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Immune responses during spontaneous control of HIV and AIDS: what is the hope for a cure?

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Abstract

HIV research has made rapid progress and led to remarkable achievements in recent decades, the most important of which are combination antiretroviral therapies (cART). However, in the absence of a vaccine, the pandemic continues, and additional strategies are needed. The "towards an HIV cure" initiative aims to eradicate HIV or at least bring about a lasting remission of infection during which the host can control viral replication in the absence of cART. Cases of spontaneous and treatment-induced control of infection offer substantial hope. Here, we describe the scientific knowledge that is lacking and the priorities that have been established for research into a cure. We discuss in detail the immunological lessons that can be learned by studying natural human and animal models of protection and spontaneous control of viremia or of disease progression. In particular, we describe the insight we have gained into the immune mechanisms of virus control, the impact of early virus-host interactions and why chronic inflammation, a hallmark of HIV infection, is an obstacle to a cure. Finally, we enumerate current interventions aimed towards improving the host immune response.

Key index words or phrases:

HIV controllers, post-treatment controllers, Natural hosts of SIV, eradication, inflammation

1. Introduction

Since the first human immunodeficiency virus (HIV) was isolated 30 years ago [1], remarkable progress has been made in research and drug development, but an efficient vaccine against HIV/AIDS is still not available. Multiple obstacles must be overcome, including the fact that HIV has a remarkable capacity to accumulate mutations and escape adaptive immune responses. During the viral life cycle, the genetic material of the virus is integrated into the cellular genome, which is believed to allow the virus to evade the host's immune responses. In this way, HIV can persist for months and years. Furthermore, HIV infection is characterised by the induction of immunological dysfunction and consequently, the host fails to control viral replication. Moreover, the preferential target cells of HIV are activated CD4+ T cells. Indeed, large quantities of virus particles are produced in activated CD4+ T cells, whereas resting CD4+ T cells are weakly or not permissive for HIV, and other CD4+ cells, such as macrophages, only produce small numbers of virions. HIV infection is characterised by a significant and persistent increase in activated CD4+ T cells. In other words, HIV creates and multiplies in its own target cells. Many open questions remain. For example, it remains a matter of debate whether a vaccine against AIDS should induce anti-HIV T cells or anti-HIV antibodies or both, which qualities anti-HIV T cells should possess to be efficient and how and where antibodies must be produced. In the absence of a vaccine, alternative strategies have become more important. Recent progress in HIV research has raised hopes for a cure for HIV. The "towards an HIV cure" initiative launched by the International AIDS Society has established a number of priorities with the aim of HIV eradication or at least lasting remission of infection during which the host can control viral replication in the absence of antiretroviral drugs (Figure 1) [2]. Timothy Brown, an HIVinfected patient who received a double stem cell transplant from CCR5 32 donors [3], has lived for more than 6 years without signs of the virus and represents the closest example to an HIV cure to date [4, 5]. However, achieving HIV eradication in a large population of patients seems farfetched at present. The natural models of AIDS control and the cases of patients able to control replication after treatment interruption encourage us to believe that HIV remission may be an achievable goal.

2. Spontaneous protection against HIV/AIDS

The capacity to control HIV replication and the speed of progression towards AIDS vary among patients. Approximately 10% of individuals infected with HIV-1 maintain their CD4+ T cell counts at near-normal levels for more than 7 years in the absence of anti-retroviral treatment. These individuals are called long-term non-progressors (LTNP). Although LTNPs are a heterogeneous population, most LTNPs exhibit low levels of viremia. Two extreme

profiles within the LTNP population have been reported (Table I). On the one hand, very few LTNPs maintain their CD4+ T cells despite high levels of viremia (at least 10.000 copies of viral RNA/ml of plasma). These individuals are called viremic non-progressors (VNP). Because viral replication is not controlled in VNPs, these individuals must possess a mechanism that protects them against CD4+ T cell loss and HIV-induced immunodeficiency. On the other hand, less than 0.5% of individuals infected with HIV-1 exhibit a spontaneous, highly efficient control of viral replication. This control is so effective that the viral load is often undetectable in the blood by routine clinical assays. Patients exhibiting such control for long periods are termed "elite controllers" or "HIV controllers (HIC)" [6]. The HLA alleles B27 and B57 are highly enriched in this population. However, the presence of these protective HLA alleles is neither sufficient nor always necessary to achieve control of infection.

3. Early virus-host interactions and their impact on disease progression

The risk of progressing rapidly or slowly towards AIDS can be somewhat predicted shortly after infection. Viremia levels at 6 months post-infection predict the rate of disease progression [7]. The levels of T cell activation as measured by the frequency of CD8+ T cells expressing HLA-DR and CD38 also predict the disease progression profile [8]. Notably, T cell activation is a stronger predictor than is viremia [8, 9] and is predictive even before seroconversion [10]. Recent studies demonstrate that inflammatory and coagulation biomarkers, such as IL-6, sCD14 and D-dimer, are better correlated with mortality than is T cell activation [11, 12]. Moreover, the levels of certain inflammatory molecules during acute infection, such as IP-10, are better predictors of rapid disease progression than viremia or CD4+ T cell counts [13].

Early virological and immunological features may be strongly prognostic because they reflect the presence of protective host factors (such as HLA-B27) and/or because the balance that is established between the virus and the host during the early phase of infection impacts the subsequent evolution of the infection [14, 15] (Figure 2). Therefore, beginning antiretroviral therapy during primary infection may provide significant benefits to HIV-infected patients. It has been suggested that early treatment could have a favourable impact on the reduction of viral reservoirs, the preservation of immune responses and protection from chronic immune activation [16]. It was recently reported that some HIV-infected patients who interrupted prolonged antiretroviral therapy that was initiated shortly after primary infection can control viremia [17]. Such post-treatment controllers (PTCs) have achieved control of infection through mechanisms that are, at least in part, different from those commonly observed in HICs. Indeed, PTCs had more severe primary infections than did HICs (Figure 2). Importantly, most PTCs lacked the protective HLA B alleles and instead carried risk-

associated HLA alleles (i.e., HLA*B35) that were largely absent among the HICs. Accordingly, PTCs had poorer CD8+ T cell responses than did HICs. PTCs also had lower levels of T cell activation than HICs. Therefore, the mechanism of virus control seems different between PTCs and HICs.

It is likely that infection control in PTCs was not achieved spontaneously but was favoured by the early initiation of therapy. The frequency of PTCs is estimated between 5% and 15% of patients beginning combination antiretroviral therapy (cART) shortly after primary HIV infection (PHI) [17-20], which is significantly higher than the proportion of HICs [21, 22]. Such a significant proportion of PTCs has only been observed after early initiation of treatment and not when therapies were begun during the chronic phase, where reported cases are even scarcer [23]. The rarity of PTCs worldwide may be explained by the fact that only a very small proportion (approximately 2%) of patients in the French Hospital Database on HIV who initiated cART early during PHI experienced a treatment interruption [17]. Indeed, in the absence of a reliable marker to predict the outcome after therapy discontinuation, even if started early, is not recommended outside clinical structured protocols. Non-controlled infection after treatment interruption increases risks of morbidity and mortality [24] and also of infection transmission.

PTCs were able to control viral replication on the long term and in some cases, even exhibited a progressive decrease in viral reservoir [17]. This may be at least partially related to the weak contribution of long-lived cells, such as central memory CD4+ T cells (T_{CM}), to the total circulating reservoir in these individuals. It will be of interest to understand why PTCs can control viral replication. The fact that it may be feasible to help the host develop protective responses gives substantial hope for the development of a cure. However, most patients are diagnosed with HIV only after several years of infection and in addition the ability to translate the PTC's mechanisms of control to other patients is as yet uncertain.

4. Animal models of spontaneous protection

Animal models allow a deeper understanding of early virus-host interactions, particularly with respect to the compartments that are crucial for the education of adaptive immune responses or that represent the major sites of viral replication (lymph nodes and mucosa). The only animal model that fully reproduces the physiopathology of AIDS consists of Asian monkeys (macaques) infected with SIVmac. In recent years, the macaque/SIVmac model has revealed key characteristics of HIV-1 pathogenesis. For example, this model has demonstrated the impact of the viral protein Nef in maintaining a high viral load *in vivo* and for disease progression [25, 26]. This model has also highlighted the role of CD4+ T_{CM} as main targets of the virus *in vivo* [27-29], as well as the dramatic and rapid depletion of CD4+ T cells in the

gut [30] and contributed to demonstrate that microbial translocation is associated with disease progression [31]. Finally, the macaque/SIVmac model has revealed the significant trafficking of immune cells, such as of natural killer (NK) cells and plasmacytoid dendritic cells (pDCs), from the periphery to the gut mucosa during infection [32, 33]. Trafficking to the gut was associated with upregulation of $\alpha4\beta7$ on NK cells and pDCs and blocking of $\alpha4\beta7$ could reduce viral loads in this tissue [34].

Macagues infected with SIVmac exhibit all the different disease progression profiles described in HIV-1-infected humans, from rapid to slow progression. Spontaneous control of viral replication has been observed in at least two macaque species (rhesus and cynomolgus) with specific MHC or TRIM5α alleles [35-37]. Some SIVagm strains (SIVagm.ver90 and SIVagm.sab92018) induce AIDS in pig-tailed macaques, but not in rhesus macaques [38-40]. Infection of rhesus macaques with SIVagm.sab92018 is characterised by high levels of viremia and dramatic mucosal CD4⁺ T cell depletion during acute infection followed by complete control of SIVagm replication defined as follows: undetectable viral load in the blood and tissues beginning at three months post-inoculation (pi) and continuing for at least 4 years; sero-reversion; complete recovery of mucosal CD4⁺ T cells by 4 years pi; normal levels of immune activation; and no disease progression [39]. Virus control was independent of MHC, APOBEC and Trim5 genotypes. This "functional cure" of SIVagm infection in rhesus macagues could be reverted by depleting CD8+ cells, which resulted in a transient rebound in viral load, suggesting that control may be at least partly immune mediated. This represents a new animal model of controlled lentiviral infection, and other, complementary models are currently under development.

Macaque models are being used to examine the effect of short-term cART initiated at different stages during acute infection on viral dissemination and replication. The Zidovudine (AZT)/Lamivudine (3TC) and Indinavir (IDV) combination efficiently reduced viral replication in all tissues when treatment was initiated before peak viremia. When the same treatment was initiated after peak viremia, the effect of treatment was stronger in the gut than in the secondary lymphoid tissues [41]. Studies are currently being conducted to evaluate pre-exposure prophylaxis (PrEP) strategies, such as rectal application of drug combinations before challenge [42].

Complete cART-associated suppression of SIVmac in rhesus macaques, even after several weeks and months of treatment, has been rarely achieved thus far. Without complete suppression, testing of strategies to reduce viral reservoirs is confounded by ongoing cycles of viral replication that can replenish such reservoirs. One major obstacle was the natural resistance of SIVmac to non-nucleoside reverse transcriptase inhibitors (NNRTIs). Efforts are currently underway to achieve the goal of drug-induced full viral suppression in the macaque model, by improving drug combinations and administration strategies, and early results are

[43]. Alternative strategies consist of chimeric encouraging simian-human immunodeficiency viruses, or SHIVs, in which the SIVmac reverse transcriptase (RT) is replaced with the RT from HIV-1 (RT-SHIV). RT-SHIVs have the advantage of being as susceptible to both nucleoside and non-nucleoside RT inhibitors as HIV-1. However, these chimeric viruses also have limitations; for example, the physiopathology of infection is not the same as with the wild-type virus. Recently, Shytai et al. succeeded in completely suppressing viral replication by intensifying cART in SIVmac-infected rhesus macaques [44]. Altogether, efficient treatment regimens in macaques will represent an essential model for answering crucial questions in the HIV cure research field, such as more precise insights into the nature of viral reservoirs in distinct body compartments during long-term treatment, the impact of early treatment on inflammation and viral reservoirs and the exact source(s) of virus during viral rebound.

Fundamental clues regarding the mechanisms that protect against AIDS also reside in the natural hosts of SIV, such as African green monkeys (AGMs), sooty mangabeys (SMs) and mandrills [45]. In contrast to macaques, these African non-human primates are natural carriers of SIV in the wild. Protection against AIDS in natural hosts occurs despite viral replication in the blood and gut at levels similar to or higher than in HIV-1-infected humans and SIVmac-infected macaques [46]. Protection is associated with an absence of both chronic T cell activation and chronic inflammation [45, 47]. The studies in natural hosts have contributed to the increased consideration of the major role of chronic immune activation in the development of AIDS. In countries where cART is accessible, the nature of HIV disease has largely shifted from one of immunodeficiency to one of chronic inflammation [48]. Deciphering the factors that predispose the natural host to control inflammation is the subject of several current studies and may have a major impact on translational research.

5. Insights into immune responses conferring spontaneous control of viral replication

HIV infection leads to a period of acute infection with vigorous viral replication, which is then partially controlled and stabilises 3-6 months after infection. The pace and level of virus control depends on both viral and host determinants. Innate responses are mobilised to first counteract the virus and to assist in the development of adaptive cellular and humoral responses against HIV. However, these defences are generally imperfect and are eventually overwhelmed by the infection. Analysis of cases of immune-driven natural control of infection offers the opportunity to examine the characteristics of optimal immune function.

(a) Innate responses

NK cells

binding to HLA class I ligands [64].

NK cells are key effectors of innate immunity. Through their capacity to mediate cytolysis and to produce numerous cytokines, NK cells can control the virus during the earliest stages of infection and shape the adaptive immune response. Thus, NK cells constitute one of the first lines of defence against HIV-1. Accordingly, several reports have linked enhanced basal and/or induced NK cell activity with protection from infection in groups of intravascular drug users, commercial sex workers and sero-discordant partners who remain sero-negative despite repeated exposure to HIV-1 [49-53] (Table I). Once established, HIV-1 infection is accompanied by an expansion of NK cells [54]. However, a skewed distribution of NK cell subpopulations and loss of cell functions occur as a consequence of exposure to HIV-1 [55]. NK cell dysregulation is at least partially driven by viral products, which suggests that HIV-1 may have evolved to escape NK cell-mediated control [56]. Evidence of a role for NK cells in the control of HIV infection comes from genetic and epidemiological studies. These studies consistently show that when linked with some HLA class I molecules carrying the Bw4-80I motif, some killer immunoglobulin-like receptor (KIR) (KIR3DS1/KIR3DL1) alleles are associated with low-level viremia and slow disease progression [57, 58]. The mechanisms underlying this control are not completely clear, but the interactions between KIR3DS1/KIR3DL1 and their Bw4 ligands may determine the expansion of specific subpopulations of NK cells [59] or the licensing of NK cells with increased responsiveness [60]. Recent studies suggest that changes in the peptides bound by HLA molecules may critically impact the way KIRs are stimulated by the HLA class I/peptide complex [61]. Interactions between KIR3DL1 and HLA class I alleles carrying the Bw4-80I motif are peptide specific [62], and some peptide residues are directly involved in the binding of KIR3DL1 to its ligand [63]. Along these lines, compelling evidence of the impact of NK cells on virus control in the context of a particular KIR background comes from the observation that HIV-1 evolves to evade NK cellmediated immune pressure by selecting for sequence variants that specifically affect KIR

Natural control of HIV-1 infection appears to begin early in most HICs who usually have lower levels of viremia than do progressor patients during acute infection [22]. The HLA-B alleles B*27 and B*57, which are commonly over-represented in HICs [65-67], are members of the Bw4-80I group [68], suggesting that NK cells may contribute to establishing HIV control in these patients through direct cytolytic or non-cytolytic anti-HIV activities or by favouring the induction of an efficient CD8+ T cell response (see below) through optimal crosstalk with dendritic cells. Various reports suggest that NK cells from HICs have increased cytolytic and secretory potential [69-71], which may be associated with particular NK cell receptor profiles [69, 71]. However, it remains unclear how this impacts the control of

infection *in vivo*, and NK cells from HICs exhibited only a modest capacity to suppress viral replication in autologous CD4+ T cells *in vitro* [72].

Plasmacytoid dendritic cells, type I interferon and intrinsic immunity

PDCs are another key component of the innate immune response. These cells influence HIV pathogenesis through their capacity to produce type I interferon (IFN-I) [73]. IFN-I upregulates interferon-stimulated genes (ISG), several of which possess antiviral activity. During acute infection, pDCs may be a critical antiviral agent. IFN-α has long been known to block HIV-1 replication in vitro and in vivo [74-77]. Several studies performed during the last few years have identified a number of IFN-stimulated cellular factors (e.g., MX2, BST-2/tetherin, TRIM5a, APOBEC3G and SAMHD1) that can restrict retroviral replication [74, 78-81]. During acute infection, transmitted founder viruses (HIV strains that succeed in establishing a persistent infection [82]) are more resistant to IFN-I than chronic-phase HIV strains [83, 84]. In addition to its antiviral effect, IFN-I may also play an important role in the stimulation of innate responses (NK) and the shaping of adaptive immune responses [85]. In contrast, during chronic HIV infection, the continuous stimulation of pDCs may be deleterious via several mechanisms. The induction of inflammatory cytokines could enhance the trafficking of new target cells to sites of viral replication [86]. The IFN-mediated induction of Indoleaminepyrrole 2,3-dioxygenase (IDO) is associated with the loss of the TH17/Treg balance [87]. PDCs with increased TRAIL, another ISG, on their surfaces could induce apoptosis of DR5-expressing CD4+ T cells [88]. During a non-controlled HIV infection, the number of circulating pDCs decreases, which is likely due to their migration to the lymph nodes and the gut where extensive HIV replication occurs [89]. In contrast, pDC levels in the blood of HICs are comparable to those found in normal donors and can produce high levels of IFN-α in response to HIV [90, 91]. PDCs from HICs do not express TRAIL on their surfaces but carry high intracellular levels that can be mobilised to the membrane upon encountering HIV [90]. These results suggest that pDCs from HICs may specifically produce IFN-α and induce the apoptosis of infected cells [90, 91]. Along these lines, pDCs from HICs can limit HIV replication in vitro when co-cultured with infected cells [91].

