analyses identified several intercontinental transmission events resulting in long-term establishment of the bacterium (Figs 1b, 1c, and 2). Transmission event T1 involved the European lineage II and led to an introduction of Sd1 in Madagascar between 1915 (95% CI 1910-1921) and 1967 (95% CI 1956-1977), during French colonization. This is consistent with the first report, which unambiguously described Sd1 there in 1927²³. Transmission event T2, involving eastern Asia and Poland, is estimated to have occurred between 1910 (95% CI 1899-1925) and 1944 (95% CI 1942-1945). All other transmission waves originated in the Indian subcontinent and affected mostly East Africa. Two of these transmission waves, T5 and T8, led to major outbreaks; according to our estimates, T5 occurred between 1970 (95% CI 1963-1975)

and 1979 (95% CI 1976-1981). This dating is consistent with the first reported outbreak in the northeastern part of what is now the Democratic Republic of the Congo in 1979, 28 years after the last isolation of Sd1 in Central Africa¹². This epidemic then spread to the Great Lakes region, where it persisted until at least 1990¹². T8 occurred between 1984 (95% CI 1978-1987) and 1987 (95% CI 1985-1989), with a first reported outbreak in Zambia in 1990-1991^{12,24}. The strain then rapidly spread across an Africa ravaged by civil unrest, war (e.g., Mozambique, Angola, Rwanda, Sierra Leone) and HIV infection^{12,24} until 2011. With the exception of a localized outbreak in the northern part of the Central African Republic in 2004²⁵ caused by sublineage IIIc (see below), all other outbreaks in Africa since 1990 have been caused by lineage IV.

The high resolution of whole-genome sequence analysis (WGS) has significantly changed our understanding of the patterns of Sd1 transmission over time at a global scale. The classical molecular epidemiology tools (Supplementary Information) previously used were unable to unravel these patterns of transmission. Furthermore, a re-evaluation of two outbreaks that occurred in the Central African Republic in 2003-2004²⁵ that we had previously investigated by pulsed-field gel electrophoresis (PFGE), the current method of choice for subtyping Sd1, revealed a lack of correlation between PFGE and WGS data (Supplementary Fig. 3 and Supplementary Information). In particular, PFGE grouped the isolates from the two outbreaks closely together, whereas they actually belonged to two different lineages, IIIc and IV, separated by ~700 SNPs. By contrast, other African T8 lineage IV isolates differing by 37 to 61 SNPs from the Central African Republic T8 lineage IV outbreak isolates, formed a more distant group. Thus, PFGE cannot attribute

profiles from different apparently geographically restricted outbreaks to a single, longer epidemic, such as that associated with the T8 transmission wave in Africa. PFGE should, therefore, no longer be used for the assessment of phylogenetic relationships in Sd1. Instead, WGS provides a robust phylogenetic framework for the epidemiological tracking of this bacterium.

One key feature in the evolution of Sd1 is the acquisition and accumulation of antibiotic resistance genes (ARGs) (Figs 3, 4, Supplementary Fig. 4, and Supplementary Information). The first antibiotic-resistant Sd1 isolates in our collection were recovered in Asia and America during the 1960s and rapidly became predominant, such that susceptible isolates had become exceptional by 1991 (100%, [67/67] susceptible isolates, between 1915 and 1960 and <1% [1/123], between 1991 and 2011). Lineage IV, the most recent of the lineages identified, is the most affected by antibiotic resistance, but almost all the contemporary circulating strains from older lineages have also become resistant to multiple antibiotics. ARGs were acquired following the first use of antibiotics in clinical practice (Fig. 4b). The first ARGs identified in Sd1 were borne on small plasmids (<10 kb), encoding resistance to streptomycin and sulfonamides. Larger plasmids (80-130 kb) of different types encoding additional resistance to tetracycline, chloramphenicol, and, for some plasmids, ampicillin (via the *bla*_{OXA-1} or *bla*_{TEM-1} genes) were then acquired in various geographic areas, from the mid-1960s to the 1980s. These plasmids belonged to the IncK and IncF groups in Asia and to the IncB/O group in Central America. The use of cotrimoxazole, beginning in the late 1960s, led to the acquisition of dihydrofolate reductase genes, mostly *dfrA1*, carried by 110-kb pST186 IncI1 and 30-kb IncX4

