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Review

Plasmacytoid dendritic cells move down on the list of suspects: In search of the immune pathogenesis of chronic hepatitis C[☆]

Matthew L. Albert^{1,2,*}, Jérémie Decalf^{1,2}, Stanislas Pol^{3,4,5}

¹*The Laboratory of Dendritic Cell Biology, Department of Immunology, Institut Pasteur, Paris, France*

²*INSERM U818, Paris, France*

³*Liver Unit (L8), Hôpital Cochin, Paris, France*

⁴*Université Paris V, René Descartes, Paris, France*

⁵*U568, INSERM, Paris, France*

Chronic hepatitis C is a major public health problem. Despite numerous clinical studies in humans and experimental observations made in chimpanzees, hepatitis C pathogenesis remains poorly understood. Here, we review the clinical features of acute and chronic disease, and discuss the role of the immune system in the pathogenesis of disease. Many are aware of the dual role of T cells: responsibility for clearance of the virus during acute phase; and liver injury during chronic phase. Nonetheless, there is an emerging belief that failure to prime HCV-specific T cells is responsible for the failure to spontaneously clear the virus, and possibly, for the lack of response to pegylated-IFN α_{2a} /ribavirin therapy. We have focused on the latest suspects, plasmacytoid dendritic cells (pDCs), considered to be the professional type I IFNs producing cells. We review the somewhat contradictory data regarding the functional capacity of pDCs in chronic HCV patients and argue that, while lower in relative concentration as compared to healthy individuals, they are not defective in their ability to initiate an innate inflammatory response. Thus, instead of being the culprit, pDCs may in fact represent a novel therapeutic target in order to improve upon existing therapies for treating HCV patients.

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According to the World Health Organization, HCV infection represents a major health concern with an estimated 180 million people infected i.e. 3% of the world's population, with 3–4 million new cases each year. Some regions (e.g. Egypt, Bolivia, Mongolia, Cameroon) are particularly affected with disease prevalence >10% of the population. Once individuals are infected, HCV rapidly accesses the liver, the dominant site of replication [1]. There are two general courses of HCV infection (Fig. 1). Approximately 20–40% of infections are generally benign, self-limited infections that clear within 6 months [2]. In fact, this may be an underestimate as

there is evidence for viral transmission (based on detection of plasma viremia) with subsequent clearance in the absence of sero-conversion [3,4]. Still, it is believed that the majority of infected individuals (60–80%) develop chronic infections that result in accumulating levels of liver damage. HCV RNA typically becomes detectable in serum within 7–14 days following exposure, with viral levels peaking at 6–10 weeks and declining rapidly thereafter [5,6]. Of those who develop chronic hepatitis C, one-third will have a lifetime risk of developing liver cirrhosis and a significant number will develop hepatocellular carcinoma (HCC) (Fig. 1) [7]. HCV is responsible for more than half of all HCC cases and two-thirds of all liver transplants in the developed world. Chronically infected patients may undergo treatment with pegylated interferon and ribavirin, but treatment is difficult to tolerate and is effective in only 40–80% of patients, depending on the infecting genotype [8].

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* Corresponding author.

E-mail address: albertm@pasteur.fr (M.L. Albert).

Pathogenesis of chronic infection is not well-defined, but it is known that HCV replicates at an extremely high rate, ranging from 10^3 to 10^7 units per ml of serum with an estimated 10^{12} new viral particles produced each day [9]. While non-cytopathic, infection results in extensive liver damage; and based on histological observations, it is believed that liver injury is immune- rather than virus-mediated. The adaptive immune response, specifically T cell activation, while slow to start, is believed to be an important criterion for clearance during acute infection [10]. The breadth (diversity of the HCV-reactive T cell repertoire) and the magnitude (number of HCV-reactive T cells) are thought to be the determinants of clearance; though insufficient, T cell activation remains robust throughout the chronic phase. In turn, tissue damage within the liver results in the production of transforming growth factor β (TGF β), which is considered to be one of the most important molecules in inflammation-induced fibrosis [11]. Long-term liver cirrhosis appears when the normal parenchyma is replaced by non-functional scar tissue, ultimately leading to liver insufficiency. Thus, the T cell response to HCV is a double-edge sword as cytotoxic T cells are essential to the destruction of infected hepatocytes, but over time, they destroy the liver.

Regarding treatment of chronic HCV patients, recent advances include the introduction of pegylated interferons in combination with ribavirin, resulting in viral eradication in 54–66% of treatment-naïve patients [12,13]. A lack of response to anti-HCV treatment can generally be categorized as either a complete non-responder (where HCV RNA levels do not significantly decline by $>2\text{-log}_{10}$ throughout therapy), or virological relapsers (where HCV RNA becomes undetectable during treatment but is detected again after discontinuation of therapy). Non-responders, and to a lesser extent relapsers, are mainly males, over 40 years, either overweight or have fatty liver, extended fibrosis (F3) or cirrhosis and infected by genotypes 1 or 4. SVR, in contrast, is usually preceded by a rapid viral decline within the first weeks of the treatment, with viral loads undetectable 6 months after discontinuation of treatment. It is widely known that HCV genotype is perhaps the single most important factor in predicting a successful therapeutic outcome [12,14]. This is reflected in current treatment guidelines [15], which advocate a more aggressive approach to treatment in patients infected with ‘difficult to treat’ HCV genotype 1, where the probability of true non-response or virological relapse is somewhat greater than in patients infected with other HCV genotypes [12,16]. Importantly, the availability of an effective therapy has also provided an important clue concerning the characterization of immune-mediated clearance of HCV.

One additional point that bears mentioning is that at no point during disease pathogenesis are HCV patients

thought to be immunosuppressed. Chronic HCV patients do not present to the clinic with evidence of opportunistic infections, a hallmark of an immunocompromised state. There is no higher rate of viral or bacterial infections in HCV-infected patients. Nor do they show evidence of poor response to vaccination (e.g. HBV vaccination). Thus, in considering the immune pathogenesis of the disease we insist that the clinical features of HCV infection do not fit with models in which global defects in immune responses are invoked (e.g. defective dendritic cells).

1. Overview of the immune correlates of spontaneous viral clearance

Much is now known about the host response to HCV infection and both chimpanzee and human studies have been instructive in determining which anti-viral mechanisms of the immune system correlate with clearance. There is a rapid IFN response during acute infection, however this does not seem to be predictive of spontaneous clearance. Similarly, HCV-specific humoral responses are commonly seen in HCV-infected chimpanzees and humans, however HCV-specific antibodies are not capable of conferring protection [17]. For reasons still unknown, they are late to emerge during acute infection with the average timing of sero-conversion being >40 days post-infection; and in chronic disease, the quasispecies diversity (in part due to the RNA polymerase being highly error-prone) results in the ability of HCV to evade humoral immune surveillance. In fact, a recent retrospective study of patient H illustrated the process of antigenic escape through the evolution and immune selection of viral glycoprotein variants, effectively outpacing the ability to generate neutralizing antibodies [18].

