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The salivary glands and saliva of Anopheles gambiae as an essential step in the Plasmodium life cycle: a global proteomics study

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## SUMMARY

Proteins synthesized in the salivary glands of the Anopheles gambiae mosquito are thought to be important in the life cycle of the malaria parasite Plasmodium. To describe Anopheles gambiae salivary gland and saliva contents, we combined several techniques: 1-DE, 2-DE and LC MS/MS. This study has identified five saliva proteins and 122 more proteins from the salivary glands, including the first proteomic description for 89 of these salivary gland proteins. Since the invasion and sporozoite maturation take place during the process of salivary glands ageing, the effect of salivary gland age on salivary component composition was examined. LC MS/MS profiling of young versus old salivary gland proteomes suggests that there is an overrepresentation of proteins involved in signalling and proteins related to the immune response in the proteins from older mosquitoes. iTRAQ labelling was used for a comparative proteomic analysis of salivary gland samples from infected or Plasmodium berghei-free mosquitoes. The expression levels of five secreted proteins were altered when the parasite was present. These observations will serve as a basis for future work concerning the possible role of these proteins in the interaction between A. gambiae, Plasmodium and the mammalian host.

## INTRODUCTION

Malaria is a parasitic disease that affects 200 million people worldwide and causes 1.5 to 2.7 million deaths per year. Of the 300-500 million clinical cases annually, nearly $90 \%$ are in the sub-Saharan countries of Africa where the malaria parasite, Plasmodium falciparum, is primarily transmitted by the mosquito Anopheles gambiae. The increasing resistance of the parasite to inexpensive drugs and the resistance of mosquitoes to insecticides have created an urgent need for innovative methods that block parasite transmission during its development within the insect. The Anopheles mosquito not only carries the parasite from infected to uninfected people, but also plays a vital role in the parasite life cycle [1]. Mosquito saliva and salivary glands are central to the interaction between parasite, vector and mammalian host. Sporozoite maturation in the mosquito salivary glands before its transmission to vertebrates is a key stage for the effective transmission to humans since it increases the sporozoite's ability to infect vertebrate hepatocytes [2]. Additionally, sporozoites are injected into the vertebrate skin with nanolitre volumes of saliva, a complex biologically active solution, which, in addition to other activities, serves as the "transmission fluid" for the malaria parasite.

The salivary glands and their diversified protein contents are essential for overcoming the challenges posed by the host: pain and itch responses, immune defences and haemostasis [3]. There is convincing evidence that the pharmacological activity of arthropod saliva affects pathogen transmission. Salivary gland lysate from the sand fly Lutzomia longipalpis facilitates the infection of mice by the protozoan parasite Leishmania major [4, 5]. However, there has been little work on the role of mosquito salivary gland proteins in promoting infection of Plasmodium species in vertebrate hosts. During the last 3 years, there have been several studies on the
transcriptome and the proteome of salivary glands of arthropod vector saliva [6-11]. Kalume et al. [12] identified 67 proteins from Anopheles gambiae salivary glands, an initial step towards the cataloging of the hundreds of proteins and peptides in the salivary proteome. However, no attempts have been made to study the proteome of Anopheles gambiae saliva in the presence of malaria parasite.

This communication presents an expanded investigation of saliva and salivary proteins in bloodfed $A$. gambiae mosquitoes determined by several proteomics approaches. These techniques ensured good coverage of salivary gland proteins of varied pIs and molecular weights. The iTRAQ labelling technology was used to quantitate differences in the proteomes of Plasmodium berghei-infected and non-infected $A$. gambiae salivary glands.

## MATERIALS AND METHODS

## Reagents

## Mosquitoes

Yaounde strain adult $A$. gambiae females were reared in insect rooms at $26 \pm 0.5^{\circ} \mathrm{C}, 70 \%$ relative humidity, with a $16 \mathrm{~h} / 8 \mathrm{~h}$ light : dark photoperiod. The adult female mosquitoes used in these experiments were either aged between 5 and 8 days or between 18 and 21 days and had blood meals 3 to 5 days after emergence. Plasmodium berghei NK65 strain parasites, transformed to express GFP at the sporozoite stage, were injected into mice by intraperitoneal injection; seven days later, female mosquitoes aged 2-3 days were fed on the infected mice. All mosquitoes were maintained on a diet of $10 \%$ Karo syrup solution. Salivary glands from either 5-8 day old or at 18-21 day old mosquitoes were dissected in 150 mM NaCl with protease inhibitors (Complete, Roche Diagnostics, Manheim, Germany ) at $4^{\circ} \mathrm{C}$ and stored at $-80^{\circ} \mathrm{C}$. Saliva was collected using artificial feeders. After lyophilisation, saliva components were re-suspended in water and stored at $-80^{\circ} \mathrm{C}$.