273 plasmids or by the Tn7 transposon inserted into the Sd1 chromosome close to the *glmS*
274 gene, as observed for *S. sonnei*²². Since the 1990s, the principal structure associated with
275 multidrug resistance in Sd1 has been a 66-kb genomic element called the *Shigella*
276 resistance locus pathogenicity island (SRL-PAI)²⁶. It was acquired four times in lineage
277 IV (South Asia or the Middle East), once in sublineage IIIc (West Africa), and once in
278 lineage II (Madagascar). Further evidence for the independent acquisition of the SRL-
279 PAI is provided by the presence of slight differences between the different acquired SRL-
280 PAIs (Supplementary Fig. 5). The SRL-A is very similar to the first SRL-PAI to be
281 described in *S. flexneri*²⁶ and it was found exclusively in lineage IV. The SRL-B, found
282 only in the lineage IV African T8 isolates, was probably derived from the SRL-A by
283 insertion sequence (IS) ISSdI-mediated rearrangements rather than being independently
284 acquired. The other SRL-PAI contained various insertions (group II introns, part of the
285 *shf* operon, region replacing orf47) not present in SRL-A. Among the 149 isolates
286 bearing the SRL-PAI, only two showed a partial deletion of the SRL-PAI, resulting in a
287 loss of the antibiotic resistance cluster (i.e., the SRL *sensu stricto*). This structure is
288 therefore quite stable over time, particularly in a bacterium containing hundreds of
289 ISs^{17,18}. This 66-kb element encodes resistance to ampicillin, streptomycin,
290 chloramphenicol and tetracycline, with no more resistance than the previously circulating
291 large plasmids. Its persistence may therefore be associated with a lower fitness cost and
292 the presence of an *fec* operon for the capture of iron, serving as selective advantages²⁶.
293 Before the principal acquisition of the SRL-A, the closest ancestral group (consisting
294 initially of South Asian and then South-East and Central Asian isolates), had acquired a
295 chromosomally encoded transposon (Fig. 2, Supplementary Fig. 6). This 10-kb structure

encodes resistance to chloramphenicol and tetracycline. The structure of the double drug-resistance module is similar to that found in the SRL and to some previously circulating large multidrug resistance IncF plasmids, such as p3099-85 and p80-547. This recent trend towards acquiring ARG-containing genomic islands or chromosomally-encoded transposons rather than plasmids is also displayed by the 7th pandemic *V. cholerae* (SXT/R391) and *Salmonella enterica* serotype Typhi H58 (24-kb composite transposon) strains, which also originate from the Indian subcontinent^{27,28}.

Resistance to nalidixic acid, a quinolone, mediated by point mutations in the DNA gyrase gene, *gyrA*, was acquired seven times in lineage IV Sd1 isolates from South Asia and Africa (Fig. 2) from the 1980s. The *gyrA* mutation leading to a serine-to-leucine substitution in the amino-acid sequence, S83L was the most frequently observed, but others, involving codon 87, such as D87G and D87Y, were observed in isolates from Central Africa and Thailand, respectively, during the 1990s. Interestingly, in the same geographic area of DRC and Rwanda in 1994, two different mutations were acquired (S83L and D87G). This may reflect the heavy use of nalidixic acid in the Rwandan refugee camps, which experienced outbreaks of disease caused by *Vibrio cholerae* O1 and Sd1²⁹.

Resistance to ciprofloxacin, a fluoroquinolone, mediated by a double mutation in *gyrA* (S83L and a second mutation in codon 87) and a mutation in the topoisomerase IV *parC* gene (S80I) was acquired only once, in a group of 20 isolates from the Indian subcontinent collected between 1995 and 2010 (Fig. 2). We observed no resistance to extended-spectrum cephalosporins, carbapenems or azithromycin in the isolates studied here, but the existence of such resistance is almost inevitable, as the area of circulation of

Sd1 overlaps with that of Enterobacteriaceae possessing mobile ARGs encoding resistance to the latest generation of antibiotics, such as NDM-1³⁰. However, the dramatic decrease in Sd1 isolation reported since the turn of the century and not explained by the findings of this genomic study, may counterbalance these pessimistic predictions.