There is recent interest in NK cells, stimulated in part by the genetic study demonstrating that genes encoding the inhibitory NK cell receptor KIR2DL3 and HLA-C1 confer a pre-disposed ability to resolve HCV infection [19]. Functional studies on humans with distinct KIR/HLA genotypes support the notion that NK cells having a greater capacity to produce IFN γ translates into more efficient viral clearance [20].

As discussed above, more is known concerning the role of T cells during the acute phase of HCV infection, where it plays an important role in viral clearance. The role of T cells in chronic disease is less clear, in part due to the diversity of the viral proteome, a direct result of the immense quasispecies swarm. As for B cells, HCV is quite efficient at escaping the T cell immune and several mechanisms have been described [6]. This includes the evolution of escape mutations, both in the presented peptide epitope and in the consensus motifs required for its processing and presentation [21]. Some studies have focused on the notion of ‘original antigen sin,’ a belief

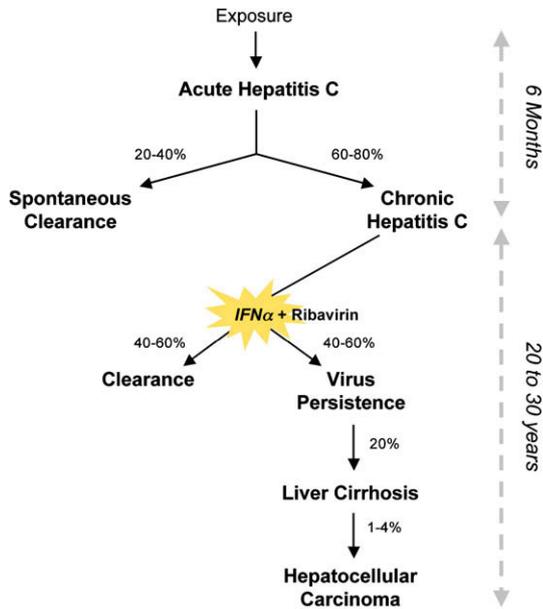


Fig. 1. HCV pathogenesis. Following exposure to the virus, acute HCV infection ensues and is asymptomatic in most of the cases. Infected individuals (60–80%) will not clear the virus and will develop a persistent infection that may last a lifetime, if untreated. Chronic infection triggers a chronic immune response and this persistent liver inflammatory liver is believed to be the cause of cirrhosis (20% of patients over the course of the disease) and can also result in the evolution of hepatocellular carcinoma (1–4% of individuals). Chronic HCV may be treated using a combination of Peg-IFN α_2 and Ribavirin, and depending on the viral genotype, response rates range from 40–80HCV.

that the existing pool of memory T cells present at the time of initial infection might influence the response to HCV [22]. This could favor viral clearance if the heterologous T cells are re-activated so as to generate a profound T cell activation during initial exposure to MHC complexes containing HCV peptide; but it may also be a disadvantage as cross-reactive low-affinity T cells could outcompete the generation of a more effective cohort of HCV-reactive T cells [23]. While this model is difficult to test, several lines of evidence would argue against its relevance to HCV immunity. First, prior exposure to influenza does not correlate with spontaneous resolution of HCV even through T cells reactive to HLA-A2.1/ influenza NA_{231–239} complexes cross-reacts with the immunodominant epitope HCV NS3_{1073–1081} [24]. Second, recent data suggest that naïve T cells outcompete memory cells, not the other way around [25]. Finally, more careful analysis indicates that there exist relatively high precursor frequencies of naïve HCV-reactive T cells within the repertoire of presumed, unexposed individuals [26]. Currently, the field is concerned with the expression of the inhibitory receptor programmed death-1 (PD-1), a member of the B7 - CD28 superfamily known to abrogate T cell activation [27]. While PD-1 expression on HCV specific CD8⁺ T cells was initially thought to be associated with cellular exhaustion [28], a recent study indicates that PD-1 expression level is not sufficient to predict infection outcome or to determine T cell func-

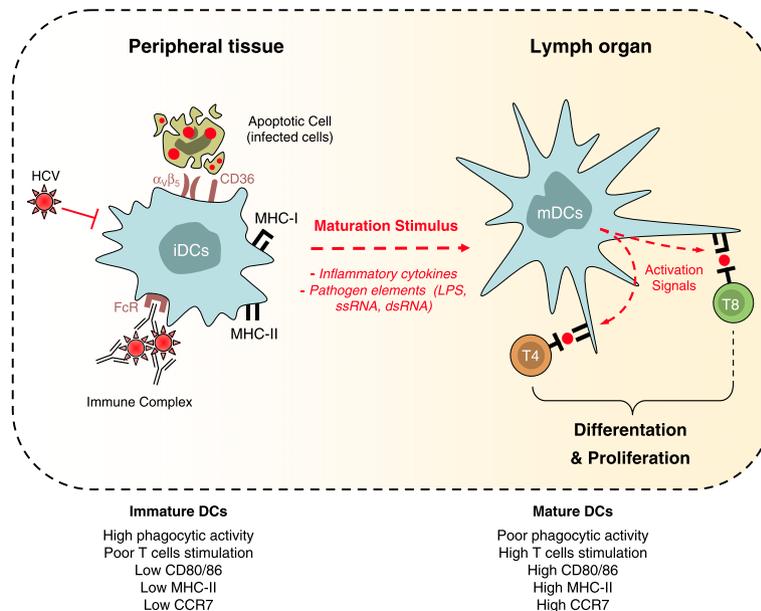


Fig. 2. Role for cDCs in inducing HCV-specific T cell immunity. In periphery, cDCs may capture HCV viral antigen by phagocytosis of infected apoptotic hepatocytes, endocytosis of immune complexes or by macropinocytosis of free virions. Antigenic elements are represented in red [Note. Based on their lack of Claudin-1 expression and the absence of miR-122 along with *in vitro* experimentation, we do not favor a role for direct infection.] During the maturation process, DCs may migrate to liver-draining lymph nodes and acquire a mature phenotype. They will also process antigen-derived peptides and present them on MHC-I and MHC-II molecules. MHC/peptides complexes associated with co-stimulatory molecules on DCs surface will trigger activation, differentiation and proliferation of antigen-specific CD4⁺ and CD8⁺ T cells. These activated T cells will go back to the site of infection to mediate the cellular immune response. In chronic HCV infection, these cells may be playing a role in chronic liver damage; but they are also critical for achieving viral clearance.