## Salivary gland extract preparation

Salivary glands were disrupted by ultrasound (Cup horn, Sonics \& Materials Inc., Newton, CT, USA) for 20 min at maximum amplitude. Salivary gland homogenates were then centrifuged for 30 min at $130,000 \mathrm{~g}$ and protein was quantified using the $\mathrm{BCA}^{\mathrm{TM}}$ protein assay (Pierce, Rockville, IL, USA). Aliquots of salivary gland extracts were stored at $-80^{\circ} \mathrm{C}$ until use.

## SDS PAGE

SG samples of $10 \mu \mathrm{~g}$ or $36 \mu \mathrm{~g}$ of salivary gland were dissolved in Laemmli sample buffer, and boiled for 5 min . After centrifugation (14000rpm, 10 min ), $20 \mu \mathrm{l}$ samples were loaded onto a $12 \%$ acrylamide, 1 mm -thick SDS PAGE Bis-Tris minigel, and subjected to electrophoresis on a Novex apparatus (Invitrogen, Carlsbad, CA, USA). Protein molecular weight markers (Precision Plus Protein standard all blue, Bio-Rad, Hercules, CA, USA) were run on the same gel. The gel was stained with Bio-Safe ${ }^{\mathrm{TM}}$ Coomassie (Bio-Rad) or silver nitrate (PlusOne ${ }^{\mathrm{TM}}$, GE Healthcare, Uppsala, Sueden). Two methods were used to isolate proteins from the gel for mass spectrometry. One method consisted of cutting out all bands visible after Coomassie or silver staining. The other method consisted of cutting the gel into 1 mm -thick slices. The plugs obtained were analyzed by mass spectrometry.

## 2-DE

Samples of salivary gland supernatant, corresponding to 50 or $120 \mu \mathrm{~g}$ of protein, were used for 2D gel analysis. To improve 2-D gel profiles, samples were treated using a ReadyPrep 2-D Cleanup kit (Bio-Rad, Hercules, CA, USA). The pellet recovered after the last centrifugation step was dissolved in $15 \mathrm{mM} \mathrm{NaCl}, 0.5 \%$ SDS (final concentration), and 2\% Triton X100 (final concentration). The sample was heated at $95^{\circ} \mathrm{C}$ for 3 min , flash-frozen in liquid nitrogen and lyophilized. The lyophilized material was dissolved in 2-DE sample buffer (7M urea, 2 M thiourea, 4\% CHAPS, 150 mM DTT, and 2\% ampholytes).

SG samples ( $30 \mu \mathrm{l}$ ) were loaded onto IEF 18 cm gels containing ampholines of pH ranging from 4 to 8 (Bio-Rad), and run for 20000 Vhrs. The second dimension was carried out on $12.5 \%$ acrylamide 22 cm slab gels. Resolved proteins were detected by SYPRO®Ruby (Invitrogen). For
each type of salivary gland extract (young blood-fed, 21 day blood-fed, infected), at least three independent sample preparations were used, and at least three independent gel analyses were carried out.

## Mass spectrometry

## MALDI-TOF-MS and database searches

Mass spectrometry was performed using a MALDI-TOF instrument (Voyager-DE-STR, Applied Biosystems, Framingham, MA) operated in positive ion reflector mode. Sample preparation for in-gel digestion was carried out as described previously [13]. Bands and spots of interest were cut out using the Investigator ProPic robot (Genomic Solutions). Plugs were washed with 100 mM ammonium bicarbonate (Sigma) and proteins reduced with 10 mM 1,4-dithiothreitol (Sigma, Saint-Louis, MO, USA), $S$-alkylated with 55 mM iodoacetamide (Sigma) and in-gel digested at $37^{\circ} \mathrm{C}$ for 4 hours with modified porcine trypsin (Promega) using the Investigator ProGest robot (Genomic Solutions, Ann Arbor, MI, USA). Peptide mixtures were desalted on ZipTip C18 (Millipore) and directly eluted onto the Maldi target using the Investigator ProMS robot (Genomic Solutions). The elution solvent consisted of a six-fold dilution of a saturated solution of CHCA ( $10 \mathrm{mg} / \mathrm{ml}$, Sigma) in 70\% ACN (J.T. Baker) containing 0.1\% TFA (Sigma). Each mass spectrum ( $700-3000 \mathrm{~m} / \mathrm{z}$ ) was acquired in automatic mode ( 12 sub-spectra of 50 laser shots were accumulated). Trypsin autolysis peptides were used as internal calibratants (fragment 108-115: $[\mathrm{M}+\mathrm{H}]^{+}=842.5100$ and fragment $\left.58-77:[\mathrm{M}+\mathrm{H}]^{+}=2211.1046\right)$. A local copy of MS-FIT 3.2 software, part of the Protein Prospector package (University of California, San Francisco) was used to search the NCBI or Anopheles Ensembl databases. Search parameters were set as follows:
only monoisotopic masses were used, a maximum peptide mass error of 50 ppm was allowed and one incomplete cleavage per peptide and a possible oxidation of methionine were considered. Moreover, no restrictions on $M_{\mathrm{r}}$ or pI were made, and a minimum of four matching peptides covering a minimum of $15 \%$ of the protein sequence, were required for protein identification. If necessary, MALDI-TOF-PSD experiments were carried out to reach protein identifications using MS-TAG software (part of Protein Prospector package).