Other IFN- α -independent cellular factors can also block viral replication (e.g., p21) [92]. Host cells appear to have evolved a number of restriction factors that can block infection at different stages of the viral replication cycle. Many of these factors, which likely play critical roles in preventing cross-species transmission [93], are counteracted by HIV-1 proteins [94]. However, it is tempting to speculate that inter-individual differences in expression levels and/or polymorphisms in these cellular factors may have an impact on HIV pathogenesis. Studies examining this question have produced conflicting results thus far. Polymorphisms in the *Trim5* α gene [95] and different expression levels of APOBEC3G [96] are associated with

greater control of infection. However, this result has not been confirmed by other studies [97, 98], and the expression of IFN-induced restriction factors may also be driven by viral replication [99]. Nevertheless, intrinsic cellular resistance to infection [e.g. linked to lack of CCR5 expression, high p21 levels] has been associated with both protection from infection among HIV-exposed seronegative individuals [100, 101] and control of infection among HICs [102, 103].

(b) Adaptive cellular responses

CD8± T cell responses

CD8+ T cell responses have been consistently associated with control of infection following acute HIV infection. The appearance of HIV-specific CD8+ T cells coincides with a decrease in viremia during primary infection [104] and the selection of viral escape mutants in regions targeted by these responses [105]. CD8+ T cell responses targeting HIV-1 Gag epitopes are associated with smaller viral loads [106, 107], which may be associated with a higher fitness cost for the virus to escape from Gag-restricted responses [108]. In vivo depletion of CD8+ cells in macaque models of pathogenic SIV infection has demonstrated that CD8-depleted macaques are unable to control infection during acute infection [109, 110]. CD8+ depletion also results in increased viral loads in chronically infected macaques [111, 112]. CD8+ T cells can counteract HIV by non-lytic (secretion of soluble factors such as β-chemokines or the as yet unidentified cellular antiviral factor CAF) [113, 114] or lytic mechanisms (cytolysis of infected cells through the Fas-Fas ligand pathway or cytotoxic granules) [115, 116]. However, CD8+ T cells can only partially control HIV. Continuous HIV replication provokes the gradual loss of CD8+ T cell functions associated with the expression of negative regulatory molecules, such as PD-1 [117]. In addition to progressive CD8+ T cell exhaustion, HIV infection is characterised by a skewed distribution of HIV-specific CD8+ T cells with low frequencies of effector cells that may be especially prone to apoptosis [118, 119].

Due to the enrichment of protective HLA class I alleles among HICs, these individuals were soon proposed as a convenient model to uncover the characteristics of efficient CD8+ T cell responses against HIV-1. Despite the low levels of circulating virus in HICs, high frequencies of HIV-specific CD8+ T cells have been observed in these individuals [67, 120]. These cells have maintained their capacities to proliferate in the presence of HIV antigens and to secrete IL-2 and other cytokines and chemokines [121, 122]. Moreover, HIV-specific CD8+ T cells in HICs have been reported to possess or rapidly up-regulate cytotoxic granule contents [123, 124] and accordingly, have a striking capacity to eliminate infected autologous CD4+ T cells [67]. This enhanced capacity to suppress HIV infection is linked to a higher frequency of further differentiated cells in association with a discordant CD38^{low}HLA-DR^{high} phenotype [67]. HIV-specific CD8+ T cell responses in HICs preferentially target epitopes in Gag, and

Gag-specific responses account for most of their capacity to suppress HIV infection [125], which may be due to faster recognition of infected cells [126].

Some of the characteristics of HIV-specific CD8+ T cells from HICs are not found in most HIV-infected patients, even during primary infection [127]. Metabolic alterations in HIV-specific CD8+ T cells have been proposed to occur very early during acute infection due to hyperproliferation associated with continual stimulation of the cells [128]. CD8+ T cells from HICs may also possess particular intrinsic characteristics. For example, selection of particular high-avidity TCR clonotypes associated with a broader capacity to recognise epitope variants and to orchestrate enhanced cytolytic functions has been shown to distinguish HICs from viremic HIV-infected patients sharing the same protective HLA class I alleles [129, 130]. Selection of such clonotypes occurs very early, although the mechanisms of selection are unknown. The function of myeloid dendritic cells, which are principally responsible for priming T cell responses, is altered during primary infection [131] (blood). In contrast, myeloid dendritic cells from HICs have enhanced antigen-presenting capacities but produce lower levels of pro-inflammatory cytokines ([132] our own unpublished results). This profile may favour T cell priming and the selection of specific optimal clonotypes in the context of reduced antigenemia and a weakly inflammatory environment.

During the chronic phase of infection, instead of a strong effector CD8+ T cell response, many HICs present with a small number of quiescent memory CD8+ T cells [125, 133, 134]. These responses may constitute a pool of preserved CD8+ T cells that are highly reactive to small quantities of antigen and can rapidly gain effector capacities in response to viral relapses from HIV reservoirs. This hypothesis is supported by *in vitro* experiments in which memory CD8+ T cells from these HICs were able to gain cytotoxic activities within a few days of stimulation with cognate peptides [135]. These experiments are not completely conclusive because cells from non-controller patients also gain anti-HIV capacities upon stimulation *in vitro* [136]. Therefore, further studies are necessary to identify clear distinguishable characteristics in memory CD8+ T cell from HICs, which may hold important clues for the development of an efficient T cell-based vaccine.

CD4± T cell responses

CD4+ T cells play a multifaceted role in HIV infection. CD4+ T cells provide crucial help to dendritic cells and B cells for the induction of HIV-specific CD8+ T cells and antibodies. Furthermore, CD4+ T cells are the main cellular target of HIV, and HIV-specific CD4+ T cells are preferentially infected by the virus [137]. Induction of activated HIV-specific CD4+ T cells in vaccine trials has been associated with a higher risk of HIV infection [138] or with faster

viral rebound in HIV-infected individuals upon interruption of treatment [139]. In contrast, induction of HIV-specific CD4+ T cell responses was not associated with an increased risk of infection in the RV144 vaccine trial [140], and higher frequencies of HIV-specific CD4+ T cell responses during primary infection have been associated with higher CD4+ T cell counts and lower viral loads after short-course antiretroviral treatment [141]. Moreover, several studies have shown that some HIV-specific CD4+ T cells develop cytolytic potential and carry high levels of granzyme A [142, 143]. These cells may be able to eliminate infected macrophages and to a lesser extent, activated CD4+ T cells expressing high levels of HLA class II molecules [142]. A recent study linked high levels of cytotoxic HIV-specific CD4+ T cells during acute infection with lower set-point viremia, supporting a direct effector activity for this subset of HIV-specific CD4+ T cells [144].

In general, primary HIV infection is accompanied by the depletion of HIV-specific CD4+ T cells and impaired cell functionality, particularly the capacity to proliferate and produce IL-2 [145]. As was the case for CD8+ T cells, in HICs, HIV-specific memory CD4+ T cells maintain their functionality [146-148]. High-quality memory CD4+ T cells in HICs have been associated with reduced expression of the negative immunoregulatory molecule cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) [149] and lower levels of FoxO3a-mediated proapoptotic transcriptional activity [150]. HIV-specific CD4+ T cells from HICs also exhibit high avidity for immunodominant Gag peptides, which may allow them to react to low levels of antigens [151]. The class II HLA alleles HLA-DRB1*13 and HLA-DQB1*06 have been associated with strong HIV-specific CD4+ T cell responses in HICs [152].

CD4+ follicular T helper (T_{FH}) cells, which regulate the development of antigen-specific B cell immunity, have received special attention in the last couple of years. T_{FH} cells are highly susceptible to HIV-1 infection *in vitro* and are a major site of viral replication and a viral reservoir [153, 154]. T_{FH} cells are infected at higher frequencies in macaques and humans than in SMs [155]. In contrast to most other CD4+ T cells, this subset is expanded and accumulates in lymph node germinal centers during HIV and SIVmac infections [153, 156-158]. Whether T_{FH} in HIV infection show an altered function that could impact anti-HIV antibody development is unclear.

Regulatory CD4+ T cells (Tregs) may play a dual role in HIV pathogenesis. Tregs may contribute to reduce pathogenesis by controlling chronic immune inflammation but may facilitate infection by suppressing the activation of effector T cells [159]. During HIV infection, Tregs accumulate in the gut [160]. The ratio of Treg:TH17 cells decreases, and this imbalance may have a deleterious effect on the integrity of the gut mucosa [161]. In contrast

to other effector CD4+ T cell subsets, Tregs preserve their suppressive capacity despite HIV-1 infection [162]. HICs appear to maintain similar or lower levels of Tregs than do healthy individuals [162-165], and their CD8+ T cells may evade Treg-mediated suppression [166]. This mitigated regulatory response in HICs may help to maintain a robust and efficient T cell response but may also explain the relatively high immune activation observed in these individuals [163], which is associated with some loss of CD4+ T cells (see chapter 6).

(c) Humoral responses

HIV infection elicits an antibody response that targets HIV envelope protein and is non-neutralising during the early stages of infection. Neutralising antibodies are only generated months after infection is established and usually lag behind viral escape mutants [167]. A blunted antibody response during HIV infection is associated with B cell dysfunction. Some individuals, elite neutralisers, can elicit broadly neutralising antibodies that recognise conserved regions of the virus envelope protein [168]. The presence of these broadly neutralising antibodies is not associated with a dramatic control of viremia *in vivo* but has been shown to strongly decrease viremia when administered to SHIV-infected macaques [169]. Neutralising IgA has been found in the genital tract of different cohorts of highly exposed but sero-negative females [170, 171], suggesting that these antibodies contribute to protection from AIDS acquisition in these subjects.

High levels of IgG2 antibodies targeting gp41 were reported as a strong correlate of slow progression to AIDS [172], and these antibodies are also found at high levels in HICs [173, 174]. The mechanism through which these antibodies contribute to HIV control is unknown, although it seems to be unrelated to direct neutralisation. In general, HICs possess heterogeneous but low levels of neutralising antibodies, suggesting that they are not a major determinant of virus control [134, 175]. In contrast, greater antibody-dependent cell-mediated cytotoxicity (ADCC) potential associated with both the quality of non-neutralising antibodies and the levels of FCy receptors on the surface of effector cells has been observed in HICs [17, 175-177]. The induction of ADCC-mediating antibodies was observed in vaccinated volunteers in the RV144 vaccine trial [178], showing marginally significant protection from HIV infection [179] and further reinforcing the potential therapeutic utility of ADCC.

Additional antibody-related activities may impact HIV infection. Non-neutralising antibodies form immune complexes with soluble HIV antigens that stimulate Fcy receptors expressed by myeloid cells, particularly macrophages. Fcy receptor aggregation provokes a blockade of viral replication through the induction of p21 and the alteration of the *de novo* synthesis pathway of dNTPs, which are necessary for the reverse transcription step of viral replication

[92, 180]. In contrast, anti-HIV antibodies may compete with complement to opsonise viral particles. The complement system is part of the innate immune response that is activated immediately upon HIV-1 infection. Among other activities [181], complement opsonisation of viral particles has been shown to favour HIV-1 capture and uptake by dendritic cells [182], which is associated with enhanced intracellular co-localisation of HIV antigens with HLA class I molecules and effective CD8+ T cell priming by dendritic cells. This effect is gradually lost with the deposition of HIV IgG on viral particles [183]. In summary, further studies need to be conducted to understand the impact of non-neutralising antibodies *in vivo*.

6. Spontaneous control of chronic inflammation in HIV/SIV infections

The proportion of infected CD4+ T cells is too small to fully account for the extent of CD4+ T cell decline. Many data point towards systemic immune activation as the factor responsible for HIV-induced immunodeficiency [12]. Indeed, studies on HIV-2 infection and in natural hosts indicate that viral replication alone is not sufficient to induce AIDS. Many studies have demonstrated that inflammation is even more closely associated with mortality in HIV-infected patients than T cell activation. Therefore, inflammatory and coagulation biomarkers (highly sensitive C-Reactive Protein (hsCRP), IL-6 and D-Dimers) are associated with immunological failure, clinical events and AIDS- and non-AIDS-related mortality [11].

(a) Natural control of inflammation in the context of high-level viremia

Natural hosts exhibit spontaneous protection against chronic immune activation despite high levels of viremia, high mucosal replication and dramatic CD4+ T cell loss in the gut [46]. During the chronic phase of infection, peripheral and tissue T cell activation levels are not or are only modestly increased. No elevation in the expression of inflammatory cytokines is observed [46]. There are no increases in coagulation markers such as D-Dimers [184]. A lack of chronic immune activation is observed despite an initial transient activation or mobilisation of pDCs, mDCs, NK cells and T cells [185-187]. Indeed, the acute phase of SIVagm infection is characterised by the recruitment of pDCs and mDCs to the lymph nodes, IFN- α production, induction of ISGs and corresponding protein expression.

Early inflammation may be essential for both the virus and the host. Inflammation would be beneficial to the virus because it attracts target cells to the site of infection and would allow the virus to establish a persistent infection. For the host, the induction of early innate antiviral responses (including IFN- α) would allow partial control of viral replication. Inflammation would then be resolved before the end of acute infection. For example, most ISGs are down-regulated back to normal levels in natural hosts in contrast to pathogenic HIV/SIVmac infection [46]. The lack of chronic inflammation would prevent immunemediated pathology and disease progression.

However, there are major differences compared with SIVmac infection in macaques: the levels of several cytokines, including IFN- α , are lower than those observed during acute SIVmac infection, which is not due to a functional defect in the ability of pDCs to sense the virus [188, 189]. Indeed, the TLR7/TLR9/IRF7 pathway is functional [188, 189]. In addition, natural hosts preserve their TH17 cells, and their epithelial barriers are not damaged and consequently, they show no signs of microbial translocation. This could at least partly explain the lack of systemic immune activation during the chronic phase. Finally, several differences in viral reservoirs in natural hosts with respect to HIV-1 and SIVmac infections have been reported: a smaller DNA reservoir (PBMC), less replication in the lymph nodes, less infection of CD4+ T_{CM} , no infection of T_{FH} , and no or rare trapping by follicular dendritic cells [46, 190]. The relevance of these observations to the lack of AIDS requires further investigation, but it is interesting to notice that small reservoirs and low contribution of T_{CM} cells have been associated with control of HIV infection in B57+ bearing-non progressor patients and also in PTCs from the VISCONTI study [17, 191].

Similar to natural SIV hosts, VNPs do not control viral replication but nonetheless, maintain close to normal CD4+ T cell counts for many years in the absence of treatment. The maintenance of CD4+ T cells is associated with a low frequency of activated (DR+CD38+) and proliferating (Ki-67+) CD4+ and CD8+ T cells [192]. Therefore, attenuated infection is equally associated with a lack of chronic immune activation. Obviously, a functional cure such as in VNPs (or natural hosts) without control of the virus is less attractive because of the risk of viral transmission. However, it is crucial to understand how VNPs avoid chronic immune activation. VNPs are rare, and unfortunately only limited information on their immune responses is available.

(b) Inflammation and CD4+ T cell loss in HIV controllers

HICs can experience modest CD4+ T cell loss despite controlled viremia. Higher CD4+ and CD8+ T cell activation is associated with a progressive loss of CD4+ cell counts in HICs [193]. CD8+ T cell activation levels in HICs are also higher than in healthy donors, efficiently treated aviremic patients [193] and PTCs. Higher levels of sCD163, sCD14, IP-10, TNF-α, sTF, D-dimers and hsCRP as well as an increased risk of atherosclerosis have been observed in some HICs compared with healthy donors or aviremic treated patients[194-197]. HICs also seem to have elevated levels of microbial translocation compared with HIV-negative and cART-suppressed individuals [163]. In a recent study, the relationship between inflammatory biomarkers and the CD4+ T cell decreases observed in some HICs has been investigated in a large HIC cohort [197]. In this study, IP-10 positively correlated with activated CD8+ and CD4+ T cells in HICs. Moreover, IP-10 and sCD163 levels in HICs

predicted the risk of CD4+ decline. Therefore, the association between inflammation and disease progression is similarly present in HICs as in other HIV-infected individuals [11, 13]. It is unclear what drives chronic inflammation in HICs despite the control of viral replication, but it could be associated with extremely low but continuous HIV replication over several years. Very low plasma levels of virus can be detected by ultrasensitive RT-PCR assays in HICs, revealing the persistence of viral replication despite maintaining viremia close to the limit of detection by standard RT-PCR [198]. Although HICs maintain a remarkable control of infection, blips in viral load levels have been observed for many of these individuals [199]. In contrast, some HICs never experience blips, even over long follow-up periods and when using ultrasensitive techniques that detect 1 RNA copy/ml of plasma. Interestingly, these HICs are similar to healthy individuals from a transcriptomic point of view [200]. HICs with blips more often exhibit CD4 T cell loss. Moreover. theoretically, chronic low-level inflammation in HICs could also be driven by higher viral replication in a few as yet unidentified sanctuaries. Such sanctuaries could correspond to immune-privileged compartments in the body, such as the brain. Recent studies in the macague model suggest that this sanctuary could also be represented by TFH cells in germinal centres (GC) [201]. In late-stage HIV infection, GCs are characterised by infiltration of CD8+ cells [202]. However, under normal conditions, GCs are devoid of CD8+ T cells. Theoretically, GCs could represent a compartment in the body where HIV could evade control by CD8+ T cell responses and replicate to higher levels than in the remaining lymph nodes and mucosal tissue cells. Finally, because most HICs have been infected for long periods, lymphoid tissues might represent some of the damage described in normal progressors, such as disruption of the epithelial barrier leading to translocation of microbial products. Indeed, HICs show increased levels of microbial translocation markers [197]. Translocation of microbial products into systemic circulation could then fuel immune activation. Studies in the non-human primate model have provided the proof of concept that higher systemic LPS levels lead to increased T cell activation [203], which could reactivate latent virus, leading to a vicious cycle.

7. Hopes and future directions for a cure

An HIV cure will succeed by targeting viral reservoirs, but *in vitro* and *in vivo* evidence suggests that host immunity should be targeted concomitantly. Very small viral reservoirs are most likely necessary but not sufficient to ensure the control of viremia off treatment, and the induction of efficient responses against HIV, eventually combined with anti-inflammatory approaches, will be necessary to eliminate HIV-producing cells in reservoir-purging protocols

(see [204] for further information on current HIV cure strategies). Several novel treatment approaches to improve host immune responses are currently under investigation.