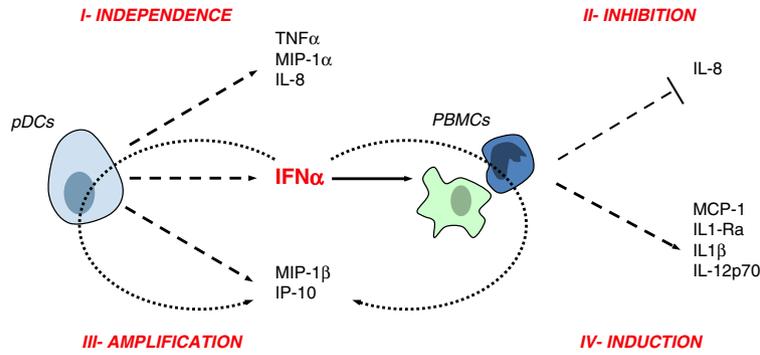


Fig. 3. Activation of pDCs induces four distinct chemokine/cytokine loops thus contributing to the initiation of an inflammatory response. Schematic representation of the four distinct cytokine loops that together help establish the pro-inflammatory response initiated by pDC activation. (I) In the first, activated pDCs secrete factors such as TNF α and MIP1 α in a manner that is triggered by TLR engagement and independent of IFNAR stimulation. (II) IL-8 is the only molecule we identified that follows a second pattern of expression – it is secreted by pDCs in response to TLR engagement its production by monocytes is inhibited by IFN α suggesting that TLR7 and TLR9 induced IL-8 production follows a different signaling pathway from TNF α -mediated IL-8 stimulation. (III) The third class of molecules is secreted by pDCs in response to TLR engagement with their expression being enhanced by autocrine IFN. Interestingly, in the case of MIP1 β and IP-10, pDC derived IFN may also induce other cell types to produce these chemokines in a manner that is apparently independent of direct TLR stimulation. (IV) In the fourth cytokine loop, illustrated by MCP-1 and also true for IL1Ra, IL1 β and IL-12p70, the pDCs do not produce but instead induce the production of these molecules by other cell types. These results suggest a coordinated set of events that support recruitment of defined cells and the production of inflammatory analytes for the initiation of an afferent immune response.

tionality in HCV infection [29]. Finally, an effort is being made to determine the relevance of elevated levels of

interferon-induced protein-10 (IP-10). Several studies have now reported that high plasma concentrations of

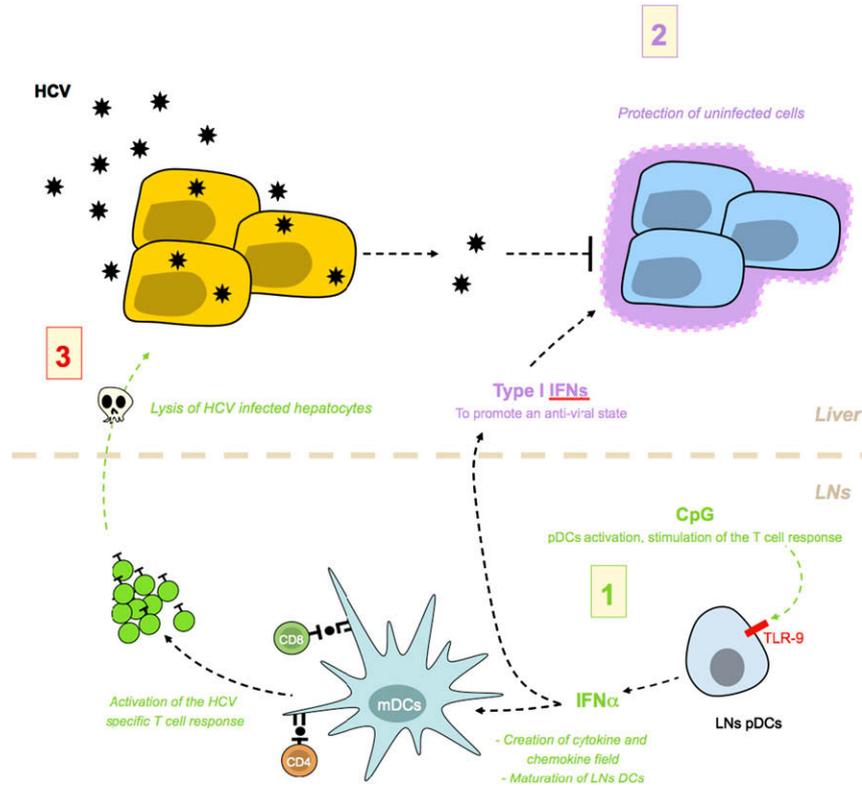


Fig. 4. Actions of pro-immune and anti-viral activity of pDC-derived endogenous IFNs. (1) CpG or other pDC agonists may be viable strategy for stimulating HCV specific T cell response in liver draining lymph nodes. Activated T cells will migrate to the liver and destroy HCV infected hepatocytes. (2) The endogenous IFN may also act to trigger anti-viral defense mechanisms within uninfected hepatocytes, inhibiting new infections and protecting from persistence of adaptive mutations. (3) Together, the dual action of pDC-derived IFN leads to better efficacy of anti-HCV therapy to achieve a definitive clearance of the virus. Advantages include the creation of a more robust cytokine/chemokine network and the avoidance of side effects that result from high doses of exogenous IFN.

IP-10 are a negative indicator in chronically infected patients, predicting failure to respond to therapy [30]. These data are somewhat paradoxical, as IP-10 is a pro-inflammatory chemokine, and is meant to attract T cells (as well as other leukocyte populations) to the liver. Current thought maintains that hormonal levels of IP-10 may in fact antagonize T cells from entering the liver, but additional work will be required to test this hypothesis.

Given the important role of T cells in HCV clearance, conventional dendritic cells (cDCs) have become the target of investigation, with the hypothesis that virus-mediated inhibition of cDC function might offer a possible explanation as to why some patients fail to mount an effective anti-viral immune response. Indeed, cDCs are critical for the priming of antigen-reactive T cells (Fig. 2) [31]. Initial studies focused on monocyte-derived cDCs, but quickly the field recognized the importance of studying cDCs *in situ*. Due to difficulties in gaining access to liver biopsies and the challenge of isolating cells from tissues, most have focused on circulating cells. Some studies have reported impaired allostimulatory function and defects in cDCs acquiring a mature or activated state [32–34]. In contrast, others have reported there to be no phenotypic or functional differences between cDCs isolated from chronically infected HCV patients *versus* those who have successfully cleared their virus [35]. Differences in isolation protocols, maturation cocktails or culturing conditions could all account for such differences, but the clinical evidence steers us away from such an explanation as there is no global impairment of the adaptive immune response in HCV infected individuals.