## LC MS/MS

## Protein digestion before identification by LC MSMS

Proteins were reduced, alkylated with 10 mM iodoacetamide, and digested with porcine trypsin (ratio 1:100) overnight at $37^{\circ} \mathrm{C}$. The trypsin digests were desalted with $\mathrm{C}_{18}$ tips (OMIX, Varian), and stored at $-80^{\circ} \mathrm{C}$ before LC MS/MS analysis.

## LC MS/MS analysis

Prior to reverse phase nanobore liquid chromatography tandem mass spectrometry (nanoLC MS/MS) analysis, samples were dissolved in Solvent A containing 5\% acetonitrile and $0.1 \%$ formic acid. The nanobore LC system was from LC Packings (Amsterdam, The Netherlands), and consisted of a Famos autosampler and an Ultimate Nano LC system. It was interfaced with a QqTOF mass spectrometer, QSTAR XL (AB/MDS Sciex, Foster City, CA), using a nanoelectrospray source (Protana Engineering A/S, Odense, Denmark). Reverse phase LC was performed using a PepMap column ( $75-\mu$ m inner diameter x $150-\mathrm{mm}$ long, LC Packings, Dionex) equilibrated with Solvant A. The peptides were eluted using a linear gradient of $5 \%$ to $40 \%$ solvent $\mathrm{B}\left(95 \%\right.$ acetonitrile, $5 \% \mathrm{H}_{2} \mathrm{O}$, and $0.1 \%$ formic acid $)$ in 90 min with a flow rate of 200 $\mathrm{nl} / \mathrm{min}$. This binary gradient was used for protein identification and iTRAQ experiments. We
operated the QSTAR XL mass spectrometer in an information-dependent-acquisition (IDA) mode; each full MS scan was followed by two MS/MS scans where the two most abundant peptide molecular ions were dynamically selected for CID, and dynamic exclusion was used to prevent repetitive selection of the same ions within a preset time. Collision energies were set to automatically adjust according to the charge state of the precursor ions.

## iTRAQ Sample Preparation Procedure.

We denatured $40 \mu \mathrm{~g}$ of each sample protein and blocked the cysteines as described in the iTRAQ protocol (Applied Biosystems, Foster City, CA). Each sample was then digested with trypsin solution overnight at $37^{\circ} \mathrm{C}$, and labelled with the iTRAQ tags as follows: non infected salivary glands, iTRAQ114; infected salivary glands by P. falciparum, iTRAQ116 or iTRAQ 117. The labelled samples were pooled and acidified for strong cation exchange (SCX) chromatography. The eluted peptides were then lyophilised and stored at $-81^{\circ} \mathrm{C}$ before analysis.

## Database search and relative quantification

MS/MS data were analyzed using ProID protein identification software version 1.1 (AB/MDS Sciex, Foster City, CA) using A. gambiae ORF database (Ensembl) [14]. In ProID, the peptide tolerance and the MS/MS tolerance were set to 0.15 Da . We manually inspected the $\mathrm{MS} / \mathrm{MS}$ spectra to validate the identified peptides.

ProQUANT 1.1 (AB/MDS Sciex, Foster City, CA) and the A. gambiae ORF database (Ensembl) were used to analyze data from the iTRAQ experiments. The confidence cut off was 95 . The tolerances set for peptide identification in ProQUANT searches were 0.15 Da for MS and 0.1 Da for MS/MS. We manually validated all identifications. Relative protein quantification in iTRAQ experiments was performed on the MS/MS scans and was the ratio of the areas under the peaks of
iTRAQ reagent tags at 114,116 , and 117 Da . The quantification results were normalized using the overall ratio obtained for all tagged peptide pairs in the sample.

## RESULTS AND DISCUSSION

# Analysis of salivary gland, saliva and saliva components of 8 day-old blood-fed Anopheles gambiae 

One-dimensional electrophoresis

## Salivary gland extracts

Two series of experiments were performed. In the first series, $12 \%$ SDS-PAGE gels were run with $10 \mu \mathrm{~g}$ of protein extract obtained from salivary glands of 8 day-old females. After Coomassie staining, protein bands were excised and the tryptic digests were analyzed by MALDI-TOF mass spectrometry. In the second series, $12 \%$ SDS-PAGE gels were run with $36 \mu \mathrm{~g}$ of protein extract. After Coomassie staining, 1 mm -thick plugs were cut from the gel (Figure 1, supplementary Table 1). Protein identification was performed as described in Methods and seventy percent of the bands were identified (Table 1).