(a) Early treatment interventions

Although current antiretroviral strategies seem to have reached their limits in terms of blocking HIV replication, early treatment initiation may provide further advantages. As previously described, treatment initiation during primary HIV infection has been linked to lasting remission of HIV infection in a group of adults [17]. Treatment initiation immediately after birth also allowed a functional cure of HIV infection in a child after treatment discontinuation [205]. Early treatment limits the establishment of viral reservoirs [206] and severely restricts viral diversity [207]. In addition, treatment initiation during primary infection has been shown to preserve CD4+ T cell homeostasis and the function of NK cells, B cells and HIV-specific T cells [55, 145, 208]. Therefore, early treatment may allow an optimal maturation of the anti-HIV response by reducing viremia and inflammation, which may favour the control of infection after treatment interruption in some individuals with low levels of infected cells. Larger studies need to be performed to identify the mechanisms associated with control after treatment interruption and to uncover predictive markers of post-treatment control.

(b) Immunotherapies

Immunotherapies based on the administration of IL- 2, IL- 7 or IL-15 alone or in combination with vaccine candidates aim to enhance immune function and restore T cell homeostasis [209]. IL-7 has garnered some interest [210, 211], but it increases the number of infected cells [210, 212], and thus far, these approaches have not shown sufficiently favourable effects *in vivo*.

Treatment with IFN- α has been shown to transiently decrease viral loads during chronic infection [74, 213], and IFN- α monotherapy was recently shown to allow control of viremia and reduction of viral reservoirs after antiretroviral treatment interruption in 48% of individuals who received IFN α for several weeks in addition to their HIV-suppressive cART regimens [214]. The mechanisms underlying this effect are still unclear, but as discussed above, IFN α treatment may enhance the immune response in treated individuals and also up-regulate HIV restriction factors, thereby rendering target cells less susceptible to HIV infection. Nevertheless, IFN α therapy requires further exploration because conflicting results have been obtained depending on whether it is administered in the absence or presence of cART or in patients with very low CD4+ T cell nadirs [20, 215].

The observation that exhaustion of HIV-specific T cells is accompanied by enhanced expression of negative immunoregulatory molecules such as PD-1 has nurtured the hypothesis that targeting these immunomodulatory pathways may be a promising therapeutic approach in the fight against HIV (review in [216]). In vitro, blockage of PD-1 interactions with PD-1 ligands restores CD4+ and CD8+ T cell functions, particularly proliferation and cytokine production. Proof of concept studies on SIV-infected macaques have shown that PD-1 blockade results in the expansion of CD8+ T cells with improved functionality [217], longer survival and decreased viral loads in infected animals [217] and may delay viral rebound after treatment interruption [218]. Anti-exhaustion strategies may not restore HIV-specific T cell functions to the levels found in HICs because some intrinsic characteristics of HIC cells are determined very early during infection (see above). However, the restoration of partial T cell function may act in synergy with additional effects of these strategies. PD-1-expressing cells are preferential targets of HIV-1 infection [219]. Furthermore, the PD-1 pathway has been linked to the establishment of HIV latency, and triggering PD-1 may help to purge HIV-1 reservoirs. Moreover, in vivo PD-1 blockage also produces a reduction of the immune activation associated with chronic SIV infection [220].

c) Anti-inflammatory therapies

Because immune activation is a major determinant of HIV pathogenesis, direct targeting of deleterious inflammation is increasingly attracting attention. Many assays are currently under investigation [221]. Chloroquine analogues are among the first anti-inflammatory molecules assessed in HIV-infected patients [222, 223]. The rationale behind this approach is that chloroquine can inhibit the recognition of HIV via TLR7 and TLR9 [224]. Whereas one study showed that short-term chloroquine administration during chronic infection in a small group of cART-naïve patients resulted in a reduction in T cell activation markers in the absence of changes in plasma viremia [225], a randomised double-blind trial with a larger group of HIVinfected patients demonstrated that longer hydroxychloroquine administration resulted in faster CD4+ T cell decay and some increase in viral replication [226]. PD-1 blockade during chronic SIV infection markedly reduced the expression of transcripts associated with type I IFN signalling in the blood and colorectal tissue of rhesus macaques, even in the presence of high levels of viremia [220]. Reduced type I IFN signalling was associated with a profound decrease in plasma LPS levels, suggesting decreased microbial translocation into the blood. PD-1 blockade enhanced immunity to gut-resident pathogenic bacteria, control of gut-associated opportunistic infections and survival of SIV-infected macaques. The effects of PD-1 blockade on reducing hyperimmune activation could be a combination of enhanced immunity against gutresident pathogenic bacteria and repair of gut barrier permeability.

Statin administration to HIV-infected patients has been shown to reduce inflammation [227] and may decrease the risk of non-AIDS-defining malignancies [228] and co-morbidities [229]. In contrast, statins may increase the risk of developing diabetes [230]. Studies with other anti-inflammatory agents (non-steroidal anti-inflammatory drugs, pyrimidine synthesis inhibitors, probiotics and Cox-2 inhibitors) have revealed similarly contrasting results. Other trials with anti-inflammatory agents, such as anti-IL-6 and JAK inhibitors, are planned or are underway [231]. In conclusion, clinical trials have thus far shown that identification of the factors responsible for chronic inflammation is needed to be able to develop more specific, better targeted approaches.

Administration of cART on its own reduces inflammation, in a large part through the reduction of viral replication [232]. The effect of early cART treatment on inflammation was assessed more recently [233]. In a pilot study, Mega-cART initiated during acute infection resulted in the reduction of viral reservoirs down to 100 copies of viral DNA per 10⁶ CD4+ T cells at 6 months pi (M6). In parallel with the decrease in the viral reservoir, some inflammatory markers were reduced, including IP-10 and D-dimers at M6, whereas LPS and sCD14 were not. In another study in which treatment was also initiated during acute infection, the plasma levels of IP-10 were decreased, whereas those of 12 other cytokines studied were not [234]. Therefore, early treatment seems to diminish inflammation to some extent. It is not clear why the effect was only observed for some inflammatory markers but could be because they are more closely associated with replication levels or earlier and better markers of inflammation [13]. Intensifying treatment in cART-suppressed individuals may help to further reduce residual viral replication and immune activation. Early intensification of treatment with the CC chemokine receptor 5 (CCR5) antagonist maraviroc may be of special interest not only because the viruses that are capable of establishing an infection generally use CCR5 as a co-receptor but also because of the guick penetration and sustained concentrations of maraviroc in the rectal mucosa [235]. The addition of maraviroc to the antiretroviral regimen resulted in a faster reduction in newly infected cells [236], a decrease in microbial translocation markers [237, 238] and a faster increase in CD4 counts [236, 237]. Paradoxically, maraviroc also induced a slower decrease in plasma viremia. It has been suggested that this result may be due to an immunoactivatory effect of maraviroc. Indeed, MIP1b levels, CD8+ T cell counts and CD4+ T cell activation were higher in the maraviroc arm of the study [236, 237]. However, higher T cell activation levels were not observed in another study [238]. In an uncontrolled trial of maraviroc intensification, plasma LPS levels were actually increased [237] and soluble inflammation markers were similar in both arms at the end of the study [236].

It has long been assumed that HICs do not need cART due to their ability to efficiently control viral replication. In light of the higher levels of immune activation among HICs than patients on cART, it has recently been questioned whether cART could be beneficial to HIC patients. A recent pilot study comprising a small number of HICs assessed whether cART leads to a reduction in inflammation in HICs [239]. Antiretroviral therapy in these HICs led to statistically significant decreases in ultrasensitive plasma HIV RNA levels and rectal cell-associated HIV RNA as well as decreased T cell activation levels (DR+CD38+) in both the blood and gut. This pilot study suggests that even low-level replication results in immune activation. Larger studies will be needed in the future to test whether the reduction in immune activation observed has any clinical relevance for HICs.

d) Therapeutic HIV vaccines

The quest for an effective HIV vaccine has been unsuccessful thus far. Different vaccine candidates have been evaluated in 6 phase III clinical trials. Most candidates did not show any efficacy, and the STEP and HVTN503 trials showed an increased risk of HIV acquisition in vaccinated individuals (http://www.hvtn.org/media/pr/step111307.html). The **HVTN** trial was also recently interrupted for futility (http://www.niaid.nih.gov/news/newsreleases/2013/Pages/HVTN505April2013.aspx). Only the RV144 trial showed marginal efficacy [179]. Although a clear correlate of protection has not been identified for this trial, vaccination was associated with the induction of ADCCmediating antibodies [178]. Overall, conventional HIV vaccine development has been disappointing, and an efficient therapeutic HIV vaccine will likely require innovative approaches.

Dendritic cell-based vaccines have garnered special attention in recent years. One approach consists of vaccinating HIV-infected patients using autologous inactivated viruses to pulse dendritic cells derived from autologous monocytes *in vitro*. Although this process is extremely laborious, a proof of concept study has shown that the approach can significantly increase HIV-specific CD8+ T cell responses and may reduce viral load set-points after treatment interruption [240]. Another strategy consists of directly targeting vaccines to dendritic cells by fusing HIV antigens to monoclonal antibodies recognising receptors such as DEC205, DC-SIGN or CD40, which are specifically expressed by different subsets of dendritic cells [241]. These approaches have shown good immunogenicity in mice and NHP animal models. A recent study suggests the attractive possibility that a DNA vaccine consisting of HIV-1 Gag p24 fused to a soluble form of PD-1 would not only efficiently target antigens to dendritic cells, which express PD-1 ligands but also block the PD-1/PD1L pathway, which may result in enhanced priming of specific responses compared with DEC205-based vaccines [242].

SIV vaccines containing persistent rhesus cytomegalovirus vectors allow control of pathogenic SIVmac239 in 50% of vaccinated animals [243]. Control of infection in these animals is such that clearance of the virus has been reported to occur several months after infection despite profound viral dissemination during primary infection [244]. Vaccination of macaques with CMV vectors is accompanied by the induction of a polyfunctional SIV-specific effector CD8+ T cell response that seems to be responsible for virus control and accounts for the progressive elimination of infected cells. Interestingly, this response targets a breadth of non-conventional epitopes that are mainly restricted by MHC II molecules [244]. Such responses may provide specific advantages, as in the case of SIV/HIV infection, by favouring the recognition of variants that escape conventional CD8+ T cell responses and also circumventing the down-regulation of MHC class I by the virus.

The observation that passive transfer of broadly neutralising antibodies allows for the control of infection in chronically SIV - infected macaques [169] and delays viral rebound in acutely treated patients after cART cessation [245] suggests that a therapeutic vaccine that induces broadly neutralising antibodies may be effective for controlling HIV-1 in infected patients. Recent knowledge of the co-evolution of the viral epitopes and neutralising antibodies [246] might prove critical to anticipate the time-course that would be established between the virus and the immune system after an eventual treatment interruption. However, inducing such antibodies by vaccination remains a challenge. Current efforts are aimed towards identifying the structural characteristics associated with the neutralising efficacy of broadly neutralising antibodies that target major sites of vulnerability in the envelope protein [247], the events that lead to their production [248] and the B cell clones that express their germline precursors [249, 250] as well as toward designing mosaic antigens that elicit broad responses [251].

In conclusion, studies of natural protection against HIV/AIDS have already provided important clues about protective host determinants. Analyses of the immune responses functioning in these models offer signatures or correlates of protection that will then need to be validated in translational research. However, many questions remain to be investigated, such as the mechanisms of residual replication, the factors driving chronic immune activation and the mechanisms underlying treatment-induced protection against viral replication. In the future, such insights will be helpful to design efficient, well-targeted strategies for a cure and may also be of use for vaccine strategies.

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References

- [1] Barre-Sinoussi, F., Chermann, J.C., Rey, F., Nugeyre, M.T., Chamaret, S., Gruest, J., Dauguet, C., Axler-Blin, C., Vezinet-Brun, F., Rouzioux, C., et al. 1983 Isolation of a T-lymphotropic retrovirus from a patient at risk for acquired immune deficiency syndrome (AIDS). *Science* **220**, 868-871.
- [2] Deeks, S.G., Autran, B., Berkhout, B., Benkirane, M., Cairns, S., Chomont, N., Chun, T.W., Churchill, M., Di Mascio, M., Katlama, C., et al. 2012 Towards an HIV cure: a global scientific strategy. *Nat Rev Immunol* **12**, 607-614. (doi:10.1038/nri3262 nri3262 [pii]).
- [3] Hutter, G., Nowak, D., Mossner, M., Ganepola, S., Mussig, A., Allers, K., Schneider, T., Hofmann, J., Kucherer, C., Blau, O., et al. 2009 Long-term control of HIV by CCR5 Delta32/Delta32 stem-cell transplantation. *N Engl J Med* **360**, 692-698. (doi:10.1056/NEJMoa0802905

(001:10.1056/NEJW0808028 260/7/602 [pii])

360/7/692 [pii]).

- [4] Allers, K., Hutter, G., Hofmann, J., Loddenkemper, C., Rieger, K., Thiel, E. & Schneider, T. 2011 Evidence for the cure of HIV infection by CCR5Delta32/Delta32 stem cell transplantation. *Blood* **117**, 2791-2799. (doi:10.1182/blood-2010-09-309591 [pii]).
- [5] Yukl, S.A., Boritz, E., Busch, M., Bentsen, C., Chun, T.W., Douek, D., Eisele, E., Haase, A., Ho, Y.C., Hutter, G., et al. 2013 Challenges in detecting HIV persistence during potentially curative interventions: a study of the Berlin patient. *PLoS Pathog* **9**, e1003347. (doi:10.1371/journal.ppat.1003347
- PPATHOGENS-D-12-02882 [pii]).
- [6] Saez-Cirion, A. & Pancino, G. 2013 HIV controllers: a genetically determined or inducible phenotype? *Immunol Rev* **254**, 281-294. (doi:10.1111/imr.12076).
- [7] Mellors, J.W., Rinaldo, C.R., Jr., Gupta, P., White, R.M., Todd, J.A. & Kingsley, L.A. 1996 Prognosis in HIV-1 infection predicted by the quantity of virus in plasma. *Science* **272**, 1167-1170.
- [8] Giorgi, J.V., Hultin, L.E., McKeating, J.A., Johnson, T.D., Owens, B., Jacobson, L.P., Shih, R., Lewis, J., Wiley, D.J., Phair, J.P., et al. 1999 Shorter survival in advanced human immunodeficiency virus type 1 infection is more closely associated with T lymphocyte activation than with plasma virus burden or virus chemokine coreceptor usage. *J Infect Dis* **179**, 859-870.
- [9] Sousa, A.E., Carneiro, J., Meier-Schellersheim, M., Grossman, Z. & Victorino, R.M. 2002 CD4 T cell depletion is linked directly to immune activation in the pathogenesis of HIV-1 and HIV-2 but only indirectly to the viral load. *J Immunol* **169**, 3400-3406.
- [10] Hazenberg, M.D., Otto, S.A., van Benthem, B.H., Roos, M.T., Coutinho, R.A., Lange, J.M., Hamann, D., Prins, M. & Miedema, F. 2003 Persistent immune activation in HIV-1 infection is associated with progression to AIDS. *Aids* **17**, 1881-1888.
- [11] Kuller, L.H., Tracy, R., Belloso, W., Wit, S.D., Drummond, F., Lane, H.C., Ledergerber, B., Lundgren, J., Neuhaus, J., Nixon, D., et al. 2008 Inflammatory and Coagulation Biomarkers and Mortality in Patients with HIV Infection. *PLoS Med* **5**, e203. (doi:10.1371/journal.pmed.0050203).
- [12] Paiardini, M. & Müller-Trutwin, M. 2013 HIV-associated chronic immune activation. *Immunological Reviews* **254**, 78-101. (doi:10.1111/imr.12079).

- [13] Liovat, A.S., Rey-Cuille, M.A., Lecuroux, C., Jacquelin, B., Girault, I., Petitjean, G., Zitoun, Y., Venet, A., Barre-Sinoussi, F., Lebon, P., et al. 2012 Acute plasma biomarkers of T cell activation set-point levels and of disease progression in HIV-1 infection. *PLoS ONE* 7, e46143. (doi:10.1371/journal.pone.0046143).
- [14] Omer, F.M., de Souza, J.B. & Riley, E.M. 2003 Differential induction of TGF-beta regulates proinflammatory cytokine production and determines the outcome of lethal and nonlethal Plasmodium yoelii infections. *J Immunol* **171**, 5430-5436.
- [15] Zuniga, E.I., Liou, L.Y., Mack, L., Mendoza, M. & Oldstone, M.B. 2008 Persistent virus infection inhibits type I interferon production by plasmacytoid dendritic cells to facilitate opportunistic infections. *Cell Host Microbe* **4**, 374-386.
- [16] Sandler, N.G. & Sereti, I. 2014 Can early therapy reduce inflammation? *Current opinion in HIV and AIDS* **9**, 72-79. (doi:10.1097/COH.0000000000000000).
- [17] Saez-Cirion, A., Bacchus, C., Hocqueloux, L., Avettand-Fenoel, V., Girault, I., Lecuroux, C., Potard, V., Versmisse, P., Melard, A., Prazuck, T., et al. 2013 Post-treatment HIV-1 controllers with a long-term virological remission after the interruption of early initiated antiretroviral therapy ANRS VISCONTI Study. *PLoS Pathog* **9**, e1003211. (doi:10.1371/journal.ppat.1003211

PPATHOGENS-D-12-02229 [pii]).