2. An important role for type I IFNs and a potential role for plasmacytoid dendritic cells

As mentioned, HCV typically reaches high serum titers within one week post-infection and studies of the innate immune response in chimpanzees have demonstrated that there is a profound host response. Transcriptional profiling of liver biopsies indicated induction if type I interferon (IFN) stimulated genes (ISGs) [36,37]. That said, it remains unclear what host sensors are responsible for the production of IFN α/β ; which cell types are producing these innate anti-viral effectors; and why type I IFN responses in the liver do not correlate with clearance even though the virus is highly sensitive to IFNs in *in vitro* experiments [38]. Some studies have demonstrated a role for RIG-I in sensing HCV RNA, pointing to the hepatocytes themselves as a possible source of the IFN β ; however recent data that the HCV protein NS3-4A is capable of cleaving Cardif would suggest that the signaling pathways of intracellular host sensors for viral RNA

are not active in infected cells [39]. Indeed, liver biopsies from HCV infected patients reveal aberrant localization of Cardif, consistent with it being inactive due its being cleaved from the mitochondrial membrane ([40] and personal communication, Jurg Tschopp). Furthermore, NS3-4A has been shown to inhibit IRF-3 phosphorylation, thus indicating that other host sensors (e.g. TLR3) are inactive in infected hepatocytes. In trying to account for the *in vivo* evidence of IFN α/β production during acute infection, one possibility is that HCV infection results in transient type I IFN production with a rapid shutdown after the virus has replicated and produced its non-structural proteins. These *in vitro* studies, however, do not fit with their being ISG expression during peak viremia [41,36] or with the data that shows endogenous IFNs being produced during the chronic phase of infection [42]. An alternative explanation, which integrates some of these findings, is that the type I IFNs are produced by non-infected hematopoietic cells.

Based on the recent advances in defining the source of type I IFN, many have proposed plasmacytoid dendritic cells (pDCs) as a prime suspect for producing IFN in chronic HCV patients; and we and others have been actively evaluating a role for these cells. pDCs are considered the natural IFN-producing cells, present in peripheral blood and capable of producing 100–1000 times more IFN α than other cell types when exposed to several viruses or bacteria [43,44]. Human pDCs are now well characterized and their ability to produce high amount of type I IFNs has earned them a place as a principal player in innate anti-viral immune responses. Human pDCs express TLR-7 and 9 [45], which recognize ssRNA and dsDNA, respectively, making them poised to respond to infectious pathogens. Whether pDCs also use cytosolic sensors to trigger type I IFN production has not been fully evaluated, but it seems as if they do not express RIG-I. Careful analysis has shown that upon activation, pDCs devote 60% of their global transcriptional activity to type I IFNs production [46] and the secretion of type I IFNs may be observed in less than 4 h after stimulation. In healthy individuals, pDCs are found in the blood and lymph organs and are believed to be absent from tissues in the stable state. Upon activation or during inflammatory responses, pDCs may migrate, both to the T cell area of lymph organs and also into the inflamed tissue parenchyma [47,48]. Regarding migration to lymph tissue, it is interesting to note that trafficking is directed to LNs that drain sites of inflammation [49] and that the path they travel differs from the one used by cDCs. In contrast to resting cDCs, which reside in tissues and migrate to the T cell area of local lymph nodes via the afferent lymphatics [31], pDCs migrate via the high endothelial venules (HEV) [50] involving L-selectin and CCR7

[49,51]. The mechanism of trafficking into the tissue remains less well defined, but may involve CXCR3, the receptor for IP-10 and two other related, interferon-induced chemokines, I-TAC and MIG. Once pDCs reach their destination, they are thought to stimulate aspects of both the innate and the adaptive immune response.

pDC derived-IFN may participate in a direct antiviral response, however increasing evidence suggests that it is through activation of other effector arms of the immune system that IFN α/β mediates HCV clearance. Indeed, both NK and CD8⁺ T cells are regulated by type I IFNs [52]. Type I IFNs have been shown to directly activate NK cells to enhance their cytotoxic activity [53] and also induce IL-15 production [54], which plays a critical role in proliferation and maintenance of NK cells. Concerning CTLs, a recent study has shown that CD8⁺ T cells lacking the IFN α/β receptor (IFNAR) are impaired in their ability to expand and differentiate into effector CTLs in the context of a viral infection [55]. Just as important may be the ability of pDCs to produce type I IFNs within lymphoid tissue, serving as an endogenous adjuvant for cDCs and provoking an enhanced production of IL-12p35 – the limiting subunit in IL-12p70, important for the differentiation of CD4⁺ T cells toward the Th1 effector lineage and the priming of CD8⁺ T cells. Whether IFNs also promote maturation of immature cDCs may turn out to be species-specific – this seems to be true in mouse models as shown in both *in vitro* and *in vivo* studies [56–58]; however human DCs do not behave in a similar manner. In addition to the production of IFNs, pDCs may directly interact with cDCs, engaging CD40 thus providing a distinct mechanism of cDC activation [59].

Our recent work has contributed in this area of study by providing a first-generation multi-analyte profile (MAP) of how pDCs serve to bridge innate and adaptive immune responses in the context of systemic viral or bacterial infections [60]. Taking advantage of high-quality data coming from medium-throughput proteomic tools such as Luminex xMAP technology, we have carried out an in-depth analysis of the cytokines and chemokines secreted when activated pDCs interact with other innate cells within the immune system (Fig. 3). Interestingly, we identified four distinct cytokine loops by which pDCs contribute to the initiation of an inflammatory response: (i) molecules secreted by the pDC itself and independent of IFN production; (ii) molecules secreted by the pDC and inhibited by paracrine IFN; (iii) molecules secreted by the pDC and amplified by paracrine IFN; and (iv) molecules not produced by pDCs but triggered by paracrine IFN. These cytokine/chemokine loops are shown here and have helped to provide a foundation for understanding the functional status of pDCs in different disease states.