## Saliva

A total of 18 saliva samples each from 400 female 8 day-old blood-fed A. gambiae were collected in water. After lyophilization, saliva components were resuspended in water and analyzed by SDS-PAGE and stained with silver nitrate (Figure 2). The stained gel bands were cut and analyzed by mass spectrometry. Five proteins were thereby identified (Table 1).

## Two-dimensional analysis

After 2-D gel electrophoresis of $120 \mu \mathrm{~g}$ of salivary gland proteins, the trypsin-digested spots were analyzed by peptide mass fingerprinting, using Maldi-Tof, or by PSD Maldi-Tof. From the total set of 204 spots (Figure 3), 29 proteins were identified and described (Table 1, supplementary Table 2). MS identification showed that $37 \%$ of these proteins produced several spots during electrophoresis. Spots at varying pIs were found for the putative 5' nucleotidase precursor in the $62-\mathrm{kDa}$ region of the gel (spots 13 to 33 in Figure 3) as well as for the D 7 precursor allergen AED A2 in the $30-\mathrm{kDa}$ region (spots $114-119,121-125$ ) and for D 7 related- 4 protein precursor in the 16 kDa region (spots 171-176). The profile of the 30 kDa protein was of particular interest with its intense spot at 32.5 kDa and a trail of spots with molecular weights between 30 to 20 kDa (Figure 3). According to the Ensembl database (release 35), two forms of the protein exist, including a long mature form of 24732.75 kDa (ensangp000000028522), and a short mature form of 13786.59 kDa (ensangp00000022344); however, only the short form remained in the Ensembl release 43. The proteomic data are consistent with a larger form of the D7 precursor that is processed by proteolytic cleavage. Several other spots identified as being secreted proteins had apparent $M_{r}$ smaller than expected according to their genomic predicted $M_{r}$ in Ensembl (Table 1). This was the case for the 5 'nucleotidase precursor protein that was identified in spots of apparent molecular weights ranging from 62 kDa to 29 kDa (Table 1, Figure 3). To determine whether this range of sizes is due to an artifact that occurs before or during sample preparation, 2-DE profiles of salivary gland extracts obtained after several freeze/thaw cycles were examined. These profiles did not differ from those of extracts obtained after our normal sonication and centrifugation procedure (data not shown). Additionally, the heating stage was not responsible for proteolysis since the numbers of spots observed with heated salivary
gland extracts and those not heated were similar (data not shown). These observations indicate that several secreted proteins may present sequence divergence or be extensively processed and/or post-translationaly modified in A. gambiae salivary glands. This idea is supported by the following points: 1) several proteins were only identified after post source decay; 2) an extensive processing of the human saliva proteome has been described [15].

## Identification of salivary gland components using LC MS/MS

LC MS/MS is an alternative strategy for large-scale protein identification that bypasses the initial protein separation step. It consists of enzymatic cleavage of a complex protein mixture and separation of the resulting peptides by chromatography before tandem mass spectrometry identification. This gel-free strategy has worked for large-scale protein identification of several biochemical systems [16].

Using this system, 30 proteins were identified with confidence (ProtScore cutoff $>$ to $95 \%$ ). Of these 30 proteins, 15 proteins ( $50 \%$ ) were matched with two or more peptides and the other $50 \%$ were identified by a single peptide hit (Table 1). Using this technique, we were able to confirm that there is a problem in the ensangp00000022344 annotation corresponding to our 30 kDa protein, since only two of the five peptide sequences identified by LC MS/MS were present in the current ensangp00000022344 sequence (Ensembl release 43).

Proteome coverage of 8 day-old blood-fed Anopheles gambiae salivary gland Together, the three technologies characterised 55 different proteins, four of which (ensangp00000028522, ensangp00000026134, ensangp00000027538, ensangp00000015472) are no longer present in the latest genome annotation (Ensembl release 43). LC MS/MS and 2-DEMS identified a similar number of proteins and both appear more effective than 1-DE-MS. Thirty
percent of the proteins identified are secreted. Ensangp000000013568, which is predicted to have aspartic-protease activity, is one of the newly identified proteins. Blast analysis has shown that this protein has $88 \%$ sequence identity with protein AAEL006169-PA in the Aedes aegypti genome and is also similar to cathepsin D enzymes of other insects such as Drosophila melanogaster and Bombyx mori. Insect cathepsins D have been shown to be involved in metamorphosis [17] and their levels are modulated in pathogen-infected insect tissues [18].