- [18] Hocqueloux, L., Prazuck, T., Avettand-Fenoel, V., Lafeuillade, A., Cardon, B., Viard, J.P. & Rouzioux, C. 2010 Long- term immunovirologic control following antiretroviral therapy interruption in patients treated at the time of primary HIV-1 infection. *Aids* **24**, 1598-1601.
- [19] Lodi, S., Meyer, L., Kelleher, A.D., Rosinska, M., Ghosn, J., Sannes, M. & Porter, K. 2012 Immunovirologic control 24 months after interruption of antiretroviral therapy initiated close to HIV seroconversion. *Archives of internal medicine* **172**, 1252-1255. (doi:10.1001/archinternmed.2012.2719).
- [20] Goujard, C., Emilie, D., Roussillon, C., Godot, V., Rouzioux, C., Venet, A., Colin, C., Pialoux, G., Girard, P.M., Boilet, V., et al. 2012 Continuous versus intermittent treatment strategies during primary HIV-1 infection: the randomized ANRS INTERPRIM Trial. *Aids* **26**, 1895-1905. (doi:10.1097/QAD.0b013e32835844d9).
- [21] Madec, Y., Boufassa, F., Rouzioux, C., Delfraissy, J.F. & Meyer, L. 2005 Undetectable viremia without antiretroviral therapy in patients with HIV seroconversion: an uncommon phenomenon? *Clin Infect Dis* **40**, 1350-1354. (doi:10.1086/429318).
- [22] Goujard, C., Chaix, M.L., Lambotte, O., Deveau, C., Sinet, M., Guergnon, J., Courgnaud, V., Rouzioux, C., Delfraissy, J.F., Venet, A., et al. 2009 Spontaneous control of viral replication during primary HIV infection: when is "HIV controller" status established? *Clin Infect Dis* **49**, 982-986. (doi:10.1086/605504).
- [23] Van Gulck, E., Bracke, L., Heyndrickx, L., Coppens, S., Atkinson, D., Merlin, C., Pasternak, A., Florence, E. & Vanham, G. 2012 Immune and viral correlates of "secondary viral control" after treatment interruption in chronically HIV-1 infected patients. *PLoS ONE* 7, e37792. (doi:10.1371/journal.pone.0037792).
- [24] El-Sadr, W.M., Lundgren, J., Neaton, J.D., Gordin, F., Abrams, D., Arduino, R.C., Babiker, A., Burman, W., Clumeck, N., Cohen, C.J., et al. 2006 CD4+ count-guided interruption of antiretroviral treatment. *N Engl J Med* **355**, 2283-2296. (doi:10.1056/NEJMoa062360).
- [25] Daniel, M.D., Kirchhoff, F., Czajak, S.C., Sehgal, P.K. & Desrosiers, R.C. 1992 Protective effects of a live attenuated SIV vaccine with a deletion in the nef gene. *Science* **258**, 1938-1941.
- [26] Schindler, M., Munch, J., Kutsch, O., Li, H., Santiago, M.L., Bibollet-Ruche, F., Muller-Trutwin, M.C., Novembre, F.J., Peeters, M., Courgnaud, V., et al. 2006 Nef-mediated suppression of T cell activation was lost in a lentiviral lineage that gave rise to HIV-1. *Cell* 125, 1055-1067.
- [27] Li, Q., Duan, L., Estes, J.D., Ma, Z.M., Rourke, T., Wang, Y., Reilly, C., Carlis, J., Miller, C.J. & Haase, A.T. 2005 Peak SIV replication in resting memory CD4+ T cells depletes gut lamina propria CD4+ T cells. *Nature* **434**, 1148-1152.

- [28] Mattapallil, J.J., Douek, D.C., Hill, B., Nishimura, Y., Martin, M. & Roederer, M. 2005 Massive infection and loss of memory CD4+ T cells in multiple tissues during acute SIV infection. *Nature* **434**, 1093-1097.
- [29] Picker, L.J., Hagen, S.I., Lum, R., Reed-Inderbitzin, E.F., Daly, L.M., Sylwester, A.W., Walker, J.M., Siess, D.C., Piatak, M., Jr., Wang, C., et al. 2004 Insufficient production and tissue delivery of CD4+ memory T cells in rapidly progressive simian immunodeficiency virus infection. *J Exp Med* **200**, 1299-1314.
- [30] Veazey, R.S. & Lackner, A.A. 2005 HIV swiftly guts the immune system. *Nat Med* 11, 469-470.
- [31] Brenchley, J.M., Price, D.A., Schacker, T.W., Asher, T.E., Silvestri, G., Rao, S., Kazzaz, Z., Bornstein, E., Lambotte, O., Altmann, D., et al. 2006 Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Nat Med* **12**, 1365-1371.
- [32] Reeves, R.K., Evans, T.I., Gillis, J. & Johnson, R.P. 2010 Simian immunodeficiency virus infection induces expansion of alpha4beta7+ and cytotoxic CD56+ NK cells. *J Virol* **84**, 8959-8963. (doi:10.1128/JVI.01126-10).
- [33] Reeves, R.K., Evans, T.I., Gillis, J., Wong, F.E., Kang, G., Li, Q. & Johnson, R.P. 2012 SIV Infection Induces Accumulation of Plasmacytoid Dendritic Cells in the Gut Mucosa. *Journal of Infectious Diseases* **206**, 1462-1468. (doi:10.1093/infdis/jis408).
- [34] Ansari, A.A., Reimann, K.A., Mayne, A.E., Takahashi, Y., Stephenson, S.T., Wang, R., Wang, X., Li, J., Price, A.A., Little, D.M., et al. 2011 Blocking of alpha4beta7 gut-homing integrin during acute infection leads to decreased plasma and gastrointestinal tissue viral loads in simian immunodeficiency virus-infected rhesus macaques. *J Immunol* **186**, 1044-1059.
- [35] Aarnink, A., Dereuddre-Bosquet, N., Vaslin, B., Le Grand, R., Winterton, P., Apoil, P.A. & Blancher, A. 2011 Influence of the MHC genotype on the progression of experimental SIV infection in the Mauritian cynomolgus macaque. *Immunogenetics* **63**, 267-274. (doi:10.1007/s00251-010-0504-6).
- [36] Karl, J.A., Bohn, P.S., Wiseman, R.W., Nimityongskul, F.A., Lank, S.M., Starrett, G.J. & O'Connor, D.H. 2013 Major histocompatibility complex class I haplotype diversity in Chinese rhesus macaques. *G3* (*Bethesda*) **3**, 1195-1201. (doi:10.1534/g3.113.006254).
- [37] Wu, F., Kirmaier, A., Goeken, R., Ourmanov, I., Hall, L., Morgan, J.S., Matsuda, K., Buckler-White, A., Tomioka, K., Plishka, R., et al. 2013 TRIM5 alpha drives SIVsmm evolution in rhesus macaques. *PLoS Pathog* **9**, e1003577. (doi:10.1371/journal.ppat.1003577).
- [38] Goldstein, S., Ourmanov, I., Brown, C.R., Plishka, R., Buckler-White, A., Byrum, R. & Hirsch, V.M. 2005 Plateau levels of viremia correlate with the degree of CD4+-T-cell loss in simian immunodeficiency virus SIVagm-infected pigtailed macaques: variable pathogenicity of natural SIVagm isolates. *J Virol* **79**, 5153-5162.
- [39] Pandrea, I., Gaufin, T., Gautam, R., Kristoff, J., Mandell, D., Montefiori, D., Keele, B.F., Ribeiro, R.M., Veazey, R.S. & Apetrei, C. 2011 Functional cure of SIVagm infection in rhesus macaques results in complete recovery of CD4+ T cells and is reverted by CD8+ cell depletion. *PLoS Pathog* **7**, e1002170. (doi:10.1371/journal.ppat.1002170).
- [40] Diop, O.M., Gueye, A., Dias-Tavares, M., Kornfeld, C., Faye, A., Ave, P., Huerre, M., Corbet, S., Barre-Sinoussi, F. & Muller-Trutwin, M.C. 2000 High levels of viral replication during primary simian immunodeficiency virus SIVagm infection are rapidly and strongly controlled in African green monkeys. *J Virol* **74**, 7538-7547.
- [41] Bourry, O., Mannioui, A., Sellier, P., Roucairol, C., Durand-Gasselin, L., Dereuddre-Bosquet, N., Benech, H., Roques, P. & Le Grand, R. 2010 Effect of a short-term HAART on SIV load in macaque tissues is dependent on time of initiation and antiviral diffusion. *Retrovirology* **7**, 78. (doi:10.1186/1742-4690-7-78).
- [42] Cranage, M., Sharpe, S., Herrera, C., Cope, A., Dennis, M., Berry, N., Ham, C., Heeney, J., Rezk, N., Kashuba, A., et al. 2008 Prevention of SIV rectal transmission and priming of T cell responses in macaques after local pre-exposure application of tenofovir gel. *PLoS Med* **5**, e157; discussion e157. (doi:10.1371/journal.pmed.0050157).

- [43] Apetrei, C., Pandrea, I. & Mellors, J.W. 2012 Nonhuman primate models for HIV cure research. *PLoS Pathog* **8**, e1002892. (doi:10.1371/journal.ppat.1002892).
- [44] Shytaj, I.L., Norelli, S., Chirullo, B., Della Corte, A., Collins, M., Yalley-Ogunro, J., Greenhouse, J., Iraci, N., Acosta, E.P., Barreca, M.L., et al. 2012 A highly intensified ART regimen induces long-term viral suppression and restriction of the viral reservoir in a simian AIDS model. *PLoS Pathog* **8**, e1002774. (doi:10.1371/journal.ppat.1002774).
- [45] Sodora, D.L., Allan, J.S., Apetrei, C., Brenchley, J.M., Douek, D.C., Else, J.G., Estes, J.D., Hahn, B.H., Hirsch, V.M., Kaur, A., et al. 2009 Toward an AIDS vaccine: lessons from natural simian immunodeficiency virus infections of African nonhuman primate hosts. *Nat Med* **15**, 861-865.
- [46] Liovat, A.S., Jacquelin, B., Ploquin, M.J., Barre-Sinoussi, F. & Muller-Trutwin, M.C. 2009 African non human primates infected by SIV why don't they get sick? Lessons from studies on the early phase of non-pathogenic SIV infection. *Curr HIV Res* **7**, 39-50.
- [47] Kornfeld, C., Ploquin, M.J., Pandrea, I., Faye, A., Onanga, R., Apetrei, C., Poaty-Mavoungou, V., Rouquet, P., Estaquier, J., Mortara, L., et al. 2005 Antiinflammatory profiles during primary SIV infection in African green monkeys are associated with protection against AIDS. *J Clin Invest* **115**, 1082-1091.
- [48] Douek, D.C. 2013 Immune activation, HIV persistence, and the cure. *Topics in antiviral medicine* **21**, 128-132.
- [49] Boulet, S., Sharafi, S., Simic, N., Bruneau, J., Routy, J.P., Tsoukas, C.M. & Bernard, N.F. 2008 Increased proportion of KIR3DS1 homozygotes in HIV-exposed uninfected individuals. *AIDS* **22**, 595-599. (doi:10.1097/QAD.0b013e3282f56b23 00002030-200803120-00005 [pii]).
- [50] Jennes, W., Verheyden, S., Demanet, C., Adje-Toure, C.A., Vuylsteke, B., Nkengasong, J.N. & Kestens, L. 2006 Cutting edge: resistance to HIV-1 infection among African female sex workers is associated with inhibitory KIR in the absence of their HLA ligands. *J Immunol* **177**, 6588-6592. (doi:177/10/6588 [pii]).
- [51] Montoya, C.J., Velilla, P.A., Chougnet, C., Landay, A.L. & Rugeles, M.T. 2006 Increased IFN-gamma production by NK and CD3+/CD56+ cells in sexually HIV-1-exposed but uninfected individuals. *Clin Immunol* **120**, 138-146. (doi:S1521-6616(06)00080-5 [pii] 10.1016/j.clim.2006.02.008).
- [52] Ravet, S., Scott-Algara, D., Bonnet, E., Tran, H.K., Tran, T., Nguyen, N., Truong, L.X., Theodorou, I., Barre-Sinoussi, F., Pancino, G., et al. 2007 Distinctive NK-cell receptor repertoires sustain high-level constitutive NK-cell activation in HIV-exposed uninfected individuals. *Blood* **109**, 4296-4305. (doi:blood-2006-08-040238 [pii] 10.1182/blood-2006-08-040238).
- [53] Scott-Algara, D., Truong, L.X., Versmisse, P., David, A., Luong, T.T., Nguyen, N.V., Theodorou, I., Barre-Sinoussi, F. & Pancino, G. 2003 Cutting edge: increased NK cell activity in HIV-1-exposed but uninfected Vietnamese intravascular drug users. *J Immunol* **171**, 5663-5667.
- [54] Alter, G., Teigen, N., Ahern, R., Streeck, H., Meier, A., Rosenberg, E.S. & Altfeld, M. 2007 Evolution of innate and adaptive effector cell functions during acute HIV-1 infection. *J Infect Dis* **195**, 1452-1460. (doi:JID37665 [pii] 10.1086/513878).
- [55] Alter, G., Teigen, N., Davis, B.T., Addo, M.M., Suscovich, T.J., Waring, M.T., Streeck, H., Johnston, M.N., Staller, K.D., Zaman, M.T., et al. 2005 Sequential deregulation of NK cell subset distribution and function starting in acute HIV-1 infection. *Blood* **106**, 3366-3369. (doi:2005-03-1100 [pii]
- 10.1182/blood-2005-03-1100).
- [56] Jost, S. & Altfeld, M. 2012 Evasion from NK cell-mediated immune responses by HIV-1. *Microbes Infect* **14**, 904-915. (doi:10.1016/j.micinf.2012.05.001 S1286-4579(12)00119-0 [pii]).
- [57] Martin, M.P., Gao, X., Lee, J.H., Nelson, G.W., Detels, R., Goedert, J.J., Buchbinder, S., Hoots, K., Vlahov, D., Trowsdale, J., et al. 2002 Epistatic interaction between KIR3DS1 and HLA-B delays the progression to AIDS. *Nat Genet* **31**, 429-434. (doi:10.1038/ng934

ng934 [pii]).

[58] Martin, M.P., Qi, Y., Gao, X., Yamada, E., Martin, J.N., Pereyra, F., Colombo, S., Brown, E.E., Shupert, W.L., Phair, J., et al. 2007 Innate partnership of HLA-B and KIR3DL1 subtypes against HIV-1. *Nat Genet* **39**, 733-740. (doi:ng2035 [pii] 10.1038/ng2035).

[59] Alter, G., Rihn, S., Walter, K., Nolting, A., Martin, M., Rosenberg, E.S., Miller, J.S., Carrington, M. & Altfeld, M. 2009 HLA class I subtype-dependent expansion of KIR3DS1+ and KIR3DL1+ NK cells during acute human immunodeficiency virus type 1 infection. *J Virol* 83, 6798-6805. (doi:10.1128/JVI.00256-09 JVI.00256-09 [pii]).

[60] Kim, S., Sunwoo, J.B., Yang, L., Choi, T., Song, Y.J., French, A.R., Vlahiotis, A., Piccirillo, J.F., Cella, M., Colonna, M., et al. 2008 HLA alleles determine differences in human natural killer cell responsiveness and potency. *Proc Natl Acad Sci U S A* **105**, 3053-3058. (doi:10.1073/pnas.0712229105 0712229105 [pii]).

[61] Fadda, L., Borhis, G., Ahmed, P., Cheent, K., Pageon, S.V., Cazaly, A., Stathopoulos, S., Middleton, D., Mulder, A., Claas, F.H., et al. 2010 Peptide antagonism as a mechanism for NK cell activation. *Proc Natl Acad Sci U S A* **107**, 10160-10165. (doi:10.1073/pnas.0913745107

0913745107 [pii]).

[62] Thananchai, H., Gillespie, G., Martin, M.P., Bashirova, A., Yawata, N., Yawata, M., Easterbrook, P., McVicar, D.W., Maenaka, K., Parham, P., et al. 2007 Cutting Edge: Allelespecific and peptide-dependent interactions between KIR3DL1 and HLA-A and HLA-B. *J Immunol* 178, 33-37. (doi:178/1/33 [pii]).

[63] Vivian, J.P., Duncan, R.C., Berry, R., O'Connor, G.M., Reid, H.H., Beddoe, T., Gras, S., Saunders, P.M., Olshina, M.A., Widjaja, J.M., et al. 2011 Killer cell immunoglobulin-like receptor 3DL1-mediated recognition of human leukocyte antigen B. *Nature* **479**, 401-405. (doi:10.1038/nature10517

nature10517 [pii]).

[64] Alter, G., Heckerman, D., Schneidewind, A., Fadda, L., Kadie, C.M., Carlson, J.M., Oniangue-Ndza, C., Martin, M., Li, B., Khakoo, S.I., et al. 2011 HIV-1 adaptation to NK-cell-mediated immune pressure. *Nature* **476**, 96-100. (doi:10.1038/nature10237 nature10237 [pii]).

[65] Migueles, S.A., Sabbaghian, M.S., Shupert, W.L., Bettinotti, M.P., Marincola, F.M., Martino, L., Hallahan, C.W., Selig, S.M., Schwartz, D., Sullivan, J., et al. 2000 HLA B*5701 is highly associated with restriction of virus replication in a subgroup of HIV-infected long term nonprogressors. *Proc Natl Acad Sci U S A* **97**, 2709-2714. (doi:10.1073/pnas.050567397 050567397 [pii]).

[66] Pereyra, F. & Jia, X. & McLaren, P.J. & Telenti, A. & de Bakker, P.I. & Walker, B.D. & Ripke, S. & Brumme, C.J. & Pulit, S.L. & Carrington, M., et al. 2010 The major genetic determinants of HIV-1 control affect HLA class I peptide presentation. *Science* **330**, 1551-1557. (doi:10.1126/science.1195271

science.1195271 [pii]).

[67] Saez-Cirion, A., Lacabaratz, C., Lambotte, O., Versmisse, P., Urrutia, A., Boufassa, F., Barre-Sinoussi, F., Delfraissy, J.F., Sinet, M., Pancino, G., et al. 2007 HIV controllers exhibit potent CD8 T cell capacity to suppress HIV infection ex vivo and peculiar cytotoxic T lymphocyte activation phenotype. *Proc Natl Acad Sci U S A* **104**, 6776-6781. (doi:0611244104 [pii]

10.1073/pnas.0611244104).

[68] Flores-Villanueva, P.O., Yunis, E.J., Delgado, J.C., Vittinghoff, E., Buchbinder, S., Leung, J.Y., Uglialoro, A.M., Clavijo, O.P., Rosenberg, E.S., Kalams, S.A., et al. 2001 Control of HIV-1 viremia and protection from AIDS are associated with HLA-Bw4 homozygosity. *Proc Natl Acad Sci U S A* **98**, 5140-5145. (doi:10.1073/pnas.071548198 071548198 [pii]).