3. Plasmacytoid DCs in HCV disease pathogenesis: Friend or Foe?

The observations concerning the role of type I IFN in facilitating cDCs to prime CD8⁺ T cells has led many to consider the possibility that HCV inhibits pDC function, thereby blocking endogenous IFN production. As it is difficult to reconcile this data with the fact that individuals chronically infected with HCV are not immunocompromised and that they have high levels of endogenous IFN [42], we evaluated the phenotypic measures and functional activity of patient pDCs as compared to healthy controls and HCV patients that had successfully cleared their virus (sustained virologic responders or SVR). We found no obvious defect in circulating pDCs. This conclusion was based on studies using both TLR7 and TLR9 agonists, assessing the ability of patient pDCs to: upregulate activation markers and homing receptors upon pDC activation; produce a broad array of cytokines and chemokines; and create an inflammatory network via their direct effects on other cell populations within the peripheral blood. While our results are in agreement with the study of Piccioli et al. [61], several groups have reported subversion of pDC in chronic HCV patients [34,62,63]. One important consideration is that we tested cytokine and chemokine production on a per-pDC basis [60,64]. This likely accounts for the differences reported by Szabo et al., who monitored IFN production within total PBMCs and did not account for the fact that there are 2–3 \times fewer pDCs in the patients with chronic HCV as compared to their normal control population [63]. Moreover, in some of the reported studies, pDCs were purified from PBMCs utilizing anti-BDCA-2 antibodies – importantly, BDCA-2 engagement is known to affect pDC function and might have confounded some of the findings [65]. It is less clear why Murakami et al. and Kanto et al. observed impaired pDC function though it may be a result of different culturing conditions used, as pDC survival *ex vivo* is quite poor in the absence of exogenous growth factors or adequate TLR stimulation.

In addition to immunologic measures of pDC function, there has been interest in defining whether there exists extra-hepatic sites of HCV replication. Indeed, demonstration that pDCs are infected by HCV might help support the notion of immune subversion. With the recent advances in the field concerning the generation of replication competent HCV and the ability to engineer reporter viruses, it has become easier to directly test this hypothesis. Using recombinant viruses engineered to express renilla luciferase and a highly sensitive measure of infection, our studies suggested that there is no direct infection of pDCs. Given the lack of an intrinsic defect in chronically infected HCV patient pDCs, and the ability of circulating cells to respond appropriately to TLR stimulation, this result is not very surpris-

ing. Moreover, it is clear that pDCs (as well as cDCs for that matter), lack the expression of claudin-1, one of the co-receptors for HCV entry into target cells [66,60].

4. Plasmacytoid DCs as a viable drug target for chronic HCV patients

These observations open up the possibility for pDCs to be harnessed for their therapeutic potential (Fig. 4). Many *in vivo* experiments using mouse models have explored how CpG could potentiate the cellular response to specific antigens [67–69], presumably acting via pDC stimulation. There are limitations in these systems, however, due to the fact that CpG will directly activate cDCs, in turn triggering maturation and T cell priming [69]. Moreover, there is evidence for direct activation of T cells in mice treated with CpG [70].

In contrast to mice, human cDCs do not express TLR-9. This makes it difficult to translate findings in experimental models to humans. Nonetheless, it is believed pDCs will be able to create an inflammatory milieu that will facilitate an enhance cellular immune response. Use of pDC agonists may in fact offer an alternative to the use of exogenous rIFN α_2 . It is worthy of mention that this approach may have three important advantages over conventional therapy: (i) it facilitates delivery of the IFN stimulation to the lymph node micro-environment. This is achieved by harnessing the biology of activated pDCs, which upregulate CCR7 and traffic to the site of T cell priming. In this way it offers a second potential benefit – (ii) by concentrating the IFN α production to the lymphoid organs, it may be possible to achieve similar activity with lower systemic levels of IFN α . Thus, the stimulation of the hypothalamic–pituitary–adrenal axis may be less significant, helping to avoid some of the more severe side effects of therapy such as mood disorders. (iii) This strategy also capitalizes on two waves of chemokine/cytokine production initiated by activated pDCs with the secretion of several known (e.g. TNF α and CCL3) and possibly many additional undefined analytes that are produced in a manner that is independent of IFN α/β [71,60].

One such clinical trial was carried out by Coley Pharmaceuticals to evaluate the efficiency of CpG as a treatment for chronic HCV. A phase 1b trial using CpG 10101 was conducted in chronically infected HCV patients and the results have been recently published [72]. They reported low levels of type I IFNs present in the plasma, but a strong IFN signature (based on IP-10 production and 2'5'-OAS expression in PBMCs). Most notably, CpG treatment was associated with a dose-dependent decrease in viral load. These results are interesting as low plasma concentration of type I IFN α yielded a clinical response, suggesting that indeed

it may be possible to segregate the pro-immune effects from the neuro-endocrine effects generally associated with rIFN α therapy. CpG treatment was also associated with a global activation of leucocytes. Activation of T cells, B cells, NK cells and pDCs were observed with a coincident reduction in the number circulating cells (also an indicator of activation) [73].

5. Plasmacytoid DCs remain on the list of suspects

While our studies concluded that pDCs isolated from patients with chronic HCV infection are phenotypically and functionally normal; and we are enthusiastic about pDC as a potential drug target, there remains some concern in using this approach and there may be some new data to argue that pDCs are not yet off the hook.

One consistent observation across most (if not all) of the published studies is that the relative percentage of circulating pDCs per total PBMCs is decreased in chronically infected HCV patients [64,60,33]. While this was also the case in patients who had achieved SVR [64,60] as well as individuals with non-viral liver disease [64], it begs the question as to whether the pDC numbers are decreased due to poor production, increased death, or differentiation and migration of the cells into sites of inflammation and/or lymphoid organs. There is no data regarding the first two proposals, but two reports have offered data concerning pDC trafficking, suggesting this is not the cause for lower numbers of circulating pDCs. Lai et al. evaluated liver DCs, comparing cDC and pDC populations from chronic HCV patients and individuals with non-viral liver disease. In contrast to the cDCs, which were more numerous and phenotypically and functionally mature, pDCs were present at lower frequency and expressed higher levels of BDCA-2 (a C-type lectin that negatively regulates IFN α/β production). Tang et al. studied the liver draining lymph nodes of chronic HCV patients, and compared to normal individuals, there was no marked increase in pDC number. One caveat that applies to both studies is that pDC differentiation/activation in settings of chronic inflammation remains poorly defined and surface expression of lineage markers may be altered in HCV infected patients.