## Analysis of salivary gland components of 21 day-old blood-fed Anopheles gambiae salivary glands and comparison of salivary gland components from young (8 days) and old (21 days) blood-fed female mosquitoes

Plasmodium berghei development in A. gambiae takes about 14 days from the infective bloodmeal until the parasite is ready to infect its mammalian host. Thus, the proteomic profile of salivary glands may be affected by ageing. To identify the molecular changes that may occur in salivary gland cells, the proteomic profile of 21 day-old female salivary glands was analyzed by LC MS/MS (Table 2). A total of 41 different proteins were characterised (Table 2). Nineteen of these proteins were described at a proteome level for the first time. Ensangp00000029528 (apolipoprotein D precursor), a protein identified as an infection-responsive protein in the Anopheles midgut [19], was one of these proteins described for the first time. iRNA silencing of the midgut transcript encoding APOD resulted in increased Plasmodium levels. Also among the newly identified intracellular proteins, ensangp00000029324 deserves particular attention. This protein belongs to the family of $\alpha 2$-macroglobulins and has $64 \%$ sequence identity with TEP15 in a FASTA comparison. These thioester-containing proteins are protease inhibitors that can play an important role in immune responses. Ensangp00000029324 has $39.42 \%$ sequence identity
with a protein described in Ornithodoros moubata [20]. This $O$. moubata protein is expressed in tick salivary glands, haemocytes and Malpighian tubules and its expression is enhanced in response to a blood meal. Using gene expression screening for immune response genes in the $A$. gambiae transcriptome, Oduol et al. [21] identified an $\alpha 2$-macroglobulin-related molecule that responded strongly to malaria parasite infection. Thus, one could propose that ensangp00000029324 is involved in the defence against pathogens such as parasites, bacteria or fungi.

Using only the LC MS/MS identified proteins, the level of salivary components after 8 days was compared to those after 21 days of salivary gland development. Their functions were compared (Figure 4). The composition of young salivary glands was less diverse than that of older salivary glands: i.e. fewer proteins could be identified in young salivary glands (29) than in older glands (42). Eighteen proteins were common to both salivary gland extracts and most were secreted proteins (30 kDa, apyrase, $5^{\prime}$ nucleotidase, D 7 precursor allergen AED A2, D7r1, D7r2, D7r3 and D7r4, maltase precursor, peroxidase precursor, GSG6, GSG7, putative gVAG); among the other identified proteins, three were glycolytic enzymes (phosphoglycerate mutase, malate dehydrogenase, triosephosphate isomerase), one was an RNAse and another one was an actinbinding protein (ensangp00000012938). All the secreted proteins identified in 8 day-old salivary glands were also found in 21 day-old salivary glands, whereas 6 additional, secreted proteins (D7 r5, GSG5, lysozyme and the hypothetical proteins $8.8,10$ and 10.2 kDa ) were specific to the 21 day-old salivary glands. Protein functions, including transcription, signalling and metabolism, assigned to some of the housekeeping proteins that were found in 8 day-old salivary glands were also identified in 21 day-old salivary glands, although a larger variety of proteins were associated
with signalling and transcription regulation. Additionally, apolipoprotein D , lysozyme and $\alpha 2$ macroglobulin, involved in the response to pathogens, were detected in 21 day-old salivary glands, but were not detected by LC MS/MS in 8 day-old salivary glands. A partial list of agerelated mammalian protein variation from the study of ageing mammalian organs [22] includes proteins involved in: (i) telomere repair, (ii) stress response, (iii) anti-oxidant defence, (iv) nicotinamide deamination, (v) insulin/insulin-like growth factor-1 signalling,,(vi) histone deacetylation, and (vii) regulation of the transcription of specific proteins, such as those involved in pituitary development. Specific age related signatures in the transcriptome of Drosophila body parts have also been investigated [23]. That study showed the presence of up-regulated mRNA levels in the aged thorax, where salivary glands are located, for immune response genes, genes linked to cellular morphogenesis as well as those for actin filament-based processes. Cellular components of the endoplasmic reticulum and the proteasome complex were also overrepresented. Thus, our observations indicating an increased level of a subset of salivary gland proteins is consistent with the transcriptional results observed in Drosophila. Interestingly, proteins involved in lipid metabolisms were only identified in 21 day-old salivary glands. Lipids are known to be important for parasite matabolism. Rosinski-Chupin et al. [11] showed that genes involved in lipid metabolism were up-regulated by Plasmodium berghei. This observation suggests that the maturation of sporozoites may require happening in ageing salivary glands.