METHODS

Bacterial isolates

The Sd1 isolates analysed in this study are listed in Supplementary Table 1 and originated from the collections of the Centers for Disease Control and Prevention, Atlanta, GA, USA ($n=56$); Institut Pasteur, Paris, France ($n=53$); Public Health England, Colindale, UK ($n=29$); Icddr,b, Dhaka, Bangladesh ($n=29$); Central Research Institute for Epidemiology, Moscow, Russian Federation ($n=22$); National Institute of Public Health, Prague, Czech Republic ($n=19$); Public Health England, Porton Down, UK ($n=17$); Iris-Lab, Brussels, Belgium ($n=11$); National Institute of Cholera and Enteric Diseases, Kolkata, India ($n=8$); Institut Pasteur de Bangui, Bangui, Central African Republic ($n=7$); Norwegian Institute of Public Health, Oslo, Norway ($n=6$); Hungarian National Collection of Medical Bacteria, Budapest, Hungary ($n=6$); Pasteur Institute of St Petersburg, St Petersburg, Russian Federation ($n=5$); National Institute of Public Health, Warsaw, Poland ($n=5$); Institut Pasteur de Dakar, Dakar, Senegal ($n=4$); New York University Langone Medical Center, New York, USA ($n=4$); Robert Koch Institut, Wernigerode, Germany ($n=4$); Institut für Hygiene und Umwelt, Hamburg, Germany

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(*n*=1); Istituto Pasteur-Fondazione Cenci Bolognetti, Rome, Italy (*n*=1); Félix d'Hérelle
reference center for bacterial viruses, Université Laval, Québec, Canada (*n*=1); National
Institute for Communicable Disease Control and Prevention, Beijing, China (*n*=1).

Bacterial DNA samples were also received from the Armed Forces Research Institute of
Medical Sciences, Bangkok, Thailand (*n*=10).

The 18 Sd1 isolates from the E.G.D. Murray collection^{15,16,31} included 14 isolates
recovered during WWI and four isolates obtained between 1926 and 1930. The WWI
isolates were obtained from different sources (Supplementary Fig. 1) and were stored at
room temperature in Douglas digest agar slant glass tubes after sealing with a gas-air
burner between August 1918 and October 1919. In 1980, the 18 tubes and the 680 other
cultures of Enterobacteriaceae from the entire collection were shipped to the National
Collection of Type Cultures (NCTC), Porton Down, UK, opened and freeze-dried.

It was confirmed that all the isolates included belonged to Sd1, by conventional methods and serotyping at the French National Reference Center for *E. coli*, *Shigella* and *Salmonella*, Institut Pasteur, Paris, as previously described³².

Antibiotic susceptibility testing

Antibiotic susceptibility was determined by disk diffusion on Mueller-Hinton (MH) agar in accordance with the guidelines of the Antibiogram Committee of the French Society for Microbiology (CA-SFM 2014) (www.sfm-microbiologie.org/). The following antimicrobial drugs (Bio-Rad, Marnes-la-Coquette, France) were tested: amoxicillin, ceftriaxone, ceftazidime, streptomycin, kanamycin, amikacin, gentamicin, nalidixic acid, ofloxacin, ciprofloxacin, sulfonamides, trimethoprim, sulfamethoxazole-trimethoprim, chloramphenicol, tetracycline, and azithromycin. *Escherichia coli* CIP 76.24 (ATCC 25922) was used as a control. For strains displaying resistance to either nalidixic acid or ciprofloxacin by the disk diffusion method, this resistance was confirmed by determination of the minimal inhibitory concentration (MIC) with the corresponding Etest strips (bioMérieux, Marcy L'Etoile, France). The MICs of azithromycin and nitrofurantoin were determined by Etests for 30 isolates chosen on the basis of resistance phenotype, and year and country of isolation.