There is also the concern that pDCs studied *ex vivo* do not recapitulate the effect of viral antigens on pDC function. While both reports that have utilized replication competent HCV to study pDC infection conclude that they are not susceptible, there remains the possibility that engagement of surface receptors by E1-E2 or soluble core might alter their function. pDCs express CD81 [74], which has been shown to engage HCV envelope [75,76]. In addition, circulating pDCs express C1qR (unpublished data), and while it has

not yet been formally demonstrated in pDCs, core binding to ClqR in several other cell types has been reported to be counter-inflammatory [77]. This may also have an impact on the use of pDCs as a drug target. In fact, one recent study suggested that HCV engagement of receptors on pDCs results in the inhibition of TLR9. While it remains to be dissected at a molecular level, and additional information is required to define how TLR9 but not TLR7 signaling is affected, this observation may impact the potential use of CpG in the treatment of HCV patients. For these types of studies we must also identify strategies to deal with the diversity of viral antigens within the quasispecies swarm as we already know, for example, that specific core variants can influence the immune system in unique ways [78].

6. Critical unknowns

Part of the confusion in defining the role of pDCs in HCV pathogenesis is that many questions remain about the mechanism by which type I IFNs mediate viral clearance. We will have to understand more about the endogenous IFNs produced during both the acute and chronic phases of infection: which cell(s) produce it; do type I IFNs mediate spontaneous clearance and if so why do they fail to limit HCV replication in chronic phase; and how are endogenous sources different from exogenous IFNs? We also believe that the working model for understanding the immune pathogenesis of HCV should be re-evaluated. HCV reactive T cells are present in chronically infected patients. In fact they are likely the effector cells responsible for the persistent liver damage. As such, pDCs may be involved in their activation and instead of looking for pDC dysfunction; perhaps they are hyper-activated as a result of a chronic infection and inflammation.

References

- [1] Nouri-Aria KT, Sallie R, Mizokami M, Portmann BC, Williams R. Intrahepatic expression of hepatitis C virus antigens in chronic liver disease. *J Pathol* 1995;175:77–83.
- [2] Seeff LB. Natural history of chronic hepatitis C. *Hepatology* 2002;36:S35–S46.
- [3] Al-Sherbiny M, Osman A, Mohamed N, Shata MT, Abdel-Aziz F, Abdel-Hamid M, et al. Exposure to hepatitis C virus induces cellular immune responses without detectable viremia or seroconversion. *Am J Trop Med Hyg* 2005;73:44–49.
- [4] Post JJ, Pan Y, Freeman AJ, Harvey CE, White PA, Palladinetti P, et al. Clearance of hepatitis C viremia associated with cellular immunity in the absence of seroconversion in the hepatitis C incidence and transmission in prisons study cohort. *J Infect Dis* 2004;189:1846–1855.
- [5] Orland JR, Wright TL, Cooper S. Acute hepatitis C. *Hepatology* 2001;33:321–327.
- [6] Bowen DG, Walker CM. Adaptive immune responses in acute and chronic hepatitis C virus infection. *Nature* 2005;436:946–952.
- [7] Lauer GM, Walker BD. Hepatitis C virus infection. *N Engl J Med* 2001;345:41–52.
- [8] Feld JJ, Hoofnagle JH. Mechanism of action of interferon and ribavirin in treatment of hepatitis C. *Nature* 2005;436:967–972.
- [9] Neumann AU, Lam NP, Dahari H, Gretch DR, Wiley TE, Layden TJ, et al. Hepatitis C viral dynamics in vivo and the antiviral efficacy of interferon-alpha therapy. *Science* 1998;282:103–107.
- [10] Freeman AJ, Pan Y, Harvey CE, Post JJ, Law MG, White PA, et al. The presence of an intrahepatic cytotoxic T lymphocyte response is associated with low viral load in patients with chronic hepatitis C virus infection. *J Hepatol* 2003;38:349–356.
- [11] Schuppan D, Krebs A, Bauer M, Hahn EG. Hepatitis C and liver fibrosis. *Cell Death Differ* 2003;10:S59–S67.
- [12] Fried MW, Shiffman ML, Reddy KR, Smith C, Marinos G, Goncalves Jr FL, et al. Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection. *N Engl J Med* 2002;347:975–982.
- [13] Zeuzem S, Diago M, Gane E, Reddy KR, Pockros P, Prati D, et al. Peginterferon alfa-2a (40 kilodaltons) and ribavirin in patients with chronic hepatitis C and normal aminotransferase levels. *Gastroenterology* 2004;127:1724–1732.
- [14] Manns MP, McHutchison JG, Gordon SC, Rustgi VK, Shiffman M, Reindollar R, et al. Peginterferon alfa-2b plus ribavirin compared with interferon alfa-2b plus ribavirin for initial treatment of chronic hepatitis C: a randomised trial. *Lancet* 2001;358:958–965.
- [15] Strader DB, Wright T, Thomas DL, Seeff LB. Diagnosis, management, and treatment of hepatitis C. *Hepatology* 2004;39:1147–1171.
- [16] Zeuzem S, Pawlotsky JM, Lukasiewicz E, von Wagner M, Gouli I, Lurie Y, et al. International, multicenter, randomized, controlled study comparing dynamically individualized versus standard treatment in patients with chronic hepatitis C. *J Hepatol* 2005;43:250–257.
- [17] Farci P, Alter HJ, Govindarajan S, Wong DC, Engle R, Lesniewski RR, et al. Lack of protective immunity against reinfection with hepatitis C virus. *Science* 1992;258:135–140.
- [18] von Hahn T, Yoon JC, Alter H, Rice CM, Rehermann B, Balfe P, et al. Hepatitis C virus continuously escapes from neutralizing antibody and T-cell responses during chronic infection in vivo. *Gastroenterology* 2007;132:667–678.
- [19] Khakoo SI, Thio CL, Martin MP, Brooks CR, Gao X, Astemborski J, et al. HLA and NK cell inhibitory receptor genes in resolving hepatitis C virus infection. *Science* 2004;305:872–874.
- [20] Ahlenstiel G, Martin MP, Gao X, Carrington M, Rehermann B. Distinct KIR/HLA compound genotypes affect the kinetics of human antiviral natural killer cell responses. *J Clin Invest* 2008;118:1017–1026.
- [21] Seifert U, Liermann H, Racanelli V, Halenius A, Wiese M, Wedemeyer H, et al. Hepatitis C virus mutation affects proteasomal epitope processing. *J Clin Invest* 2004;114:250–259.
- [22] Rehermann B, Nascimbeni M. Immunology of hepatitis B virus and hepatitis C virus infection. *Nat Rev Immunol* 2005;5:215–229.
- [23] Welsh RM, Selin LK. No one is naive: the significance of heterologous T-cell immunity. *Nat Rev Immunol* 2002;2:417–426.
- [24] Urbani S, Amadei B, Fiscicaro P, Pilli M, Missale G, Bertoletti A, et al. Heterologous T cell immunity in severe hepatitis C virus infection. *J Exp Med* 2005;201:675–680.
- [25] Helft J, Jacquet A, Joncker N, Grandjean I, Dorothee G, Kissenpennig A, et al. Antigen specific T–T interactions regulate CD4 T cell expansion. *Blood* 2008;112:1249–1258.
- [26] Neveu B, Debeaupuis E, Echasserieau K, Le Moullac-Vaidye B, Gassin M, Jegou L, et al. Selection of high avidity CD8 T cells