## Comparison of infected and non-infected salivary gland composition

iTRAQ, an isotope labelling approach, was used for the quantitative study of gene expression at the proteome level. This approach is based on chemical isobaric tagging of the N -terminus of peptides generated from trypsin digests of proteins isolated from cells or tissues in different
states. The labelled samples are combined, fractionated together by strong cation exchange chromatography and analysed by nanoLC mass spectrometry. The labelled peptides and hence the corresponding proteins are then identified by database searching using the MS/MS data. The fragmentation of the tag attached to the peptides generates a low molecular mass reporter ion which is unique to the tag used. Comparison of the intensities of these reporter ions gives relative protein quantification. Table 3 shows the list of proteins identified and quantified using iTRAQ.

Twelve identical, secreted proteins were found in uninfected and infected salivary glands during three separate comparisons (Table 3). The ratios of reporter ion peaks of infected versus noninfected salivary glands varied between 0.65 and 1.97. From the ratio values, it was deduced that the expression of five of the proteins, was altered. The level of gVAG is increased two-fold in infected salivary glands, whereas the levels of GSG6, apyrase, D7 related-1 protein precursor and D7 precursor allergen AED A2 are decreased with ratios ranging from 0.67 to 0.77 for these proteins (Table 3). The presence of pathogens in salivary glands has been reported to induce modifications in insect behaviour and/or modifications in saliva composition. gVAG is a protein of the antigen 5 family and it has similarities with the mammalian cysteine-rich secretory proteins, vespid antigen 5 and plant-pathogenesis-related proteins [27]. The precise function of these secreted proteins is unknown. The level of gVAG mRNA was shown to be increased in the midgut of mosquitoes infected with Plasmodium falciparum compared to level in uninfected midguts [19]. The silencing of this gene resulted in increased Plasmodium levels, suggesting that gVAG is a defence-related protein [19]. We therefore expect a similar role of gVAG in $A$. gambiae salivary glands.

The level of apyrase was reduced by a factor of 1.5 in $P$. berghei-infected $A$. gambiae salivary
glands. Apyrase inhibits ADP-induced platelet aggregation and, therefore, affects blood-feeding. The level of apyrase influences the probing time of Anopheles gambiae [28]. The reduction of apyrase abundancy by $85 \%$ in salivary glands from A. gambiae due to the injection of doublestranded apyrase encoding RNA was correlated with increased probing time. Decreased apyrase levels in Plasmodium gallinaceum infected Aedes aegypti salivary glands caused an increase in mosquito probing time [24]. An increase in probing time has also been observed for Anopheles gambiae infected with Plasmodium falciparum [29]. Additionally, transcription of the apyrase encoding gene appears to be repressed in $P$. berghei-infected $A$. gambiae mosquitoes [11]. Our observation is, therefore, consistent with these data and we can expect an increase probing time for $P$. berghei-infected A. gambiae.

The levels of D7 precursor allergen AED A2 and D7 related-1 protein precursor proteins were decreased in infected salivary glands by a factor of 1.3 and 1.5 respectively. The D7 short proteins bind serotonin with high affinity, as well as histamine and norepinephrine, thus antagonizing the vasoconstrictor, platelet-aggregating, and pain-inducing level of these factors [30]. The decreased production of D7 related-1 protein precursor may induce an increased local inflammatory response to mosquito bites, thus modifying the immune response to the parasite. Although we did not observe a change in D7 related-4 protein precursor protein levels in our analysis, Rosinski-Chupin et al. [11] observed variable D7 related-4 protein precursor gene expression using SAGE. The proteins D7 precursor allergen AED A2 and GSG6 have no known function, thus we cannot anticipate the consequence of reducing their expression on parasite development and transmission.

Table 3 also shows that the iTRAQ technique identified forty three proteins not observed using
by LC MS/MS analysis of salivary gland extracts from insects of the same age. This result is consistent with the previous observations that better fragmentation is obtained using this technology, giving more peptides per protein and allowing the identification of less abundant proteins [31, 32]. One of the newly identified proteins was a homolog of "translationally controlled tumour protein" (Tctp). Tctp homologues have been described in mammals and in many other species, including plants, earthworm, parasites, hydra and yeast [33-38]. They are heat stable, calcium-binding proteins [39] and their expression is induced in response to various stimuli within cells [38]. Tctps also bind haem, and tubulin [40]. Tctps induce the release of histamine [41] and the secretion of interleukin-4 [42] from basophils. Despite having a ubiquitous tissue distribution, multiple specific potencies [43] and highly conserved amino acid sequences, their primary physiological role remains unclear [40]. The A. gambiae protein has the highest identity scores with its Aedes albopictus and Aedes aegypti homologs (85\%). An identity score of $44 \%$ was observed with the ticks Dermacentor and Ixodes salivary histamine-releasing factors (HRF) [44, 45]. The tick HRF recombinant protein induced histamine secretion from a rat basophilic leukaemic cell line, in a dose-dependent manner. We suggest that the Anopheles gambiae Tctp homolog is present in saliva and contributes to the allergic inflammation associated with the Anopheles gambiae bite. Thus, if it is similarly able to trigger cutaneous mast cell histamine release, as observed with the Schistosoma mansoni Tctp homolog [46], the resulting vasodilation could facilitate Plasmodium sporozoite migration into blood vessels.