- [69] Marras, F., Nicco, E., Bozzano, F., Di Biagio, A., Dentone, C., Pontali, E., Boni, S., Setti, M., Orofino, G., Mantia, E., et al. 2013 Natural killer cells in HIV controller patients express an activated effector phenotype and do not up-regulate NKp44 on IL-2 stimulation. *Proc Natl Acad Sci U S A* **110**, 11970-11975. (doi:10.1073/pnas.1302090110 1302090110 [pii]).
- [70] Tomescu, C., Duh, F.M., Hoh, R., Viviani, A., Harvill, K., Martin, M.P., Carrington, M., Deeks, S.G. & Montaner, L.J. 2012 Impact of protective killer inhibitory receptor/human leukocyte antigen genotypes on natural killer cell and T-cell function in HIV-1-infected controllers. *AIDS* **26**, 1869-1878. (doi:10.1097/QAD.0b013e32835861b0).
- [71] Vieillard, V., Fausther-Bovendo, H., Samri, A. & Debre, P. 2010 Specific phenotypic and functional features of natural killer cells from HIV-infected long-term nonprogressors and HIV controllers. *J Acquir Immune Defic Syndr* **53**, 564-573. (doi:10.1097/QAI.0b013e3181d0c5b4).
- [72] O'Connell, K.A., Han, Y., Williams, T.M., Siliciano, R.F. & Blankson, J.N. 2009 Role of natural killer cells in a cohort of elite suppressors: low frequency of the protective KIR3DS1 allele and limited inhibition of human immunodeficiency virus type 1 replication in vitro. *J Virol* **83**, 5028-5034.
- [73] Muller-Trutwin, M. & Hosmalin, A. 2005 Role for plasmacytoid dendritic cells in anti-HIV innate immunity. *Immunol Cell Biol* **83**, 578-583. (doi:ICB1394 [pii] 10.1111/j.1440-1711.2005.01394.x).
- [74] Pillai, S.K., Abdel-Mohsen, M., Guatelli, J., Skasko, M., Monto, A., Fujimoto, K., Yukl, S., Greene, W.C., Kovari, H., Rauch, A., et al. 2012 Role of retroviral restriction factors in the interferon-alpha-mediated suppression of HIV-1 in vivo. *Proc Natl Acad Sci U S A* **109**, 3035-3040. (doi:10.1073/pnas.1111573109 [pii]).
- [75] Poli, G., Orenstein, J.M., Kinter, A., Folks, T.M. & Fauci, A.S. 1989 Interferon-alpha but not AZT suppresses HIV expression in chronically infected cell lines. *Science* **244**, 575-577. [76] Torriani, F.J., Rodriguez-Torres, M., Rockstroh, J.K., Lissen, E., Gonzalez-Garcia, J., Lazzarin, A., Carosi, G., Sasadeusz, J., Katlama, C., Montaner, J., et al. 2004 Peginterferon Alfa-2a plus ribavirin for chronic hepatitis C virus infection in HIV-infected patients. *N Engl J Med* **351**, 438-450. (doi:10.1056/NEJMoa040842 351/5/438 [pii]).
- [77] Yamamoto, J.K., Barre-Sinoussi, F., Bolton, V., Pedersen, N.C. & Gardner, M.B. 1986 Human alpha- and beta-interferon but not gamma- suppress the in vitro replication of LAV, HTLV-III, and ARV-2. *Journal of interferon research* **6**, 143-152.
- [78] Berger, A., Sommer, A.F., Zwarg, J., Hamdorf, M., Welzel, K., Esly, N., Panitz, S., Reuter, A., Ramos, I., Jatiani, A., et al. 2011 SAMHD1-deficient CD14+ cells from individuals with Aicardi-Goutieres syndrome are highly susceptible to HIV-1 infection. *PLoS Pathog* 7, e1002425. (doi:10.1371/journal.ppat.1002425 PPATHOGENS-D-11-01498 [pii]).
- [79] Carthagena, L., Bergamaschi, A., Luna, J.M., David, A., Uchil, P.D., Margottin-Goguet, F., Mothes, W., Hazan, U., Transy, C., Pancino, G., et al. 2009 Human TRIM gene expression in response to interferons. *PLoS One* **4**, e4894. (doi:10.1371/journal.pone.0004894).
- [80] Goujon, C., Moncorge, O., Bauby, H., Doyle, T., Ward, C.C., Schaller, T., Hue, S., Barclay, W.S., Schulz, R. & Malim, M.H. 2013 Human MX2 is an interferon-induced postentry inhibitor of HIV-1 infection. *Nature* **502**, 559-562. (doi:10.1038/nature12542 nature12542 [pii]).
- [81] Kane, M., Yadav, S.S., Bitzegeio, J., Kutluay, S.B., Zang, T., Wilson, S.J., Schoggins, J.W., Rice, C.M., Yamashita, M., Hatziioannou, T., et al. 2013 MX2 is an interferon-induced inhibitor of HIV-1 infection. *Nature* **502**, 563-566. (doi:10.1038/nature12653 nature12653 [pii]).
- [82] Keele, B.F., Giorgi, E.E., Salazar-Gonzalez, J.F., Decker, J.M., Pham, K.T., Salazar, M.G., Sun, C., Grayson, T., Wang, S., Li, H., et al. 2008 Identification and characterization of

transmitted and early founder virus envelopes in primary HIV-1 infection. *Proc Natl Acad Sci U S A* **105**, 7552-7557. (doi:10.1073/pnas.0802203105 0802203105 [pii]).

[83] Fenton-May, A.E., Dibben, O., Emmerich, T., Ding, H., Pfafferott, K., Aasa-Chapman, M.M., Pellegrino, P., Williams, I., Cohen, M.S., Gao, F., et al. 2013 Relative resistance of HIV-1 founder viruses to control by interferon-alpha. *Retrovirology* **10**, 146. (doi:10.1186/1742-4690-10-

146 1742-4690-10-146 [pii]).

- [84] Parrish, N.F., Gao, F., Li, H., Giorgi, E.E., Barbian, H.J., Parrish, E.H., Zajic, L., Iyer, S.S., Decker, J.M., Kumar, A., et al. 2013 Phenotypic properties of transmitted founder HIV-1. *Proc Natl Acad Sci U S A* **110**, 6626-6633. (doi:10.1073/pnas.1304288110 1304288110 [pii]).
- [85] Seo, Y.J. & Hahm, B. 2010 Type I interferon modulates the battle of host immune system against viruses. *Adv Appl Microbiol* **73**, 83-101. (doi:10.1016/S0065-2164(10)73004-5

S0065-2164(10)73004-5 [pii]).

- [86] Haase, A.T. 2010 Targeting early infection to prevent HIV-1 mucosal transmission. *Nature* **464**, 217-223. (doi:10.1038/nature08757 nature08757 [pii]).
- [87] Favre, D., Mold, J., Hunt, P.W., Kanwar, B., Loke, P., Seu, L., Barbour, J.D., Lowe, M.M., Jayawardene, A., Aweeka, F., et al. 2010 Tryptophan catabolism by indoleamine 2,3-dioxygenase 1 alters the balance of TH17 to regulatory T cells in HIV disease. *Sci Transl Med* 2, 32ra36. (doi:10.1126/scitranslmed.3000632 2/32/32ra36 [pii]).
- [88] Herbeuval, J.P., Nilsson, J., Boasso, A., Hardy, A.W., Kruhlak, M.J., Anderson, S.A., Dolan, M.J., Dy, M., Andersson, J. & Shearer, G.M. 2006 Differential expression of IFN-alpha and TRAIL/DR5 in lymphoid tissue of progressor versus nonprogressor HIV-1-infected patients. *Proc Natl Acad Sci U S A* **103**, 7000-7005. (doi:0600363103 [pii] 10.1073/pnas.0600363103).
- [89] Malleret, B., Maneglier, B., Karlsson, I., Lebon, P., Nascimbeni, M., Perie, L., Brochard, P., Delache, B., Calvo, J., Andrieu, T., et al. 2008 Primary infection with simian immunodeficiency virus: plasmacytoid dendritic cell homing to lymph nodes, type I interferon, and immune suppression. *Blood* 112, 4598-4608. (doi:10.1182/blood-2008-06-162651 blood-2008-06-162651 [pii]).
- [90] Barblu, L., Machmach, K., Gras, C., Delfraissy, J.F., Boufassa, F., Leal, M., Ruiz-Mateos, E., Lambotte, O. & Herbeuval, J.P. 2012 Plasmacytoid dendritic cells (pDCs) from HIV controllers produce interferon-alpha and differentiate into functional killer pDCs under HIV activation. *J Infect Dis* **206**, 790-801. (doi:jis384 [pii] 10.1093/infdis/jis384).
- [91] Machmach, K., Leal, M., Gras, C., Viciana, P., Genebat, M., Franco, E., Boufassa, F., Lambotte, O., Herbeuval, J.P. & Ruiz-Mateos, E. 2012 Plasmacytoid dendritic cells reduce HIV production in elite controllers. *J Virol* **86**, 4245-4252. (doi:10.1128/JVI.07114-11 JVI.07114-11 [pii]).
- [92] Bergamaschi, A., David, A., Le Rouzic, E., Nisole, S., Barre-Sinoussi, F. & Pancino, G. 2009 The CDK inhibitor p21Cip1/WAF1 is induced by FcgammaR activation and restricts the replication of human immunodeficiency virus type 1 and related primate lentiviruses in human macrophages. *J Virol* 83, 12253-12265. (doi:10.1128/JVI.01395-09 JVI.01395-09 [pii]).
- [93] Krupp, A., McCarthy, K.R., Ooms, M., Letko, M., Morgan, J.S., Simon, V. & Johnson, W.E. 2013 APOBEC3G polymorphism as a selective barrier to cross-species transmission and emergence of pathogenic SIV and AIDS in a primate host. *PLoS Pathog* **9**, e1003641. (doi:10.1371/journal.ppat.1003641

PPATHOGENS-D-13-01187 [pii]).

[94] Malim, M.H. & Bieniasz, P.D. 2012 HIV Restriction Factors and Mechanisms of Evasion. *Cold Spring Harb Perspect Med* **2**, a006940. (doi:10.1101/cshperspect.a006940

a006940 [pii]).

[95] van Manen, D., Rits, M.A., Beugeling, C., van Dort, K., Schuitemaker, H. & Kootstra, N.A. 2008 The effect of Trim5 polymorphisms on the clinical course of HIV-1 infection. *PLoS Pathog* **4**, e18. (doi:10.1371/journal.ppat.0040018 07-PLPA-RA-0431 [pii]).

[96] De Pasquale, M., Kourteva, Y., Allos, T. & D'Aquila, R.T. 2013 Lower HIV Provirus Levels Are Associated with More APOBEC3G Protein in Blood Resting Memory CD4+ T Lymphocytes of Controllers In Vivo. *PLoS One* **8**, e76002.

(doi:10.1371/journal.pone.0076002

PONE-D-13-23416 [pii]).

[97] Do, H., Vasilescu, A., Diop, G., Hirtzig, T., Heath, S.C., Coulonges, C., Rappaport, J., Therwath, A., Lathrop, M., Matsuda, F., et al. 2005 Exhaustive genotyping of the CEM15 (APOBEC3G) gene and absence of association with AIDS progression in a French cohort. *J Infect Dis* **191**, 159-163. (doi:JID33034 [pii] 10.1086/426826).

[98] Sewram, S., Singh, R., Kormuth, E., Werner, L., Mlisana, K., Karim, S.S. & Ndung'u, T. 2009 Human TRIM5alpha expression levels and reduced susceptibility to HIV-1 infection. *J Infect Dis* **199**, 1657-1663. (doi:10.1086/598861).

[99] Rotger, M., Dang, K.K., Fellay, J., Heinzen, E.L., Feng, S., Descombes, P., Shianna, K.V., Ge, D., Gunthard, H.F., Goldstein, D.B., et al. 2010 Genome-wide mRNA expression correlates of viral control in CD4+ T-cells from HIV-1-infected individuals. *PLoS Pathog* **6**, e1000781. (doi:10.1371/journal.ppat.1000781).

[100] Paxton, W.A., Martin, S.R., Tse, D., O'Brien, T.R., Skurnick, J., VanDevanter, N.L., Padian, N., Braun, J.F., Kotler, D.P., Wolinsky, S.M., et al. 1996 Relative resistance to HIV-1 infection of CD4 lymphocytes from persons who remain uninfected despite multiple high-risk sexual exposure. *Nat Med* **2**, 412-417.

[101] Saez-Cirion, A., Versmisse, P., Truong, L.X., Chakrabarti, L.A., Carpentier, W., Barre-Sinoussi, F., Scott-Algara, D. & Pancino, G. 2006 Persistent resistance to HIV-1 infection in CD4 T cells from exposed uninfected Vietnamese individuals is mediated by entry and post-entry blocks. *Retrovirology* **3**, 81. (doi:1742-4690-3-81 [pii] 10.1186/1742-4690-3-81).

[102] Chen, H., Li, C., Huang, J., Cung, T., Seiss, K., Beamon, J., Carrington, M.F., Porter, L.C., Burke, P.S., Yang, Y., et al. 2011 CD4+ T cells from elite controllers resist HIV-1 infection by selective upregulation of p21. *J Clin Invest* **121**, 1549-1560. (doi:10.1172/JCl44539 [pii]).

[103] Saez-Cirion, A., Hamimi, C., Bergamaschi, A., David, A., Versmisse, P., Melard, A., Boufassa, F., Barre-Sinoussi, F., Lambotte, O., Rouzioux, C., et al. 2011 Restriction of HIV-1 replication in macrophages and CD4+ T cells from HIV controllers. *Blood* **118**, 955-964. (doi:10.1182/blood-2010-12-327106

blood-2010-12-327106 [pii]).

[104] Koup, R.A., Safrit, J.T., Cao, Y., Andrews, C.A., McLeod, G., Borkowsky, W., Farthing, C. & Ho, D.D. 1994 Temporal association of cellular immune responses with the initial control of viremia in primary human immunodeficiency virus type 1 syndrome. *J Virol* **68**, 4650-4655.

[105] Borrow, P., Lewicki, H., Wei, X., Horwitz, M.S., Peffer, N., Meyers, H., Nelson, J.A., Gairin, J.E., Hahn, B.H., Oldstone, M.B., et al. 1997 Antiviral pressure exerted by HIV-1-specific cytotoxic T lymphocytes (CTLs) during primary infection demonstrated by rapid selection of CTL escape virus. *Nat Med* 3, 205-211.

[106] Kiepiela, P., Ngumbela, K., Thobakgale, C., Ramduth, D., Honeyborne, I., Moodley, E., Reddy, S., de Pierres, C., Mncube, Z., Mkhwanazi, N., et al. 2007 CD8+ T-cell responses to different HIV proteins have discordant associations with viral load. *Nat Med* 13, 46-53. (doi:10.1038/nm1520).

[107] Riviere, Y., McChesney, M.B., Porrot, F., Tanneau-Salvadori, F., Sansonetti, P., Lopez, O., Pialoux, G., Feuillie, V., Mollereau, M., Chamaret, S., et al. 1995 Gag-specific cytotoxic

- responses to HIV type 1 are associated with a decreased risk of progression to AIDS-related complex or AIDS. AIDS Res Hum Retroviruses 11, 903-907.
- [108] Martinez-Picado, J., Prado, J.G., Fry, E.E., Pfafferott, K., Leslie, A., Chetty, S., Thobakgale, C., Honeyborne, I., Crawford, H., Matthews, P., et al. 2006 Fitness cost of escape mutations in p24 Gag in association with control of human immunodeficiency virus type 1. *J Virol* **80**, 3617-3623. (doi:10.1128/JVI.80.7.3617-3623.2006).
- [109] Matano, T., Shibata, R., Siemon, C., Connors, M., Lane, H.C. & Martin, M.A. 1998 Administration of an anti-CD8 monoclonal antibody interferes with the clearance of chimeric simian/human immunodeficiency virus during primary infections of rhesus macaques. *J Virol* **72**, 164 169.
- [110] Okoye, A., Park, H., Rohankhedkar, M., Coyne-Johnson, L., Lum, R., Walker, J.M., Planer, S.L., Legasse, A.W., Sylwester, A.W., Piatak, M., et al. 2009 Profound CD4+/CCR5+ T cell expansion is induced by CD8+ lymphocyte depletion but does not account for accelerated SIV pathogenesis. *J Exp Med* **206**, 1575 1588.
- [111] Jin, X., Bauer, D.E., Tuttleton, S.E., Lewin, S., Gettie, A., Blanchard, J., Irwin, C.E., Safrit, J.T., Mittler, J., Weinberger, L., et al. 1999 Dramatic rise in plasma viremia after CD8+ T cell depletion in simian immunodeficiency virus-infected macaques. *J Exp Med* **189**, 991 998.
- [112] Schmitz, J.E., Simon, M.A., Kuroda, M.J., Lifton, M.A., Ollert, M.W., Vogel, C.W., Racz, P., Tenner-Racz, K., Scallon, B.J., Dalesandro, M., et al. 1999 A nonhuman primate model for the selective elimination of CD8+ lymphocytes using a mouse-human chimeric monoclonal antibody. *Am J Pathol* **154**, 1923 1932.
- [113] Cocchi, F., DeVico, A.L., Garzino-Demo, A., Arya, S.K., Gallo, R.C. & Lusso, P. 1995 Identification of RANTES, MIP-1 alpha, and MIP-1 beta as the major HIV-suppressive factors produced by CD8+ T cells. *Science* **270**, 1811-1815.
- [114] Walker, C.M., Moody, D.J., Stites, D.P. & Levy, J.A. 1986 CD8+ lymphocytes can control HIV infection in vitro by suppressing virus replication. *Science* **234**, 1563-1566.
- [115] Kagi, D., Ledermann, B., Burki, K., Hengartner, H. & Zinkernagel, R.M. 1994 CD8+ T cell-mediated protection against an intracellular bacterium by perforin-dependent cytotoxicity. *Eur J Immunol* **24**, 3068-3072. (doi:10.1002/eji.1830241223).
- [116] Lowin, B., Hahne, M., Mattmann, C. & Tschopp, J. 1994 Cytolytic T-cell cytotoxicity is mediated through perforin and Fas lytic pathways. *Nature* **370**, 650-652. (doi:10.1038/370650a0).
- [117] Youngblood, B., Wherry, E.J. & Ahmed, R. 2012 Acquired transcriptional programming in functional and exhausted virus-specific CD8 T cells. *Curr Opin HIV AIDS* **7**, 50-57. (doi:10.1097/COH.0b013e32834ddcf2).
- [118] Champagne, P., Ogg, G.S., King, A.S., Knabenhans, C., Ellefsen, K., Nobile, M., Appay, V., Rizzardi, G.P., Fleury, S., Lipp, M., et al. 2001 Skewed maturation of memory HIV-specific CD8 T lymphocytes. *Nature* **410**, 106-111. (doi:10.1038/35065118 [pii]).
- [119] Papagno, L., Spina, C.A., Marchant, A., Salio, M., Rufer, N., Little, S., Dong, T., Chesney, G., Waters, A., Easterbrook, P., et al. 2004 Immune activation and CD8+ T-cell differentiation towards senescence in HIV-1 infection. *PLoS Biol* **2**, E20. (doi:10.1371/journal.pbio.0020020).
- [120] Gea-Banacloche, J.C., Migueles, S.A., Martino, L., Shupert, W.L., McNeil, A.C., Sabbaghian, M.S., Ehler, L., Prussin, C., Stevens, R., Lambert, L., et al. 2000 Maintenance of large numbers of virus-specific CD8+ T cells in HIV-infected progressors and long-term nonprogressors. *J Immunol* **165**, 1082-1092.
- [121] Betts, M.R., Nason, M.C., West, S.M., De Rosa, S.C., Migueles, S.A., Abraham, J., Lederman, M.M., Benito, J.M., Goepfert, P.A., Connors, M., et al. 2006 HIV nonprogressors preferentially maintain highly functional HIV-specific CD8+ T cells. *Blood* **107**, 4781-4789. (doi:10.1182/blood-2005-12-4818).
- [122] Migueles, S.A., Laborico, A.C., Shupert, W.L., Sabbaghian, M.S., Rabin, R., Hallahan, C.W., Van Baarle, D., Kostense, S., Miedema, F., McLaughlin, M., et al. 2002 HIV-specific

CD8+ T cell proliferation is coupled to perforin expression and is maintained in nonprogressors. *Nat Immunol* **3**, 1061-1068. (doi:10.1038/ni845).