- correlates with control of hepatitis C virus infection. *Hepatology* 2008;48:713–722.
- [27] Sharpe AH, Freeman GJ. The B7-CD28 superfamily. *Nat Rev Immunol* 2002;2:116–126.
- [28] Urbani S, Amadei B, Tola D, Massari M, Schivazappa S, Missale G, et al. PD-1 expression in acute hepatitis C virus (HCV) infection is associated with HCV-specific CD8 exhaustion. *J Virol* 2006;80:11398–11403.
- [29] Kasprovicz V, Schulze Zur Wiesch J, Kuntzen T, Nolan BE, Longworth S, Berical A, et al. High level of PD-1 expression on hepatitis C virus (HCV)-specific CD8+ and CD4+ T cells during acute HCV infection, irrespective of clinical outcome. *J Virol* 2008;82:3154–3160.
- [30] Butera D, Marukian S, Iwamaye AE, Hembrador E, Chambers TJ, Di Bisceglie AM, et al. Plasma chemokine levels correlate with the outcome of antiviral therapy in patients with hepatitis C. *Blood* 2005;106:1175–1182.
- [31] Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature* 1998;392:245–252.
- [32] Auffermann-Gretzinger S, Keeffe EB, Levy S. Impaired dendritic cell maturation in patients with chronic, but not resolved, hepatitis C virus infection. *Blood* 2001;97:3171–3176.
- [33] Bain C, Fatmi A, Zoulim F, Zarski JP, Trepo C, Inchauspe G. Impaired allostimulatory function of dendritic cells in chronic hepatitis C infection. *Gastroenterology* 2001;120:512–524.
- [34] Kanto T, Inoue M, Miyatake H, Sato A, Sakakibara M, Yakushijin T, et al. Reduced numbers and impaired ability of myeloid and plasmacytoid dendritic cells to polarize T helper cells in chronic hepatitis C virus infection. *J Infect Dis* 2004;190:1919–1926.
- [35] Longman RS, Talal AH, Jacobson IM, Albert ML, Rice CM. Presence of functional dendritic cells in patients chronically infected with hepatitis C virus. *Blood* 2004;103:1026–1029.
- [36] Bigger CB, Brasky KM, Lanford RE. DNA microarray analysis of chimpanzee liver during acute resolving hepatitis C virus infection. *J Virol* 2001;75:7059–7066.
- [37] Su AI, Pezacki JP, Wodicka L, Brideau AD, Supkova L, Thimme R, et al. Genomic analysis of the host response to hepatitis C virus infection. *Proc Natl Acad Sci USA* 2002;99:15669–15674.
- [38] Lindenbach BD, Evans MJ, Syder AJ, Wolk B, Tellinghuisen TL, Liu CC, et al. Complete replication of hepatitis C virus in cell culture. *Science* 2005;309:623–626.
- [39] Meylan E, Curran J, Hofmann K, Moradpour D, Binder M, Bartenschlager R, et al. Cardif is an adaptor protein in the RIG-I antiviral pathway and is targeted by hepatitis C virus. *Nature* 2005;437:1167–1172.
- [40] Loo YM, Owen DM, Li K, Erickson AK, Johnson CL, Fish PM, et al. Viral and therapeutic control of IFN-beta promoter stimulator 1 during hepatitis C virus infection. *Proc Natl Acad Sci USA* 2006;103:6001–6006.
- [41] Thimme R, Bukh J, Spangenberg HC, Wieland S, Pemberton J, Steiger C, et al. Viral and immunological determinants of hepatitis C virus clearance, persistence, and disease. *Proc Natl Acad Sci USA* 2002;99:15661–15668.
- [42] Mihm S, Frese M, Meier V, Wietzke-Braun P, Scharf JG, Bartenschlager R, et al. Interferon type I gene expression in chronic hepatitis C. *Lab Invest* 2004;84:1148–1159.
- [43] Peter HH, Dallugge H, Zawatzky R, Euler S, Leibold W, Kirchner H. Human peripheral null lymphocytes. II. Producers of type-I interferon upon stimulation with tumor cells, Herpes simplex virus and *Corynebacterium parvum*. *Eur J Immunol* 1980;10:547–555.
- [44] Trinchieri G, Santoli D, Dee RR, Knowles BB. Anti-viral activity induced by culturing lymphocytes with tumor-derived or virus-transformed cells. Identification of the anti-viral activity as interferon and characterization of the human effector lymphocyte subpopulation. *J Exp Med* 1978;147:1299–1313.
- [45] Kadowaki N, Antonenko S, Lau JY, Liu YJ. Natural interferon alpha/beta-producing cells link innate and adaptive immunity. *J Exp Med* 2000;192:219–226.
- [46] Ito T, Kanzler H, Duramad O, Cao W, Liu YJ. Specialization, kinetics, and repertoire of type I interferon responses by human plasmacytoid dendritic cells. *Blood* 2006;107:2423–2431.
- [47] Cella M, Jarrossay D, Facchetti F, Alebardi O, Nakajima H, Lanzavecchia A, et al. Plasmacytoid monocytes migrate to inflamed lymph nodes and produce large amounts of type I interferon. *Nat Med* 1999;5:919–923.
- [48] Lai WK, Curbishley SM, Goddard S, Alabraba E, Shaw J, Youster J, et al. Hepatitis C is associated with perturbation of intrahepatic myeloid and plasmacytoid dendritic cell function. *J Hepatol* 2007;47:338–347.
- [49] Yoneyama H, Matsuno K, Zhang Y, Nishiwaki T, Kitabatake M, Ueha S, et al. Evidence for recruitment of plasmacytoid dendritic cell precursors to inflamed lymph nodes through high endothelial venules. *Int Immunol* 2004;16:915–928.
- [50] Matsutani T, Tanaka T, Tohya K, Otani K, Jang MH, Umemoto E, et al. Plasmacytoid dendritic cells employ multiple cell adhesion molecules sequentially to interact with high endothelial venule cells molecular basis of their trafficking to lymph nodes. *Int Immunol* 2007;19:1031–1037.
- [51] Penna G, Sozzani S, Adorini L. Cutting edge: selective usage of chemokine receptors by plasmacytoid dendritic cells. *J Immunol* 2001;167:1862–1866.
- [52] Stetson DB, Medzhitov R. Type I interferons in host defense. *Immunity* 2006;25:373–381.
- [53] Lee CK, Rao DT, Gertner R, Gimeno R, Frey AB, Levy DE. Distinct requirements for IFNs and STAT1 in NK cell function. *J Immunol* 2000;165:3571–3577.
- [54] Nguyen KB, Salazar-Mather TP, Dalod MY, Van Deusen JB, Wei XQ, Liew FY, et al. Coordinated and distinct roles for IFN-alpha beta, IL-12, and IL-15 regulation of NK cell responses to viral infection. *J Immunol* 2002;169:4279–4287.
- [55] Kolumam GA, Thomas S, Thompson LJ, Sprent J, Murali-Krishna K. Type I interferons act directly on CD8 T cells to allow clonal expansion and memory formation in response to viral infection. *J Exp Med* 2005;202:637–650.
- [56] Gallucci S, Lolkema M, Matzinger P. Natural adjuvants: endogenous activators of dendritic cells. *Nat Med* 1999;5:1249–1255.
- [57] Luft T, Pang KC, Thomas E, Hertzog P, Hart DN, Trapani J, et al. Type I IFNs enhance the terminal differentiation of dendritic cells. *J Immunol* 1998;161:1947–1953.
- [58] Montoya M, Schiavoni G, Mattei F, Gresser I, Belardelli F, Borrow P, et al. Type I interferons produced by dendritic cells promote their phenotypic and functional activation. *Blood* 2002;99:3263–3271.
- [59] Yoneyama H, Matsuno K, Toda E, Nishiwaki T, Matsuo N, Nakano A, et al. Plasmacytoid DCs help lymph node DCs to induce anti-HSV CTLs. *J Exp Med* 2005;202:425–435.
- [60] Decalf J, Fernandes S, Longman R, Ahloulay M, Audat F, Lefrerre F, et al. Plasmacytoid dendritic cells initiate a complex chemokine and cytokine network and are a viable drug target in chronic HCV patients. *J Exp Med* 2007;204:2423–2437.
- [61] Piccioli D, Tavarini S, Nuti S, Colombatto P, Brunetto M, Bonino F, et al. Comparable functions of plasmacytoid and monocyte-derived dendritic cells in chronic hepatitis C patients and healthy donors. *J Hepatol* 2005;42:61–67.
- [62] Murakami H, Akbar SM, Matsui H, Horiike N, Onji M. Decreased interferon-alpha production and impaired T helper 1 polarization by dendritic cells from patients with chronic hepatitis C. *Clin Exp Immunol* 2004;137:559–565.
- [63] Szabo G, Dolganiuc A. Subversion of plasmacytoid and myeloid dendritic cell functions in chronic HCV infection. *Immunobiology* 2005;210:237–247.