Our data also identify Serpin 9 (ensangp00000016680) at the proteomic level for the first time. Serpins are a very large family of serine protease inhibitors with various biological functions that are found in all higher eukaryotes and viruses [47]. The mosquito genome contains 14 annotated serpin genes, 10 of which are inhibitory protease substrates. Some of these serpins are involved
in immune signal amplification cascades. Serpin 9 is involved in the arthropod immune response and during Staphylococcus aureus infection, it is only induced late in infection [48]. In contrast, during the Plasmodium life cycle in mosquitoes, serpin 9 is primarily activated when the midgut epithelium is invaded by ookinetes [48]. However, a tag corresponding to Serpin 9 was identified in the A. gambiae salivary gland using SAGE [11], but the level of this tag was not modified by Plasmodium infection.

## CONCLUDING REMARKS

In this study, complementary proteomic approaches were used to catalogue 122 Anopheles gambiae salivary gland proteins from blood-fed 8 day-old and 21 day-old females (supplementary Table 3). The most acidic proteins identified were the 30 kDa protein ( $\mathrm{pI}, 3.8$ ) and calmodulin ( $\mathrm{pI}, 3.9$ ) and the most basic proteins were retrovirus-related pol polyprotein (pI, 11.28) and ensangp00000015472 ( $\mathrm{pI}, 10.38$ ). The smallest proteins identified were hypothetical $8.8 \mathrm{kDa}\left(\mathrm{M}_{\mathrm{r}}, 8.8 \mathrm{kDa}\right)$ and retrovirus related pol polyprotein $\left(\mathrm{M}_{\mathrm{r}}, 9.6 \mathrm{kDa}\right)$ and the largest was Ryanodin receptor $1\left(\mathrm{M}_{\mathrm{r}}, 577.8 \mathrm{kDa}\right)$. Our approach confirmed the presence of seven proteins identified in earlier Ensembl annotations but not listed in the latest version (version 43). This observation emphasizes the complementarity of proteomic and genomic approaches for accurate genome annotation, an idea previously suggested by Kalume et al. [49].

LC MS/MS was clearly the most powerful technique (Figure 6A). iTRAQ labelling led to the identification of 78 proteins, 39 of which were not identified by classical LC MS/MS, illustrating the value of using the two technologies in parallel for maximum proteome coverage. The proteins identified in this study were sorted into functional categories based on their annotations in the database and the results are summarised in Figure 6B. A large proportion of the identified proteins are involved in energy pathways, blood or sugar feeding, protein folding, modification and in amino acid metabolism, but the largest group (37\%) is composed of proteins with no known function. The same situation is also encountered in the proteomic analysis of human saliva [50]. In Anopheles gambiae, twenty five percent of the identified proteins are predicted to have a signal sequence and are, therefore, putatively present in saliva. The largest category of peptide sequences was that derived from secreted proteins, demonstrating that they are the most
abundant proteins in salivary gland extracts. This observation is consistent with the findings of Kalume et al. [12].

Seventy-five percent of the 122 proteins reported here are identified in an Anopheles gambiae salivary gland proteomic study for the first time. Most of these newly identified proteins are housekeeping proteins and only few, such as GSG5, GSG3, ensangp00000029324, serpin 9, hypothetical 10 kDa and apolipoprotein D precursor, are secreted. The 2- D gel analysis suggests that some secreted proteins, including 5'nucleotidase, D7 precursor allergen AED A2, D7 related-4 protein precursor and 30 kDa , are extensively processed, although the consequence of such modifications on their activity is unknown. LC/MS-MS profiling of young versus old salivary gland proteomes suggests that there is an over-representation of proteins involved in signalling, proteins implied in carbohydrate and lipid metabolism and proteins related to the immune response in older glands. As the invasion and the maturation of sporozoites occurs during the ageing process of salivary glands, it would be interesting to know whether the age of the salivary gland affects parasite transmission. Finally, we detected a change in the level of five salivary proteins in the presence of Plasmodium berghei sporozoites. These observations will serve as a basis for future work to determine the possible role of these proteins in the interaction between A. gambiae, Plasmodium and the mammal host.