Determination of the mutator phenotype of strain M115

The mutation rate of M115 was estimated by monitoring the capacity of this strain to generate mutations conferring resistance to rifampin in two independent experiments including duplicates, as previously described³³. *E. coli* strain ECOR48 (CIP 106023) was used as a strong mutator positive control³⁴, the Sd1 97-13397 isolate was used as a putative strong mutator isolate (deletion of the *mutS* gene), Sd1 M116 and Sd197 were used as putative normomutator isolates (integrity of the *mutS*, *mutH*, *mutL* and *uvrD* methyl-directed mismatch repair genes).

Total DNA extraction

Total DNA was extracted with the InstaGene matrix kit (Bio-Rad) for the PCR identification of antibiotic resistance genes, the Wizard Genomic DNA Kit (Promega, Madison, WI, USA) for multilocus sequence typing and Illumina sequencing and the phenol chloroform method³⁵ for Illumina sequencing and PacBio sequencing.

Multi-locus sequence typing

Conventional multi-locus sequence typing (MLST) was performed on a subset of 33 Sd1 isolates, as previously described³⁶. Sequencing was performed at the *Plateforme de Génomique des Pathogènes et Santé Publique*, PF8 (Institut Pasteur). The nucleotide sequences and deduced protein sequences were analysed with EditSeq and Megalign software (DNASTAR, Madison, WI, USA). The BLASTN program of NCBI was used for database searches (<http://www.ncbi.nlm.nih.gov/BLAST/>).

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410 **PCR identification of antibiotic resistance genes**

411

412 The *bla*_{TEM}, *bla*_{SHV}, *bla*_{OXA-1}, *cat1*, *sul1*, *dfrA1*, and *aadA1* resistance genes and the class
413 1 and 2 integron gene cassettes were amplified by PCR, as previously described³⁷.

414

415 The presence of the *Shigella* resistance locus pathogenicity island (SRL-PAI) was
416 assessed by PCR, as previously described³⁸. The structure of the SRL-PAI was assessed
417 by PCR mapping with the primers described or with new primers designed on the basis of
418 GenBank accession no. AF326777. Amplicons not of the expected size were sequenced.

419

420 **Plasmid analyses**

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422 Plasmids were obtained from *E. coli* transconjugants or transformants, as previously
423 described³⁷, except that ampicillin (50 mg/L) or chloramphenicol (20 mg/L) was used as
424 a selective agent.

425

426 Plasmid size was determined in parental and transconjugant or transformants strains by
427 S1 nuclease treatment and pulsed-field gel electrophoresis, as previously described³⁷.

428 PCR-based replicon-typing analysis was performed as previously described³⁹.

429

430 Eight 30-130 kb plasmids conferring antimicrobial resistance were sequenced. Plasmid
431 DNA was extracted with the Large-Construct Kit (Qiagen, Courtaboeuf, France) and

sequenced through services provided by GATC Biotech (Konstanz, Germany), using
shotgun sequencing runs on a 454/Roche GS FLX Analyzer (Roche, Basel, Switzerland).
The resulting sequences were assembled into a unique scaffold. Gap closure was carried
out by PCR followed by Sanger DNA sequencing with the Big Dye® Terminator V3.1
Cycle Sequencing Kit (Applied Biosystems, Foster City, CA, USA) and a 96-capillary
3730xl DNA Analyzer (Applied Biosystems), by Eurofins MGW Operon (Cochin
Platform, Paris, France). Automatic annotation was performed with the RAST⁴⁰
server (<http://rast.nmpdr.org/>), followed by manual inspection and correction. The
sequences obtained have been deposited in GenBank under the accession numbers
KT754160 (p80-547), KT754161 (pCAR10), KT754162 (pBU53M1), KT754163
(pA5468), KT754164 (p3099-85), KT754165 (p93-531-1), KT754166 (p92-9000),
KT754167 (p69-3818).

Whole-genome sequencing

High-throughput genome sequencing was carried out at the genomics platform of the
Pasteur Institute, GATC Biotech, Beckman Coulter Genomics (Danvers, MA, USA) or at
the Wellcome Trust Sanger Institute, on Illumina platforms generating 100 to 146 bp
paired-end reads, yielding a mean of 206-fold coverage (minimum 37-fold, maximum
990-fold) (Supplementary Table 2). Short-read sequence data were submitted to the
European Nucleotide Archive (ENA) (<http://www.ebi.ac.uk/ena>) and the genome
accession numbers are provided in Supplementary Table 1.