- [64] Longman RS, Talal AH, Jacobson IM, Rice CM, Albert ML. Normal functional capacity in circulating myeloid and plasmacytoid dendritic cells in patients with chronic hepatitis C. *J Infect Dis* 2005;192:497–503.
- [65] Dzionek A, Sohma Y, Nagafune J, Cella M, Colonna M, Facchetti F, et al. BDCA-2, a novel plasmacytoid dendritic cell-specific type II C-type lectin, mediates antigen capture and is a potent inhibitor of interferon alpha/beta induction. *J Exp Med* 2001;194:1823–1834.
- [66] Evans MJ, von Hahn T, Tscherne DM, Syder AJ, Panis M, Wolk B, et al. Claudin-1 is a hepatitis C virus co-receptor required for a late step in entry. *Nature* 2007;446:801–805.
- [67] Kraft AR, Krux F, Schimmer S, Ohlen C, Greenberg PD, Dittmer U. CpG oligodeoxynucleotides allow for effective adoptive T-cell therapy in chronic retroviral infection. *Blood* 2007;109:2982–2984.
- [68] Lonsdorf AS, Kuekrek H, Stern BV, Boehm BO, Lehmann PV, Tary-Lehmann M. Intratumor CpG-oligodeoxynucleotide injection induces protective antitumor T cell immunity. *J Immunol* 2003;171:3941–3946.
- [69] Vabulas RM, Pircher H, Lipford GB, Hacker H, Wagner H. CpG-DNA activates in vivo T cell epitope presenting dendritic cells to trigger protective antiviral cytotoxic T cell responses. *J Immunol* 2000;164:2372–2378.
- [70] Gelman AE, LaRosa DF, Zhang J, Walsh PT, Choi Y, Sunyer JO, et al. The adaptor molecule MyD88 activates PI-3 kinase signaling in CD4+ T cells and enables CpG oligodeoxynucleotide-mediated costimulation. *Immunity* 2006;25:783–793.
- [71] Piqueras B, Connolly J, Freitas H, Palucka AK, Banchereau J. Upon viral exposure, myeloid and plasmacytoid dendritic cells produce 3 waves of distinct chemokines to recruit immune effectors. *Blood* 2006;107:2613–2618.
- [72] McHutchison JG, Bacon BR, Gordon SC, Lawitz E, Shiffman M, Afdhal NH, et al. Phase 1B, randomized, double-blind, dose-escalation trial of CPG 10101 in patients with chronic hepatitis C virus. *Hepatology* 2007;46:1341–1349.
- [73] McHutchison JG. Immunophenotyping profile of CpG 10101 (ACTILON), a new TLR9 agonist antiviral for hepatitis C. *Hepatology* 2005;42:539A.
- [74] Yamada E, Montoya M, Schuettler CG, Hickling TP, Tarr AW, Vitelli A, et al. Analysis of the binding of hepatitis C virus genotype 1a and 1b E2 glycoproteins to peripheral blood mononuclear cell subsets. *J Gen Virol* 2005;86:2507–2512.
- [75] Pileri P, Uematsu Y, Campagnoli S, Galli G, Falugi F, Petracca R, et al. Binding of hepatitis C virus to CD81. *Science* 1998;282:938–941.
- [76] Scarselli E, Ansuini H, Cerino R, Roccasecca RM, Acali S, Filocamo G, et al. The human scavenger receptor class B type I is a novel candidate receptor for the hepatitis C virus. *EMBO J* 2002;21:5017–5025.
- [77] Yao ZQ, Ray S, Eisen-Vandervelde A, Waggoner S, Hahn YS. Hepatitis C virus: immunosuppression by complement regulatory pathway. *Viral Immunol* 2001;14:277–295.
- [78] Shiina M, Rehermann B. Cell culture-produced hepatitis C virus impairs plasmacytoid dendritic cell function. *Hepatology* 2007;47:385–395.