[123] Hersperger, A.R., Makedonas, G. & Betts, M.R. 2008 Flow cytometric detection of perforin upregulation in human CD8 T cells. *Cytometry. Part A: the journal of the International Society for Analytical Cytology* **73**, 1050-1057. (doi:10.1002/cyto.a.20596).

[124] Migueles, S.A., Osborne, C.M., Royce, C., Compton, A.A., Joshi, R.P., Weeks, K.A., Rood, J.E., Berkley, A.M., Sacha, J.B., Cogliano-Shutta, N.A., et al. 2008 Lytic granule loading of CD8+ T cells is required for HIV-infected cell elimination associated with immune control. *Immunity* **29**, 1009-1021. (doi:10.1016/j.immuni.2008.10.010).

[125] Saez-Cirion, A., Sinet, M., Shin, S.Y., Urrutia, A., Versmisse, P., Lacabaratz, C., Boufassa, F., Avettand-Fenoel, V., Rouzioux, C., Delfraissy, J.F., et al. 2009 Heterogeneity in HIV suppression by CD8 T cells from HIV controllers: association with Gag-specific CD8 T cell responses. *J Immunol* **182**, 7828-7837. (doi:10.4049/jimmunol.0803928 182/12/7828 [pii]).

[126] Sacha, J.B., Chung, C., Rakasz, E.G., Spencer, S.P., Jonas, A.K., Bean, A.T., Lee, W., Burwitz, B.J., Stephany, J.J., Loffredo, J.T., et al. 2007 Gag-specific CD8+ T lymphocytes recognize infected cells before AIDS-virus integration and viral protein expression. *J Immunol* **178**, 2746-2754.

[127] Lecuroux, C., Girault, I., Cheret, A., Versmisse, P., Nembot, G., Meyer, L., Rouzioux, C., Pancino, G., Venet, A. & Saez-Cirion, A. 2013 CD8 T-cells from most HIV-infected patients lack ex vivo HIV-suppressive capacity during acute and early infection. *PLoS One* **8**, e59767. (doi:10.1371/journal.pone.0059767 PONE-D-12-36880 [pii]).

[128] Trautmann, L., Mbitikon-Kobo, F.M., Goulet, J.P., Peretz, Y., Shi, Y., Van Grevenynghe, J., Procopio, F.A., Boulassel, M.R., Routy, J.P., Chomont, N., et al. 2012 Profound metabolic, functional, and cytolytic differences characterize HIV-specific CD8 T cells in primary and chronic HIV infection. *Blood* **120**, 3466-3477. (doi:10.1182/blood-2012-04-422550

blood-2012-04-422550 [pii]).

[129] Ladell, K., Hashimoto, M., Iglesias, M.C., Wilmann, P.G., McLaren, J.E., Gras, S., Chikata, T., Kuse, N., Fastenackels, S., Gostick, E., et al. 2013 A molecular basis for the control of preimmune escape variants by HIV-specific CD8+ T cells. *Immunity* **38**, 425-436. (doi:10.1016/j.immuni.2012.11.021

S1074-7613(13)00106-4 [pii]).

[130] Chen, H., Ndhlovu, Z.M., Liu, D., Porter, L.C., Fang, J.W., Darko, S., Brockman, M.A., Miura, T., Brumme, Z.L., Schneidewind, A., et al. 2012 TCR clonotypes modulate the protective effect of HLA class I molecules in HIV-1 infection. *Nat Immunol* **13**, 691-700. (doi:10.1038/ni.2342

ni.2342 [pii]).

[131] Huang, J., Yang, Y., Al-Mozaini, M., Burke, P.S., Beamon, J., Carrington, M.F., Seiss, K., Rychert, J., Rosenberg, E.S., Lichterfeld, M., et al. 2011 Dendritic cell dysfunction during primary HIV-1 infection. *J Infect Dis* **204**, 1557-1562. (doi:10.1093/infdis/jir616).

[132] Huang, J., Burke, P.S., Cung, T.D., Pereyra, F., Toth, I., Walker, B.D., Borges, L., Lichterfeld, M. & Yu, X.G. 2010 Leukocyte immunoglobulin-like receptors maintain unique antigen-presenting properties of circulating myeloid dendritic cells in HIV-1-infected elite controllers. *J Virol* 84, 9463-9471. (doi:10.1128/JVI.01009-10 JVI.01009-10 [pii]).

[133] Emu, B., Sinclair, E., Hatano, H., Ferre, A., Shacklett, B., Martin, J.N., McCune, J.M. & Deeks, S.G. 2008 HLA class I-restricted T-cell responses may contribute to the control of human immunodeficiency virus infection, but such responses are not always necessary for long-term virus control. *J Virol* 82, 5398-5407. (doi:10.1128/JVI.02176-07).

[134] Pereyra, F., Addo, M.M., Kaufmann, D.E., Liu, Y., Miura, T., Rathod, A., Baker, B., Trocha, A., Rosenberg, R., Mackey, E., et al. 2008 Genetic and immunologic heterogeneity among persons who control HIV infection in the absence of therapy. *J Infect Dis* **197**, 563-571. (doi:10.1086/526786).

- [135] Ndhlovu, Z.M., Proudfoot, J., Cesa, K., Alvino, D.M., McMullen, A., Vine, S., Stampouloglou, E., Piechocka-Trocha, A., Walker, B.D. & Pereyra, F. 2012 Elite controllers with low to absent effector CD8+ T cell responses maintain highly functional, broadly directed central memory responses. *J Virol* **86**, 6959-6969. (doi:10.1128/JVI.00531-12).
- [136] Shan, L., Deng, K., Shroff, N.S., Durand, C.M., Rabi, S.A., Yang, H.C., Zhang, H., Margolick, J.B., Blankson, J.N. & Siliciano, R.F. 2012 Stimulation of HIV-1-specific cytolytic T lymphocytes facilitates elimination of latent viral reservoir after virus reactivation. *Immunity* **36**, 491-501. (doi:10.1016/j.immuni.2012.01.014).
- [137] Douek, D.C., Brenchley, J.M., Betts, M.R., Ambrozak, D.R., Hill, B.J., Okamoto, Y., Casazza, J.P., Kuruppu, J., Kunstman, K., Wolinsky, S., et al. 2002 HIV preferentially infects HIV-specific CD4+ T cells. *Nature* **417**, 95-98. (doi:10.1038/417095a).
- [138] Benlahrech, A., Harris, J., Meiser, A., Papagatsias, T., Hornig, J., Hayes, P., Lieber, A., Athanasopoulos, T., Bachy, V., Csomor, E., et al. 2009 Adenovirus vector vaccination induces expansion of memory CD4 T cells with a mucosal homing phenotype that are readily susceptible to HIV-1. *Proc Natl Acad Sci U S A* **106**, 19940-19945. (doi:10.1073/pnas.0907898106).
- [139] Papagno, L., Alter, G., Assoumou, L., Murphy, R.L., Garcia, F., Clotet, B., Larsen, M., Braibant, M., Marcelin, A.G., Costagliola, D., et al. 2011 Comprehensive analysis of virus-specific T-cells provides clues for the failure of therapeutic immunization with ALVAC-HIV vaccine. *Aids* **25**, 27-36. (doi:10.1097/QAD.0b013e328340fe55).
- [140] de Souza, M.S., Ratto- Kim, S., Chuenarom, W., Schuetz, A., Chantakulkij, S., Nuntapinit, B., Valencia-Micolta, A., Thelian, D., Nitayaphan, S., Pitisuttithum, P., et al. 2012 The Thai phase III trial (RV144) vaccine regimen induces T cell responses that preferentially target epitopes within the V2 region of HIV-1 envelope. *J Immunol* 188, 5166-5176. (doi:10.4049/jimmunol.1102756).
- [141] Frater, J., Ewings, F., Hurst, J., Brown, H., Robinson, N., Fidler, S., Babiker, A., Weber, J., Porter, K. & Phillips, R.E. 2014 HIV-1 specific CD4 responses in primary HIV-1 infection predict disease progression in the SPARTAC trial. *AIDS*. (doi:10.1097/QAD.000000000000130).
- [142] Zheng, N., Fujiwara, M., Ueno, T., Oka, S. & Takiguchi, M. 2009 Strong ability of Nefspecific CD4+ cytotoxic T cells to suppress human immunodeficiency virus type 1 (HIV-1) replication in HIV-1-infected CD4+ T cells and macrophages. *J Virol* **83**, 7668-7677. (doi:10.1128/JVI.00513-09).
- [143] Appay, V. 2004 The physiological role of cytotoxic CD4(+) T-cells: the holy grail? *Clin Exp Immunol* **138**, 10-13. (doi:10.1111/j.1365-2249.2004.02605.x CEI2605 [pii]).
- [144] Soghoian, D.Z., Jessen, H., Flanders, M., Sierra-Davidson, K., Cutler, S., Pertel, T., Ranasinghe, S., Lindqvist, M., Davis, I., Lane, K., et al. 2012 HIV-specific cytolytic CD4 T cell responses during acute HIV infection predict disease outcome. *Sci Transl Med* 4, 123ra125. (doi:10.1126/scitranslmed.3003165 4/123/123ra25 [pii]).
- [145] Oxenius, A., Price, D.A., Easterbrook, P.J., O'Callaghan, C.A., Kelleher, A.D., Whelan, J.A., Sontag, G., Sewell, A.K. & Phillips, R.E. 2000 Early highly active antiretroviral therapy for acute HIV-1 infection preserves immune function of CD8+ and CD4+ T lymphocytes. *Proc Natl Acad Sci U S A* **97**, 3382-3387.
- [146] Emu, B., Sinclair, E., Favre, D., Moretto, W.J., Hsue, P., Hoh, R., Martin, J.N., Nixon, D.F., McCune, J.M. & Deeks, S.G. 2005 Phenotypic, functional, and kinetic parameters associated with apparent T-cell control of human immunodeficiency virus replication in individuals with and without antiretroviral treatment. *J Virol* **79**, 14169-14178.
- [147] Potter, S.J., Lacabaratz, C., Lambotte, O., Perez-Patrigeon, S., Vingert, B., Sinet, M., Colle, J.H., Urrutia, A., Scott-Algara, D., Boufassa, F., et al. 2007 Preserved central memory and activated effector memory CD4+ T-cell subsets in human immunodeficiency virus controllers: an ANRS EP36 study. *J Virol* 81, 13904-13915.
- [148] Vingert, B., Benati, D., Lambotte, O., de Truchis, P., Slama, L., Jeannin, P., Galperin, M., Perez-Patrigeon, S., Boufassa, F., Kwok, W.W., et al. 2012 HIV controllers maintain a

population of highly efficient Th1 effector cells in contrast to patients treated in the long term. *J Virol* **86**, 10661-10674. (doi:10.1128/JVI.00056-12 JVI.00056-12 [pii]).

[149] Kaufmann, D.E., Kavanagh, D.G., Pereyra, F., Zaunders, J.J., Mackey, E.W., Miura, T., Palmer, S., Brockman, M., Rathod, A., Piechocka-Trocha, A., et al. 2007 Upregulation of CTLA-4 by HIV-specific CD4+ T cells correlates with disease progression and defines a reversible immune dysfunction. *Nat Immunol* **8**, 1246-1254.

[150] van Grevenynghe, J., Procopio, F.A., He, Z., Chomont, N., Riou, C., Zhang, Y., Gimmig, S., Boucher, G., Wilkinson, P., Shi, Y., et al. 2008 Transcription factor FOXO3a controls the persistence of memory CD4(+) T cells during HIV infection. *Nat Med* **14**, 266-274.

[151] Vingert, B., Perez-Patrigeon, S., Jeannin, P., Lambotte, O., Boufassa, F., Lemaitre, F., Kwok, W.W., Theodorou, I., Delfraissy, J.F., Theze, J., et al. 2010 HIV controller CD4+ T cells respond to minimal amounts of Gag antigen due to high TCR avidity. *PLoS Pathog* **6**, e1000780. (doi:10.1371/journal.ppat.1000780).

[152] Ferre, A.L., Hunt, P.W., McConnell, D.H., Morris, M.M., Garcia, J.C., Pollard, R.B., Yee, H.F., Jr., Martin, J.N., Deeks, S.G. & Shacklett, B.L. 2010 HIV controllers with HLA-DRB1*13 and HLA-DQB1*06 alleles have strong, polyfunctional mucosal CD4+ T-cell responses. *J Virol* **84**, 11020-11029.

[153] Perreau, M., Savoye, A.L., De Crignis, E., Corpataux, J.M., Cubas, R., Haddad, E.K., De Leval, L., Graziosi, C. & Pantaleo, G. 2013 Follicular helper T cells serve as the major CD4 T cell compartment for HIV- 1 infection, replication, and production. *J Exp Med* 210, 143-156. (doi:10.1084/jem.20121932 jem.20121932 [pii]).

[154] Xu, Y., Weatherall, C., Bailey, M., Alcantara, S., De Rose, R., Estaquier, J., Wilson, K., Suzuki, K., Corbeil, J., Cooper, D.A., et al. 2013 Simian immunodeficiency virus infects follicular helper CD4 T cells in lymphoid tissues during pathogenic infection of pigtail macaques. *J Virol* 87, 3760-3773. (doi:10.1128/JVI.02497-12).

[155] Brenchley, J.M., Vinton, C., Tabb, B., Hao, X.P., Connick, E., Paiardini, M., Lifson, J.D., Silvestri, G. & Estes, J.D. 2012 Differential infection patterns of CD4+ T cells and lymphoid tissue viral burden distinguish progressive and nonprogressive lentiviral infections. *Blood* **120**, 4172-4181. (doi:10.1182/blood-2012-06-437608) .

[156] Hong, J.J., Amancha, P.K., Rogers, K., Ansari, A.A. & Villinger, F. 2012 Spatial alterations between CD4(+) T follicular helper, B, and CD8(+) T cells during simian immunodeficiency virus infection: T/B cell homeostasis, activation, and potential mechanism for viral escape. *J Immunol* 188, 3247-3256. (doi:10.4049/jimmunol.1103138).

[157] Lindqvist, M., van Lunzen, J., Soghoian, D.Z., Kuhl, B.D., Ranasinghe, S., Kranias, G., Flanders, M.D., Cutler, S., Yudanin, N., Muller, M.I., et al. 2012 Expansion of HIV-specific T follicular helper cells in chronic HIV infection. *J Clin Invest* **122**, 3271-3280. (doi:10.1172/JCI64314).

[158] Petrovas, C., Yamamoto, T., Gerner, M.Y., Boswell, K.L., Wloka, K., Smith, E.C., Ambrozak, D.R., Sandler, N.G., Timmer, K.J., Sun, X., et al. 2012 CD4 T follicular helper cell dynamics during SIV infection. *J Clin Invest* **122**, 3281-3294. (doi:10.1172/JCl63039).

[159] Chevalier, M.F. & Weiss, L. 2012 The split personality of regulatory T cells in HIV infection. *Blood*. (doi:blood-2012-07-409755 [pii]

10.1182/blood-2012-07-409755).

[160] Epple, H.-J.r., Loddenkemper, C., Kunkel, D.e., Tröger, H., Maul, J., Moos, V., Berg, E., Ullrich, R., Schulzke, J.r.-D., Stein, H., et al. 2006 Mucosal but not peripheral FOXP3+ regulatory T cells are highly increased in untreated HIV infection and normalize after suppressive HAART. *Blood* **108**, 3072-3078. (doi:10.1182/blood-2006-04-016923).

[161] Dandekar, S., George, M.D. & Baumler, A.J. 2010 Th17 cells, HIV and the gut mucosal barrier. *Curr Opin HIV AIDS* **5**, 173-178. (doi:10.1097/COH.0b013e328335eda3).