We optimised the resolution of the chromosome-encoded antibiotic resistance structures and ensured that representative isolates from the various lineages were included, by sequencing 10 isolates on the PacBIO RS II platform (Pacific Biosciences, CA, USA), as previously described²⁸. The PacBio data were submitted to the ENA and the genome accession numbers are provided in Supplementary Table 1.

Other studied genomes

Sd1 strain Sd197¹⁷ was used as the reference genome. A second Sd1 genome Sd1617¹⁸ was used as a second reference genome, to confirm the population structure found with Sd197.

Short-read sequences from the following six Sd1 genomes published by Rohmer *et al.*² were downloaded from the ENA and included in this study: 2735 (USA, 1974, SRR765065), 91R17 (Guatemala, 1991, SRR765098), 91R14 (Guatemala, 1991, SRR765104), DH03 (Central African Republic, 1996, SRR765110), DH05 (Central African Republic, 1996, SRR765112), and DH06 (Central African Republic, 1996, SRR765113).

The following genomes were used as outgroups: *E. coli* O157:H7 strain Sakai (GenBank accession no. NC_002695), *E. coli* strain K-12 MG1655 (GenBank accession no. NC_000913), *S. flexneri* type 2a strain 2457T (GenBank accession no. AE014073), *S.*

boydii strain Sb227 (GenBank accession no. NC_007613), and *S. sonnei* strain Ss046 (GenBank accession no. NC_007384).

Read alignment and SNP detection

For the analysis of single-nucleotide polymorphisms (SNPs), Illumina-generated paired-end reads and the simulated paired-end reads from publicly available assembled genomes, were mapped to the reference genome of Sd1 strain Sd197, including the chromosome (CP000034) and plasmids pSD1_197 (CP000035) and pSD197_spA (CP000640), with SMALT (version 0.7.4) (<http://www.sanger.ac.uk/resources/software/smalt/>) as previously described²⁸.

***De novo* assembly**

The reads for each strain were assembled *de novo* with Velvet⁴¹ version 1.2.09, with parameters optimised with VelvetOptimiser version 2.2.5 (<https://github.com/tseemann/VelvetOptimiser>). They were scaffolded with SSPACE⁴² version v2.0. The gaps were closed with GapFiller⁴³ version 1.11, and the sequences were annotated with Prokka⁴⁴ version 1.5, as previously described²⁸. CLC Assembly Cell version 4.2.0 (CLC bio, Aarhus, Denmark) was also used to investigate antibiotic resistance determinants.

Phylogenetic analyses

500

501 The maximum likelihood (ML) phylogenetic tree shown in Supplementary Fig. 7 was
502 built from a 140,385-chromosomal SNP alignment generated by snp_sites software
503 (https://github.com/sanger-pathogens/snp_sites) from all 331 short-read sequences, plus
504 Sd1 genomes Sd197 (used as a reference) and Sd1617, together with the six *E. coli* and
505 *Shigella* sp. genomes used as outgroups. RAxML⁴⁵ version 7.8.6 was used with the
506 generalised time-reversible model and a Gamma distribution to model site-specific rate
507 variation (the GTR+ Γ substitution model; GTRGAMMA in RAxML). Support for the
508 ML phylogeny was assessed by 100 bootstrap pseudo-analyses of the alignment data, and
509 the final tree was visualised in FigTree version 1.4.2
510 (<http://tree.bio.ed.ac.uk/software/figtree/>).

511

512 The ML phylogenetic trees shown in Figs 1a, 3a, 3c, Supplementary Figs 1a, 3b, 4, 9, 11
513 and 14 were built from a 14,677-chromosomal SNP alignment of all 331 Sd1 short-read
514 sequences, plus Sd1 genome Sd197, used as the reference. Repetitive regions (within the
515 chromosome, between the chromosome and the virulence plasmid (VP) or the SRL-PAI)
516 were removed manually with the Artemis⁴⁶ genome browser. Recombinogenic regions
517 were also removed with the Gubbins⁴⁷ software. The remaining 14,677 chromosomal
518 SNPs were randomly distributed along the non-repetitive non-recombinant core genome
519 (3,750,125 bp), with a spacing of about one SNP per 256 bp or a nucleotide divergence of
520 0.39% (Supplementary Fig. 12). RAxML version 7.8.6 (GTRGAMMA substitution
521 model) was used to construct the tree. We performed 500 bootstrap pseudoreplicate
522 analyses to assess support for the ML phylogeny. The tree was rooted on M115, which

was shown to be the most closely related to the ancestral strain of Sd1 by two different approaches (ML and Bayesian) and was visualised with MEGA⁴⁸ version 6, iTOL^{49,50} or FigTree version 1.4.2.

The ML phylogenetic trees shown in Supplementary Figs 10 and 11 were built from a 15,752-chromosomal SNP alignment of all 331 Sd1 short-read sequences, plus Sd1 genome Sd1617, used as the reference. The method used was similar to that described above, except that the repetitive regions were not removed manually and phylogenetic support was assessed by 100 bootstrap pseudo-analyses.

The VP phylogenetic tree shown in Supplementary Fig. 15 was constructed similarly, from the 226 plasmid-containing isolates (> 90% coverage at read depth > 10x), based on 290 SNPs randomly distributed along the non-repetitive non-recombinant pSD1_197 sequence (99,704 bp, 54.6% of pSD1_197). The tree was unrooted.

Phylogenetic clustering

We clustered the isolates of Sd1 into various lineages by eye and by applying hierarchical Bayesian analysis of population structure (BAPS)⁵¹ software to the 14,677-chromosomal SNP alignment. Five iterations (L value) were run with a maximum cluster number (K value) of 6 or 10 and three iterations were run with $K=6$.

Temporal analysis

546

547 We investigated the temporal signal in the ML phylogeny for Sd1, using Path-O-Gen
548 (<http://tree.bio.ed.ac.uk/software/pathogen/>). The relationships between root-to-tip
549 distances, year of isolation and lineage were analysed by linear regression methods.

550

551 We used Bayesian Evolutionary Analysis by Sampling Trees (BEAST)⁵² version 1.8 to
552 date the important nodes. The analyses were conducted on a subsample of 125 isolates
553 from across the ML tree, covering the full temporal and geographic range of this
554 pathogen. The concatenated 10,798 chromosomal SNP alignments of these 125 strains
555 were subjected to multiple BEAST analyses with both constant-size and Bayesian skyline
556 population size change models, in combination with either a strict molecular clock or a
557 relaxed clock, to identify the best-fit model^{22,53}. For the BEAST analysis, the GTR+ Γ
558 substitution model was selected and tip dates were defined as the year of isolation. For all
559 model combinations, three independent chains of 100 million generations each were run
560 to ensure convergence, with sampling every 1,000 iterations. Convergence and effective
561 sample size (ESS) values were inspected using Tracer⁵² version 1.5. A marginal
562 likelihood estimation was carried out, with path sampling and stepping stone sampling
563 for each run that had converged, to compare the different combinations of clock and tree
564 models^{54,55}. The marginal likelihood estimation was then used to determine which model
565 gave the best fit, by calculating the Bayes Factor. The relaxed, uncorrelated lognormal
566 clock model, which allows evolutionary rates to vary among the branches of the tree
567 together with the skyline demographic model proved a much better fit for the data, as
568 found previously for *S. sonnei*²² and *S. flexneri*⁵³. The parameter and tree estimates of the

three runs were combined with LogCombiner⁵² version 1.7.5, with the first 20% of states in each chain removed as burn-in. Maximum clade credibility (MCC) trees were generated with TreeAnnotator⁵² version 1.7.5 on the combined files, and visualised with FigTree version 1.4.2. Estimates are reported as median values with the 95% highest posterior density (HPD, hereafter referred to as the credible interval). The Bayesian skyline plot was calculated and visualised with Tracer⁵² version 1.5, to investigate changes in the effective population size of Sd1 over time. To confirm the dating estimates, ten other random subsamples were generated from clusters calculated using the Prosperi method⁵⁶ (code here: http://figshare.com/articles/clustertree.R_Code_for_clustering_phylogenetic_trees/97225) with a threshold of 0.03. All singleton isolates were included (n=86) and one isolate from each of the 33 clusters was randomly selected to generate the ten subsamples. These alignments were analysed in BEAST using the same model and showed similar dating for each of the lineages (Supplementary Table 2).