[162] Angin, M., Kwon, D.S., Streeck, H., Wen, F., King, M., Rezai, A., Law, K., Hongo, T.C., Pyo, A., Piechocka-Trocha, A., et al. 2012 Preserved function of regulatory T cells in chronic

- HIV-1 infection despite decreased numbers in blood and tissue. *J Infect Dis* **205**, 1495-1500. (doi:jis236 [pii]
- 10.1093/infdis/jis236).
- [163] Hunt, P.W., Landay, A.L., Sinclair, E., Martinson, J.A., Hatano, H., Emu, B., Norris, P.J., Busch, M.P., Martin, J.N., Brooks, C., et al. 2011 A low T regulatory cell response may contribute to both viral control and generalized immune activation in HIV controllers. *PLoS ONE* **6**, e15924. (doi:10.1371/journal.pone.0015924).
- [164] Owen, R.E., Heitman, J.W., Hirschkorn, D.F., Lanteri, M.C., Biswas, H.H., Martin, J.N., Krone, M.R., Deeks, S.G. & Norris, P.J. 2010 HIV+ elite controllers have low HIV-specific T-cell activation yet maintain strong, polyfunctional T-cell responses. *Aids* **24**, 1095-1105. (doi:10.1097/QAD.0b013e3283377a1e).
- [165] Simonetta, F., Lecuroux, C., Girault, I., Goujard, C., Sinet, M., Lambotte, O., Venet, A. & Bourgeois, C. 2012 Early and long-lasting alteration of effector CD45RA(-)Foxp3(high) regulatory T-cell homeostasis during HIV infection. *J Infect Dis* **205**, 1510-1519. (doi:jis235 [pii]
- 10.1093/infdis/jis235).
- [166] Elahi, S., Dinges, W.L., Lejarcegui, N., Laing, K.J., Collier, A.C., Koelle, D.M., McElrath, M.J. & Horton, H. 2011 Protective HIV-specific CD8+ T cells evade Treg cell suppression. *Nat Med* 17, 989-995. (doi:10.1038/nm.2422 nm.2422 [pii]).
- [167] Overbaugh, J. & Morris, L. 2012 The Antibody Response against HIV-1. *Cold Spring Harb Perspect Med* **2**, a007039. (doi:10.1101/cshperspect.a007039).
- [168] Simek, M.D., Rida, W., Priddy, F.H., Pung, P., Carrow, E., Laufer, D.S., Lehrman, J.K., Boaz, M., Tarragona-Fiol, T., Miiro, G., et al. 2009 Human immunodeficiency virus type 1 elite neutralizers: individuals with broad and potent neutralizing activity identified by using a high-throughput neutralization assay together with an analytical selection algorithm. *J Virol* 83, 7337-7348. (doi:10.1128/JVI.00110-09).
- [169] Barouch, D.H., Whitney, J.B., Moldt, B., Klein, F., Oliveira, T.Y., Liu, J., Stephenson, K.E., Chang, H.W., Shekhar, K., Gupta, S., et al. 2013 Therapeutic efficacy of potent neutralizing HIV-1-specific monoclonal antibodies in SHIV-infected rhesus monkeys. *Nature* 503, 224-228. (doi:10.1038/nature12744 nature12744 [pii]).
- [170] Hirbod, T. & Broliden, K. 2007 Mucosal immune responses in the genital tract of HIV-1-exposed uninfected women. *J Intern Med* **262**, 44-58. (doi:10.1111/j.1365-2796.2007.01822.x).
- [171] Kaul, R., Trabattoni, D., Bwayo, J.J., Arienti, D., Zagliani, A., Mwangi, F.M., Kariuki, C., Ngugi, E.N., MacDonald, K.S., Ball, T.B., et al. 1999 HIV-1-specific mucosal IgA in a cohort of HIV-1-resistant Kenyan sex workers. *Aids* **13**, 23-29.
- [172] Ngo-Giang-Huong, N., Candotti, D., Goubar, A., Autran, B., Maynart, M., Sicard, D., Clauvel, J.P., Agut, H., Costagliola, D. & Rouzioux, C. 2001 HIV type 1-specific IgG2 antibodies: markers of helper T cell type 1 response and prognostic marker of long-term nonprogression. *AIDS Res Hum Retroviruses* 17, 1435-1446. (doi:10.1089/088922201753197105).
- [173] French, M.A., Center, R.J., Wilson, K.M., Fleyfel, I., Fernandez, S., Schorcht, A., Stratov, I., Kramski, M., Kent, S.J. & Kelleher, A.D. 2013 Isotype-switched immunoglobulin G antibodies to HIV Gag proteins may provide alternative or additional immune responses to 'protective' human leukocyte antigen-B alleles in HIV controllers. *Aids* **27**, 519-528. (doi:10.1097/QAD.0b013e32835cb720).
- [174] Lai, J.I., Licht, A.F., Dugast, A.S., Suscovich, T., Choi, I., Bailey-Kellogg, C., Alter, G. & Ackerman, M.E. 2013 Divergent antibody subclass and specificity profiles but not protective HLA-B alleles are associated with variable antibody effector function among HIV-1 controllers. *J Virol*. (doi:10.1128/JVI.03130-13).
- [175] Lambotte, O., Ferrari, G., Moog, C., Yates, N.L., Liao, H.X., Parks, R.J., Hicks, C.B., Owzar, K., Tomaras, G.D., Montefiori, D.C., et al. 2009 Heterogeneous neutralizing antibody

and antibody-dependent cell cytotoxicity responses in HIV-1 elite controllers. *Aids* **23**, 897-906. (doi:10.1097/QAD.0b013e328329f97d).

[176] Deepe, R.N., Kistner-Griffin, E., Martin, J.N., Deeks, S.G. & Pandey, J.P. 2012 Epistatic interactions between Fc (GM) and FcgammaR genes and the host control of human immunodeficiency virus replication. *Human immunology* **73**, 263-266. (doi:10.1016/j.humimm.2011.12.008).

[177] Liu, Q., Sun, Y., Rihn, S., Nolting, A., Tsoukas, P.N., Jost, S., Cohen, K., Walker, B. & Alter, G. 2009 Matrix metalloprotease inhibitors restore impaired NK cell-mediated antibody-dependent cellular cytotoxicity in human immunodeficiency virus type 1 infection. *Journal of virology* **83**, 8705-8712. (doi:10.1128/JVI.02666-08).

[178] Bonsignori, M., Pollara, J., Moody, M.A., Alpert, M.D., Chen, X., Hwang, K.K., Gilbert, P.B., Huang, Y., Gurley, T.C., Kozink, D.M., et al. 2012 Antibody-dependent cellular cytotoxicity-mediating antibodies from an HIV-1 vaccine efficacy trial target multiple epitopes and preferentially use the VH1 gene family. *J Virol* **86**, 11521-11532. (doi:10.1128/JVI.01023-12

[179] Rerks-Ngarm, S., Pitisuttithum, P., Nitayaphan, S., Kaewkungwal, J., Chiu, J., Paris, R., Premsri, N., Namwat, C., de Souza, M., Adams, E., et al. 2009 Vaccination with ALVAC and AIDSVAX to prevent HIV-1 infection in Thailand. *N Engl J Med* **361**, 2209-2220. (doi:10.1056/NEJMoa0908492).

[180] Allouch, A., David, A., Amie, S.M., Lahouassa, H., Chartier, L., Margottin-Goguet, F., Barre-Sinoussi, F., Kim, B., Saez-Cirion, A. & Pancino, G. 2013 p21-mediated RNR2 repression restricts HIV-1 replication in macrophages by inhibiting dNTP biosynthesis pathway. *Proc Natl Acad Sci U S A* **110**, E3997-4006. (doi:10.1073/pnas.1306719110 1306719110 [pii]).

[181] Stoiber, H., Banki, Z., Wilflingseder, D. & Dierich, M.P. 2008 Complement-HIV interactions during all steps of viral pathogenesis. *Vaccine* **26**, 3046-3054. (doi:10.1016/j.vaccine.2007.12.003

S0264-410X(07)01448-X [pii]).

JVI.01023-12 [pii]).

[182] Banki, Z., Posch, W., Ejaz, A., Oberhauser, V., Willey, S., Gassner, C., Stoiber, H., Dittmer, U., Dierich, M.P., Hasenkrug, K.J., et al. 2010 Complement as an endogenous adjuvant for dendritic cell-mediated induction of retrovirus -specific CTLs. *PLoS Pathog* **6**, e1000891. (doi:10.1371/journal.ppat.1000891).

[183] Posch, W., Cardinaud, S., Hamimi, C., Fletcher, A., Muhlbacher, A., Loacker, K., Eichberger, P., Dierich, M.P., Pancino, G., Lass-Florl, C., et al. 2012 Antibodies attenuate the capacity of dendritic cells to stimulate HIV-specific cytotoxic T lymphocytes. *J Allergy Clin Immunol* **130**, 1368-1374 e1362. (doi:10.1016/j.jaci.2012.08.025).

[184] Pandrea, I., Cornell, E., Wilson, C., Ribeiro, R.M., Ma, D., Kristoff, J., Xu, C., Haret-Richter, G.S., Trichel, A., Apetrei, C., et al. 2012 Coagulation biomarkers predict disease progression in SIV-infected nonhuman primates. *Blood* **120**, 1357-1366. (doi:10.1182/blood-2012-03-414706).

[185] Diop, O.M., Ploquin, M.J., Mortara, L., Faye, A., Jacquelin, B., Kunkel, D., Lebon, P., Butor, C., Hosmalin, A., Barre-Sinoussi, F., et al. 2008 Plasmacytoid dendritic cell dynamics and alpha interferon production during Simian immunodeficiency virus infection with a nonpathogenic outcome. *J Virol* 82, 5145-5152. (doi:10.1128/JVI.02433-07).

[186] Wijewardana, V., Kristoff, J., Xu, C., Ma, D., Haret-Richter, G., Stock, J.L., Policicchio, B.B., Mobley, A.D., Nusbaum, R., Aamer, H., et al. 2013 Kinetics of myeloid dendritic cell trafficking and activation: impact on progressive, nonprogressive and controlled SIV infections. *PLoS Pathog* **9**, e1003600. (doi:10.1371/journal.ppat.1003600).

[187] Pereira, L.E., Johnson, R.P. & Ansari, A.A. 2008 Sooty mangabeys and rhesus macaques exhibit significant divergent natural killer cell responses during both acute and chronic phases of SIV infection. *Cellular Immunology* **254**, 10-19. (doi:10.1016/j.cellimm.2008.06.006).

[188] Bosinger, S.E., Johnson, Z.P., Folkner, K.A., Patel, N., Hashempour, T., Jochems, S.P., del Rio Estrada, P.M., Paiardini, M., Lin, R., Vanderford, T.H., et al. 2013 Intact Type I

- Interferon Production and IRF7 Function in Sooty Mangabeys. *PLoS Pathog* **9**, e1003597. (doi:10.1371/journal.ppat.1003597).
- [189] Jacquelin, B., Mayau, V., Targat, B., Liovat, A.S., Kunkel, D., Petitjean, G., Dillies,
- M.A., Roques, P., Butor, C., Silvestri, G., et al. 2009 Nonpathogenic SIV infection of African green monkeys induces a strong but rapidly controlled type I IFN response. *J Clin Invest* **119**, 3544-3555. (doi:10.1172/JCl40093).
- [190] Jacquelin, B., Zahn, R.C., Barré-Sinoussi, F., Schmitz, J.E., Kaur, A. & Müller-Trutwin, M.C. 2011 Natural SIV Infection: Immunological Aspects. In *Models of Protection Against HIV/SIV: Avoiding AIDS in humans and monkeys* (eds. G. Pancino, G. Silvestri & K. Fowke), pp. 47-80, Academic Press is an imprint of Elsevier.
- [191] Descours, B., Avettand-Fenoel, V., Blanc, C., Samri, A., Melard, A., Supervie, V., Theodorou, I., Carcelain, G., Rouzioux, C. & Autran, B. 2012 Immune responses driven by protective human leukocyte antigen alleles from long-term nonprogressors are associated with low HIV reservoir in central memory CD4 T cells. *Clin Infect Dis* **54**, 1495-1503. (doi:10.1093/cid/cis188).
- [192] Choudhary, S.K., Vrisekoop, N., Jansen, C.A., Otto, S.A., Schuitemaker, H., Miedema, F. & Camerini, D. 2007 Low immune activation despite high levels of pathogenic human immunodeficiency virus type 1 results in long-term asymptomatic disease. *J Virol* **81**, 8838-8842.
- [193] Hunt, P.W., Brenchley, J., Sinclair, E., McCune, Joseph M., Roland, M., Page-Shafer, K., Hsue, P., Emu, B., Krone, M., Lampiris, H., et al. 2008 Relationship between T Cell Activation and CD4+ T Cell Count in HIV-Seropositive Individuals with Undetectable Plasma HIV RNA Levels in the Absence of Therapy. *J Infect Dis* **197**, 126-133. (doi:doi:10.1086/524143).
- [194] Hsue, P.Y., Hunt, P.W., Schnell, A., Kalapus, S.C., Hoh, R., Ganz, P., Martin, J.N. & Deeks, S.G. 2009 Role of viral replication, antiretroviral therapy, and immunodeficiency in HIV-associated atherosclerosis. *Aids* 23, 1059-1067. (doi:10.1097/QAD.0b013e32832b514b).
- [195] Pereyra, F., Lo, J., Triant, V.A., Wei, J., Buzon, M.J., Fitch, K.V., Hwang, J., Campbell, J.H., Burdo, T.H., Williams, K.C., et al. 2012 Increased coronary atherosclerosis and immune activation in HIV-1 elite controllers. *Aids* **26**, 2409-2412. (doi:10.1097/QAD.0b013e32835a9950).
- [196] Krishnan, S., Wilson, E.M., Sheikh, V., Rupert, A., Mendoza, D., Yang, J., Lempicki, R., Migueles, S.A. & Sereti, I. 2013 Evidence for Innate Immune System Activation in HIV Type 1-Infected Elite Controllers. *J Infect Dis.* (doi:10.1093/infdis/jit581).
- [197] Noel, N., Boufassa, F., Lecuroux, C., Saez-Cirion, A., Bourgeois, C., Dunyach-Remy, C., Goujard, C., Rouzioux, C., Meyer, L., Pancino, G., et al. 2013 Elevated IP10 levels are associated with immune activation and low CD4+ T-cell counts in HIV controller patients. *Aids*. (doi:10.1097/QAD.0000000000000174).
- [198] Hatano, H., Delwart, E.L., Norris, P.J., Lee, T.H., Dunn-Williams, J., Hunt, P.W., Hoh, R., Stramer, S.L., Linnen, J.M., McCune, J.M., et al. 2009 Evidence for persistent low-level viremia in individuals who control human immunodeficiency virus in the absence of antiretroviral therapy. *J Virol* 83, 329-335. (doi:10.1128/JVI.01763-08).
- [199] Boufassa, F., Saez-Cirion, A., Lechenadec, J.r., Zucman, D., Avettand-Fenoel, V.r., Venet, A., Rouzioux, C., Delfraissy, J.-F.o., Lambotte, O., Meyer, L., et al. 2011 CD4 Dynamics over a 15 Year-Period among HIV Controllers Enrolled in the ANRS French Observatory. *PLoS One* **6**, e18726.
- [200] Vigneault, F., Woods, M., Buzon, M.J., Li, C., Pereyra, F., Crosby, S.D., Rychert, J., Church, G., Martinez-Picado, J., Rosenberg, E.S., et al. 2011 Transcriptional profiling of CD4 T cells identifies distinct subgroups of HIV-1 elite controllers. *J Virol* **85**, 3015-3019. (doi:10.1128/JVI.01846-10).
- [201] Fukazawa, Y., Lum, R., Bae, J.Y., Okoye, A.A., Hagen, S.I., Park, H., Legasse, A.W., Axthelm, M.K., Estes, J.D., Piatak, M.J., et al. 2013 Effective CD8+ T cell responses restrict SIV replication to follivular helper T cells. In *31st annual symposium on nonhuman primate models for AIDS* (Atlanta, GA, USA).

- [202] Racz, P., Tenner-Racz, K., van Vloten, F., Schmidt, H., Dietrich, M., Gluckman, J.C., Letvin, N.L. & Janossy, G. 1990 Lymphatic tissue changes in AIDS and other retrovirus infections: tools and insights. *Lymphology* **23**, 85-91.
- [203] Pandrea, I., Gaufin, T., Brenchley, J.M., Gautam, R., Monjure, C., Gautam, A., Coleman, C., Lackner, A.A., Ribeiro, R.M., Douek, D.C., et al. 2008 Cutting edge: Experimentally induced immune activation in natural hosts of simian immunodeficiency virus induces significant increases in viral replication and CD4+ T cell depletion. *J Immunol* **181**, 6687-6691.
- [204] Passaes, C.P. & Saez-Cirion, A. 2014 HIV cure research: advances and prospects. *Virology IN PRESS*.
- [205] Persaud, D., Gay, H., Ziemniak, C., Chen, Y.H., Piatak, M., Jr., Chun, T.W., Strain, M., Richman, D. & Luzuriaga, K. 2013 Absence of detectable HIV-1 viremia after treatment cessation in an infant. *N Engl J Med* **369**, 1828-1835. (doi:10.1056/NEJMoa1302976).
- [206] Ngo-Giang-Huong, N., Deveau, C., Da Silva, I., Pellegrin, I., Venet, A., Harzic, M., Sinet, M., Delfraissy, J.F., Meyer, L., Goujard, C., et al. 2001 Proviral HIV-1 DNA in subjects
- followed since primary HIV-1 infection who suppress plasma viral load after one year of highly active antiretroviral therapy. *Aids* **15**, 665-673.
- [207] Delwart, E., Magierowska, M., Royz, M., Foley, B., Peddada, L., Smith, R., Heldebrant,
- C., Conrad, A. & Busch, M. 2002 Homogeneous quasispecies in 16 out of 17 individuals during very early HIV-1 primary infection. *Aids* **16**, 189-195.
- [208] Moir, S., Buckner, C.M., Ho, J., Wang, W., Chen, J., Waldner, A.J., Posada, J.G., Kardava, L., O'Shea, M.A., Kottilil, S., et al. 2010 B cells in early and chronic HIV infection: evidence for preservation of immune function associated with early initiation of antiretroviral therapy. *Blood* **116**, 5571-5579. (doi:10.1182/blood-2010-05-285528).
- [209] Leone, A., Picker, L.J. & Sodora, D.L. 2009 IL-2, IL-7 and IL-15 as immuno-modulators during SIV/HIV vaccination and treatment. *Curr HIV Res* **7**, 83-90.
- [210] Levy, Y., Lacabaratz, C., Weiss, L., Viard, J.P., Goujard, C., Lelievre, J.D., Boue, F., Molina, J.M., Rouzioux, C., Avettand-Fenoel, V., et al. 2009 Enhanced T cell recovery in HIV-1-infected adults through IL-7 treatment. *J Clin Invest* **119**, 997-1007. (doi:10.1172/JCl38052).
- [211] Levy, Y., Sereti, I., Tambussi, G., Routy, J.P., Lelievre, J.D., Delfraissy, J.F., Molina, J.M., Fischl, M., Goujard, C., Rodriguez, B., et al. 2012 Effects of recombinant human interleukin 7 on T-cell recovery and thymic output in HIV-infected patients receiving antiretroviral therapy: results of a phase I/IIa randomized, placebo-controlled, multicenter study. *Clin Infect Dis* **55**, 291-300. (doi:10.1093/cid/cis383).
- [212] Vandergeeten, C., Fromentin, R., DaFonseca, S., Lawani, M.B., Sereti, I., Lederman, M.M., Ramgopal, M., Routy, J.P., Sekaly, R.P. & Chomont, N. 2013 Interleukin-7 promotes HIV persistence during antiretroviral therapy. *Blood* **121**, 4321-4329. (doi:10.1182/blood-2012-11-465625).
- [213] Asmuth, D.M., Murphy, R.L., Rosenkranz, S.L., Lertora, J.J., Kottilil, S., Cramer, Y., Chan, E.S., Schooley, R.T., Rinaldo, C.R., Thielman, N., et al. 2010 Safety, tolerability, and mechanisms of antiretroviral activity of pegylated interferon Alfa-2a in HIV-1-monoinfected participants: a phase II clinical trial. *J Infect Dis* **201**, 1686-1696. (doi:10.1086/652420).
- [214] Azzoni, L., Foulkes, A.S., Papasavvas, E., Mexas, A.M., Lynn, K.M., Mounzer, K., Tebas, P., Jacobson, J.M., Frank, I., Busch, M.P., et al. 2013 Pegylated Interferon alfa-2a monotherapy results in suppression of HIV type 1 replication and decreased cell-associated HIV DNA integration. *J Infect Dis* **207**, 213-222. (doi:10.1093/infdis/jis663 iis663 [piil).
- [215] Boue, F., Reynes, J., Rouzioux, C., Emilie, D., Souala, F., Tubiana, R., Goujard, C., Lancar, R. & Costagliola, D. 2011 Alpha interferon administration during structured interruptions of combination antiretroviral therapy in patients with chronic HIV-1 infection: INTERVAC ANRS 105 trial. *Aids* **25**, 115-118. (doi:10.1097/QAD.0b013e328340a1e7).
- [216] Porichis, F. & Kaufmann, D.E. 2012 Role of PD-1 in HIV pathogenesis and as target for therapy. *Curr HIV/AIDS Rep* **9**, 81-90. (doi:10.1007/s11904-011-0106-4).