Genetic analyses

In silico MLST was then carried out by MLST version 1.8 (<https://cge.cbs.dtu.dk/services/MLST/>) on assembled sequences for all the dataset. New alleles were confirmed by Sanger sequencing and submitted to the MLST database website (<http://mlst.warwick.ac.uk/mlst/>).

The presence and type of antibiotic resistance genes (ARGs) or ARG-containing structures (Fig. 3b and Supplementary Fig. 4) were determined with ResFinder⁵⁷ version 2.1 (<https://cge.cbs.dtu.dk/services/ResFinder/>), BLAST analysis against defined reference sequences (plasmids or chromosomally encoded structures), PlasmidFinder⁵⁸ version 1.3 (<https://cge.cbs.dtu.dk/services/PlasmidFinder/>), Plasmid MLST locus/sequence definitions database (<http://pubmlst.org/plasmid/>), and pMLST version 1.2 (<https://cge.cbs.dtu.dk/services/pMLST/>) on CLC or Velvet assemblies. The new alleles and STs of IncI⁵⁹ and IncN⁶⁰ plasmids have been deposited in the PubMLST database (<http://pubmlst.org/plasmid/>). The presence of mutations in the quinolone-resistance determining region of the DNA gyrase and topoisomerase IV genes was determined manually on *de novo* assembled sequences. PacBio sequences were used to analyse the structure of the SRL-PAI variants and the composite transposon inserted into the chromosome in genome CDC 87-3330. The *in silico* results were compared with PCR data, when available.

Pan-genome analysis

Roary⁶¹ version 3.2.4 was used on Velvet-annotated assemblies, to construct a pan-genome. The pan-genome analysis identified genome 2735² as an outlier. Further investigation revealed an extreme AT bias, therefore this sample was excluded from subsequent analyses. A more sensitive annotation was performed on the resulting clusters of proteins with InterPro⁶², to provide Gene Ontology⁶³ classifications for each gene.

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SUPPLEMENTARY INFORMATION

Supplementary Information is linked to the online version of the paper at www.nature.com/nature.

The authors declare no competing financial interests.

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AUTHOR CONTRIBUTIONS

R.B., P.A.D.G., S.B., N.R.T and F.-X.W. designed the study. N.S., C.J., K.A.T., R.B., K.K., R.K., J.E.R., L.K., A.K., O.V., C.J.M., T.R., C. Bizet, A.G.S, A.G., A.L.W., M.-C.F., S.L.H., M.J.B, C.J., A.M., A.-L.P., P.R., A.F., E.D., M.V., H.B., M.H., P.A.D.G., P.S., L.B., C.-S.C., D.C., B.C., S.E., G.P.P., A.V.E., H.I., A.K.-K., A.L., M.G., F.G., C.L., M.M., L.A.M.P, G.P.-P., A.P., G.A.S., D.T., C.Y., H.Z., P.S. and F.-X.W. selected and provided characterized isolates and their epidemiological information. E.N.-N., M.L.-C., I. C., C.R., A.T.-D., M. A.-D. and L.B. did the phenotypic experiments and DNA extractions. A.E.M. and S.R.H provided guidance for genomic analyses. C. Bouchier performed the whole-genome sequencing. M.A. processed the short reads. E.N.-N., N.F., K.K., S. B., K.E.H, J.H, A.J.P., G.G., E.S., and F.-X.W. analysed the genomic sequence

data. F.-X.W. wrote the manuscript with major contributions from A.E.M., P.A.D.G., E.D., J.P., P.S., K.E.H., S.B. and N.R.T. All authors contributed to manuscript editing. F.-X.W. oversaw the project.