- [217] Velu, V., Titanji, K., Zhu, B., Husain, S., Pladevega, A., Lai, L., Vanderford, T.H., Chennareddi, L., Silvestri, G., Freeman, G.J., et al. 2009 Enhancing SIV-specific immunity in vivo by PD-1 blockade. *Nature* **458**, 206-210. (doi:10.1038/nature07662).
- [218] Velu, V., Mylvaganam, G., Freeman, G., Ahmed, R. & Amara, R. 2013 PD-1 blockade as an adjunct therapy for ART: Improves anti-SIV immunity, virologic response to ART and gut CD4 T cell restoration. In 31st annual symposium on nonhuman primate models for AIDS (Atlanta, GA, USA).
- [219] Chomont, N., El-Far, M., Ancuta, P., Trautmann, L., Procopio, F.A., Yassine-Diab, B., Boucher, G., Boulassel, M.R., Ghattas, G., Brenchley, J.M., et al. 2009 HIV reservoir size and persistence are driven by T cell survival and homeostatic proliferation. *Nat Med* **15**, 893-900. (doi:10.1038/nm.1972).
- [220] Dyavar Shetty, R., Velu, V., Titanji, K., Bosinger, S.E., Freeman, G.J., Silvestri, G. & Amara, R.R. 2012 PD-1 blockade during chronic SIV infection reduces hyperimmune activation and microbial translocation in rhesus macaques. *J Clin Invest* **122**, 1712-1716. (doi:10.1172/JCl60612).
- [221] Hatano, H. 2013 Immune activation and HIV persistence: considerations for novel therapeutic interventions. *Curr Opin HIV AIDS* **8**, 211-216. (doi:10.1097/COH.0b013e32835f9788).
- [222] Sperber, K., Chiang, G., Chen, H., Ross, W., Chusid, E., Gonchar, M., Chow, R. & Liriano, O. 1997 Comparison of hydroxychloroquine with zidovudine in asymptomatic patients infected with human immunodeficiency virus type 1. *Clin Ther* **19**, 913-923. (doi:S0149-2918(97)80045-8 [pii]) .
- [223] Sperber, K., Louie, M., Kraus, T., Proner, J., Sapira, E., Lin, S., Stecher, V. & Mayer, L. 1995 Hydroxychloroquine treatment of patients with human immunodeficiency virus type 1. *Clin Ther* **17**, 622-636. (doi:0149-2918(95)80039-5 [pii]).
- [224] Martinson, J.A., Montoya, C.J., Usuga, X., Ronquillo, R., Landay, A.L. & Desai, S.N. 2010 Chloroquine modulates HIV-1-induced plasmacytoid dendritic cell alpha interferon: implication for T-cell activation. *Antimicrobial agents and chemotherapy* **54**, 871-881. (doi:10.1128/AAC.01246-09).
- [225] Murray, S.M., Down, C.M., Boulware, D.R., Stauffer, W.M., Cavert, W.P., Schacker, T.W., Brenchley, J.M. & Douek, D.C. 2010 Reduction of immune activation with chloroquine therapy during chronic HIV infection. *J Virol* **84**, 12082-12086.
- [226] Paton, N.I., Goodall, R.L., Dunn, D.T., Franzen, S., Collaco-Moraes, Y., Gazzard, B.G., Williams, I.G., Fisher, M.J., Winston, A., Fox, J., et al. 2012 Effects of hydroxychloroquine on immune activation and disease progression among HIV-infected patients not receiving antiretroviral therapy: a randomized controlled trial. *JAMA* **308**, 353-361. (doi:10.1001/jama.2012.6936 1221697 [pii]).
- [227] Funderburg, N.T., Jiang, Y., Debanne, S.M., Storer, N., Labbato, D., Clagett, B., Robinson, J., Lederman, M.M. & McComsey, G.A. 2013 Rosuvastatin Treatment Reduces Markers of Monocyte Activation in HIV-Infected Subjects on Antiretroviral Therapy. *Clin Infect Dis.* (doi:cit748 [pii]
- 10.1093/cid/cit748).
- [228] Overton, E.T., Kitch, D., Benson, C.A., Hunt, P.W., Stein, J.H., Smurzynski, M., Ribaudo, H.J. & Tebas, P. 2013 Effect of statin therapy in reducing the risk of serious non-AIDS-defining events and nonaccidental death. *Clin Infect Dis* **56**, 1471-1479. (doi:10.1093/cid/cit053 [pii]).
- [229] Rasmussen, L.D., Kronborg, G., Larsen, C.S., Pedersen, C., Gerstoft, J. & Obel, N. 2013 Statin therapy and mortality in HIV-infected individuals; a Danish nationwide population-based cohort study. *PLoS One* **8**, e52828. (doi:10.1371/journal.pone.0052828 PONE-D-12-27304 [pii]).
- [230] Lichtenstein, K., Debes, R., Wood, K. *et al.*, 2013 Statin Use Is Associated with Incident Diabetes Mellitus among Patients in the HIV Outpatient Study. *In 20th CROI*. Abstract # 767, (Atlanta, GA, USA).

- [231] Takahashi, Y., Byrareddy, S., Albrecht, C., Brameier, M., Walter, L., Mayne, A.E., Dunbar, P., Russo, R., Little, D.M., Villinger, T., et al. 2014 In vivo Administration of a JAK3 Inhibitor During Acute SIV Infection Leads To Significant Increases in Viral Load During Chronic Infection. *PLoS Pathog.* (doi:in press).
- [232] Morlat, P., Pereira, E., Clayette, P., Derreudre-Bosquet, N., Ecobichon, J.L., Benveniste, O., Saves, M. & Leport, C. 2008 Early evolution of plasma soluble TNF-alpha p75 receptor as a marker of progression in treated HIV-infected patients. *AIDS Res Hum Retroviruses* **24**, 1383-1389. (doi:10.1089/aid.2007.0293).
- [233] Ananworanich, J., Schuetz, A., Vandergeeten, C., Sereti, I., de Souza, M., Rerknimitr, R., Dewar, R., Marovich, M., van Griensven, F., Sekaly, R., et al. 2012 Impact of multitargeted antiretroviral treatment on gut T cell depletion and HIV reservoir seeding during acute HIV infection. *PLoS ONE* **7**, e33948. (doi:10.1371/journal.pone.0033948).
- [234] Gay, C., Dibben, O., Anderson, J.A., Stacey, A., Mayo, A.J., Norris, P.J., Kuruc, J.D., Salazar-Gonzalez, J.F., Li, H., Keele, B.F., et al. 2011 Cross-Sectional Detection of Acute HIV Infection: Timing of Transmission, Inflammation and Antiretroviral Therapy. *PLoS One* **6**, e19617.
- [235] Brown, K.C., Patterson, K.B., Malone, S.A., Shaheen, N.J., Prince, H.M., Dumond, J.B., Spacek, M.B., Heidt, P.E., Cohen, M.S. & Kashuba, A.D. 2011 Single and multiple dose pharmacokinetics of maraviroc in saliva, semen, and rectal tissue of healthy HIV-negative men. *J Infect Dis* **203**, 1484- 1490. (doi:10.1093/infdis/jir059).
- [236] Puertas, M.C., Massanella, M., Llibre, J.M., Ballestero, M., Buzon, M.J., Ouchi, D., Esteve, A., Boix, J., Manzardo, C., Miro, J.M., et al. 2013 Intensification of a raltegravir-based regimen with maraviroc in early HIV-1 infection. *Aids*. (doi:10.1097/QAD.000000000000066).
- [237] Hunt, P.W., Lederman, M.M. & Deeks, S.G. 2013 Response: Maraviroc intensification and microbial translocation. *Blood* **122**, 2283-2284. (doi:10.1182/blood-2013-08-516930).
- [238] Psomas, C., Lavigne, J.P., Barbuat, C., Trabelsi, S., Ghosn, J., Lascoux-Combe, C., Flandre, P., Cuzin, L., Reynes, J., Autran, B., et al. 2013 Maraviroc-induced decrease in circulating bacterial products is not linked to an increase in immune activation in HIV-infected individuals. *Blood* **122**, 2282-2283. (doi:10.1182/blood-2013-06-507012).
- [239] Hatano, H., Yukl, S.A., Ferre, A.L., Graf, E.H., Somsouk, M., Sinclair, E., Abdel-Mohsen, M., Liegler, T., Harvill, K., Hoh, R., et al. 2013 Prospective antiretroviral treatment of asymptomatic, HIV-1 infected controllers. *PLoS Pathog* **9**, e1003691. (doi:10.1371/journal.ppat.1003691).
- [240] Garcia, F., Climent, N., Guardo, A.C., Gil, C., Leon, A., Autran, B., Lifson, J.D., Martinez-Picado, J., Dalmau, J., Clotet, B., et al. 2013 A dendritic cell-based vaccine elicits T cell responses associated with control of HIV-1 replication. *Sci Transl Med* **5**, 166ra162. (doi:10.1126/scitranslmed.3004682).
- [241] Trumpfheller, C., Longhi, M.P., Caskey, M., Idoyaga, J., Bozzacco, L., Keler, T., Schlesinger, S.J. & Steinman, R.M. 2012 Dendritic cell-targeted protein vaccines: a novel approach to induce T-cell immunity. *J Intern Med* **271**, 183-192. (doi:10.1111/j.1365-2796.2011.02496.x).
- [242] Zhou, J., Cheung, A.K., Tan, Z., Wang, H., Yu, W., Du, Y., Kang, Y., Lu, X., Liu, L., Yuen, K.Y., et al. 2013 PD1-based DNA vaccine amplifies HIV-1 GAG-specific CD8+ T cells in mice. *J Clin Invest* **123**, 2629-2642. (doi:10.1172/JCl64704 [pii]).
- [243] Hansen, S.G., Ford, J.C., Lewis, M.S., Ventura, A.B., Hughes, C.M., Coyne-Johnson, L., Whizin, N., Oswald, K., Shoemaker, R., Swanson, T., et al. 2011 Profound early control of highly pathogenic SIV by an effector memory T-cell vaccine. *Nature* **473**, 523-527. (doi:10.1038/nature10003).
- [244] Hansen, S.G., Sacha, J.B., Hughes, C.M., Ford, J.C., Burwitz, B.J., Scholz, I., Gilbride, R.M., Lewis, M.S., Gilliam, A.N., Ventura, A.B., et al. 2013 Cytomegalovirus vectors violate CD8+ T cell epitope recognition paradigms. *Science* **340**, 1237874. (doi:10.1126/science.1237874).

[245] Trkola, A., Kuster, H., Rusert, P., Joos, B., Fischer, M., Leemann, C., Manrique, A., Huber, M., Rehr, M., Oxenius, A., et al. 2005 Delay of HIV-1 rebound after cessation of antiretroviral therapy through passive transfer of human neutralizing antibodies. *Nat Med* 11, 615-622. (doi:10.1038/nm1244).

[246] Moore, P.L., Gray, E.S., Wibmer, C.K., Bhiman, J.N., Nonyane, M., Sheward, D.J., Hermanus, T., Bajimaya, S., Tumba, N.L., Abrahams, M.R., et al. 2012 Evolution of an HIV glycan-dependent broadly neutralizing antibody epitope through immune escape. *Nat Med* 18, 1688-1692. (doi:10.1038/nm.2985).

[247] Kwong, P.D. & Mascola, J.R. 2012 Human antibodies that neutralize HIV-1: identification, structures, and B cell ontogenies. *Immunity* **37**, 412-425. (doi:10.1016/j.immuni.2012.08.012

S1074-7613(12)00378-0 [pii]).

[248] Liao, H.X., Lynch, R., Zhou, T., Gao, F., Alam, S.M., Boyd, S.D., Fire, A.Z., Roskin, K.M., Schramm, C.A., Zhang, Z., et al. 2013 Co-evolution of a broadly neutralizing HIV-1 antibody and founder virus. *Nature* **496**, 469-476. (doi:10.1038/nature12053 pii]).

[249] Jardine, J., Julien, J.P., Menis, S., Ota, T., Kalyuzhniy, O., McGuire, A., Sok, D., Huang, P.S., MacPherson, S., Jones, M., et al. 2013 Rational HIV immunogen design to target specific germline B cell receptors. *Science* **340**, 711-716. (doi:10.1126/science.1234150

science.1234150 [pii]).

[250] McGuire, A.T., Hoot, S., Dreyer, A.M., Lippy, A., Stuart, A., Cohen, K.W., Jardine, J., Menis, S., Scheid, J.F., West, A.P., et al. 2013 Engineering HIV envelope protein to activate germline B cell receptors of broadly neutralizing anti-CD4 binding site antibodies. *J Exp Med* 210, 655-663. (doi:10.1084/jem.20122824 jem.20122824 [pii]).

[251] Barouch, D.H., Stephenson, K.E., Borducchi, E.N., Smith, K., Stanley, K., McNally, A.G., Liu, J., Abbink, P., Maxfield, L.F., Seaman, M.S., et al. 2013 Protective efficacy of a global HIV-1 mosaic vaccine against heterologous SHIV challenges in rhesus monkeys. *Cell* **155**, 531-539. (doi:10.1016/j.cell.2013.09.061 S0092-8674(13)01280-4 [pii]).

Figure captions

Figure 1: Priorities for HIV cure research based on recommendations from the International

AIDS Society (IAS).

Figure 2: Schematic of the hypothetical variation in viremia, CD4+ T cell counts and

inflammation in post-treatment controllers (PTC, orange) compared with HIV controllers (HIC,

green), non-pathogenic infections such as SIV infection of natural hosts (NPI, pink) and

viremic non progressors (blue). The orange box on the PTC curve indicates the period of

cART initiation, and the dashed line indicates hypothetical levels.

Short title for page headings: Natural immune control against HIV/AIDS

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Table I: Extreme profiles of natural protection against HIV/AIDS

	Protection against HIV infection	Protection against AIDS		
Type of natural protection	Highly HIV-exposed seronegative individuals	HIV controllers	Viremic non progressors	African nonhuman primates
Characteristics	No sign of infection despite repeated exposure to HIV	 <0.5% of the HIV+ population Undetectable viremia Stable CD4+ T cell counts 	Very rareLong-term asymptomaticHigh viremiaStable CD4+ T cell counts	 African green monkeys, sooty mangabeys, mandrills Asymptomatic High viremia Stable CD4+ T cell counts
Mechanisms potentially involved	 Strong innate responses Humoural responses in mucosa Low levels of CD4+ T cell activation Post-virus entry blockade Host genetic polymorphism 	 Genetic background (HLA-B27 and HLA-B57) Early control of viral replication Strong CD8+ T cell responses Enhanced ADCC activity Reduced susceptibility of CD4+ T cells to HIV infection 	Low levels of immune activation	 Early and efficient resolution of inflammation and T cell activation Less infection of TCM and TFH cells No microbial translocation
Proofs of concept or possible clinical applications	 CCR5Δ32 homozygous bone marrow transplant B chemokines to block CCR5 Restriction factors 	 HAART treatment during acute infection, post- treatment controllers Boosting of immune responses 	Design of well-targeted anti-inflammatory treatments	

BASIC RESEARCH



TRANSLATIONAL RESEARCH



CLINICAL RESEARCH



CLINICAL CARE

- How hosts control HIV replication in the absence of therapy?
- What are the mechanisms contribute that to the establishment and maintenance of latent infection. including the respective role of ongoing viral replication and/or homeostatic proliferation?
- What are the tissue and cellular reservoirs of HIV in individuals on long-term antiretroviral therapy?
- What are the origins of immune activation and inflammation in the presence of antiretroviral therapy and their consequences for HIV persistence?



Develop strategies to stop the inflammation which fuels new target cells and weakens the host defenses

to detect latently infected

cells

Develop and test strategies to enhance the capacity of the host immune response to control active viral replication



Develop and test therapeutic agents or immunological strategies to safely eliminate latent infection or control viral reservoirs in animal models and in individuals on antiretroviral therapy