AUTHOR INFORMATION

Short-read sequences have been deposited at EBI-ENA, under study accession numbers PRJEB10304, PRJEB2846 and PRJEB3255. PacBio sequences have been deposited at EBI-ENA, under study accession number PRJEB7928. Plasmid, SRL-PAI, and Tn87-3330 sequences have been deposited in GenBank, under accession numbers KT754160–KT754167, KT777637–KT777641, and KT777642, respectively.

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TABLE

None

FIGURE LEGENDS

Figure 1. Geographic distribution and transmission patterns of *Shigella dysenteriae* type 1 genetic lineages. a, Maximum likelihood (ML) phylogeny of the 332 genomes studied, showing the four lineages, I to IV, and the four sublineages of lineage III: IIIa to IIIc. The tree was rooted on M115, the most closely related to the *S. dysenteriae* type 1

ancestral strain. The tips of the tree are coloured to indicate the continent on which the infection occurred. T1 to T8 indicate intercontinental transmission events. **b**, Geographic presence (circles), inferred arrivals (thick arrows) and principal long-distance transmission events (thin arrows) of lineages I to III based on phylogeographic analysis. Intercontinental transmission events are indicated by the letter T. The date ranges shown for transmission events are the median values for the MRCA (taken from BEAST) with the first number indicating the median MRCA of the transmitted strains, and the second number indicating the median MRCA of the transmitted strains and their closest relative from the source location. **c**, Geographic presence (circles, thunderbolts) and intercontinental transmission events of lineage IV based on phylogeographic analysis. Isolate assignment to the corresponding transmission event is indicated by coloured halos.

Figure 2. Timed phylogeny of a subsample of 125 *Shigella dysenteriae* type 1 isolates.

a, Bayesian skyline plot showing temporal changes since 1747 in effective population size (black curve) with 95% confidence intervals (cyan). World War I (WWI) is indicated by a red bar. **b**, Maximum clade credibility tree produced using BEAST (lognormal relaxed clock model; Bayesian skyline) also presenting information about the ortho-nitrophenyl- β -galactoside (ONPG) test. Resistance to nalidixic acid (NAL^R) is indicated by a purple circle and resistance to ciprofloxacin (CIP^R) is indicated by a purple triangle. Acquisition of the antibiotic resistance element, *Shigella* resistance locus pathogenicity island (SRL-PAI), is indicated by a black thunderbolt. Acquisition of the resistance transposon (Tn87-3330), originally found in isolate CDC 87-3330, is indicated by an

orange thunderbolt. T1 to T8 indicate intercontinental transmission events. Estimated dates for the intercontinental transmission events are provided in dataset S7 of Supplementary Table 2.

Figure 3. Phenotypic and genetic characterization of antibiotic resistance in *Shigella*

dysenteriae type 1. a, Resistance phenotype for eight antibiotics (ampicillin, AMP; streptomycin, STR; sulfonamides, SUL; trimethoprim, TMP; chloramphenicol, CHL; tetracycline, TET; nalidixic acid, NAL; and ciprofloxacin, CIP), according to the lineages (I to IV) defined on the basis of the maximum likelihood (ML) phylogeny (as in Fig. 1a). Resistance is indicated in red and susceptibility in grey, whereas no antibiotic susceptibility data is indicated in white. **b,** Principal genetic structures bearing antibiotic resistance genes (ARGs) as a function of genetic lineage (defined by ML phylogeny), time period and geography. A more detailed figure is provided in Supplementary Fig. 4.

Figure 4. Evolution of antibiotic resistance of *Shigella dysenteriae* type 1. a, Change in the number of antibiotic resistance genes (ARGs) per isolate over time. The logarithmic trendline and the correlation coefficient of determination (R^2) are shown in red. **b,** Timeline of the first detection of the main ARGs in our collection. The antibiotics (AMP, ampicillin; STR, streptomycin; SUL, sulfonamides; TMP, trimethoprim; CHL, chloramphenicol; TET, tetracycline; NAL, nalidixic acid; and CIP, ciprofloxacin) for which the ARGs convey resistance to are indicated. Asteriks indicate the mutation of chromosomal genes of the core genome.

Figure 1.

Figure 2.

Figure 3.

a **b**

Figure 4.