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## LEGENDS TO FIGURES

## Figure 1: SDS-PAGE of salivary gland extracts from 8 day-old blood-fed A. gambiae

Salivary components were separated by a $12 \%$ NU-PAGE Bis-Tris gel under denaturating and reducing conditions. Molecular mass markers are shown on the left. After Coomassie staining, the gel was cut into millimeter slices as indicated by the numbers on the right side of the figure. The plugs obtained were analyzed by mass spectrometry as described in the Methods section.

Figure 2: SDS-PAGE of 8 day-old blood-fed A. gambiae saliva
Saliva was collected from 7200 females using artificial feeders. After lyophilisation, saliva components were re-suspended in water and aliquots were analyzed by SDS-PAGE. Following silver nitrate staining, the numbered protein bands were analyzed by mass spectrometry.

Figure 3: 2-DE analysis of salivary gland extracts from 8 day-old blood-fed A. gambiae Salivary gland extracts were purified by ReadyPrep 2D Cleanup kit and $120 \mu \mathrm{~g}$ of proteins were solubilized in 2D sample buffer, as described in the Methods section. Proteins were separated in the first dimension using carrier ampholyte gradient gels between pH 4 and pH 8 . Separation in the second dimension was performed using $12.5 \%$ SDS acrylamide gel. The gel was stained using SYPRO® Ruby.

Figure 4: Comparison of 8 day-old and 21 day-old salivary component functional annotations.
A) 8 day-old salivary gland components; B) 21 day-old salivary gland components.

Figure 5: Functional annotation of the 122 salivary components identified in $\mathbf{8}$ day-old and 21 day-old blood-fed Anopheles gambiae.
A) Contribution of various proteomic approaches to protein identification; B) Biological processes in which they are involved.

Table 1 : Proteins identified in salivary gland extracts of 8 day-old blood fed Anopheles gambiae

| Ensembl | Protein | Predicted | Identification | 1-DE-MS | 2-D | -MS | LC MS/MS | Comments | Subcell | Found in |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Identification (Ensembl release 43) | Family/Description | $M_{\mathrm{r}} / \mathrm{pI}$ |  | \% coverage | spot number | \% coverage ${ }^{\text {a) }}$ | Peptide sequence |  | localization <br> b) | other <br> proteomic ( P ) <br> or <br> transcriptomic <br> (T) studies |
| Ensangp00000028522 ${ }^{\text {c }}$ | 30 kDa protein | 26.9/3.8 | 2-DE-MS | - | $\begin{aligned} & \hline 84,184- \\ & 188,190, \\ & 192-193, \\ & 196-199 \\ & 201-202 \end{aligned}$ | PSD | EQELSDCIVDKR <br> IKECFSSLDK <br> ELDDGLIEREQELSDCIVDK <br> LMNPTIDLVSTIEKYSK <br> ECFSSLDKDVSAMVK <br> KDDAEEDSEEGGEEGGDGASG <br> G <br> EGGEKESPR | GE rich salivary gland | secreted | P [12], [9] |
| Ensangp00000018590 | 5 aminolevulinate synthase erythroid specific mitochondrial precursor * | 46.31/7.54 | 2-DE-MS | - | 110 | 25 | - | Metabolism of amino acid | mitochondria 1 matrix | this work |
| Ensangp00000015067 | Ambiguous* | 35.7/10.4 | 2-DE-MS | - | 186 | 16 | - | ? | mitochondria 1 | this work |
| Ensangp00000015256 | Ambiguous/candidate odorant receptor* | 44.85/7.01 | LC MS/MS | - | - | - | AQRPVGITAGK | Olfactory receptor (drosophila) | membranar | this work |
| Ensangp00000022917 | Ambiguous* | 72.38/10.16 | LC MS/MS | - | - | - | GRPILPLLKTVQSYK | Tropomyosin domain | intracellular | this work |
| Ensangp00000024702 | Ambiguous* | 30.31/9.58 | LC MS/MS | - | - | - | IHDGVTHAAK | ? | ? | this work |
| Ensangp00000026134 ${ }^{\text {c }}$ | Ambiguous* | 23.01/10 | 2-DE-MS | - | 169 | PSD | - | ? | ? | this work |
| Ensangp00000015382 | Apyrase | 61.79/8.6 | $\begin{aligned} & \text { 1-DE-MS, LC } \\ & \text { MS/MS, } \end{aligned}$ | 20\% | - | - | AAEEGDTCIAGIAR LNVAQVAGLR GDITNEEAIGASPFSNTVDLLT LR | Anti-platelet | secreted | P [12] |
| Ensangp00000011707 | Aspartate amino transferase* | 44.71/6.78 | 2-DE-MS | - | 95 | 17 | - | Metabolism of amino acid | cytoplasmic | this work |
| Ensangp00000024137 <br> and/or <br> Ensangp00000016868 ${ }^{\text {d) }}$ | ATP synthase subunit beta mitochondrial precursor | $22.69 / 4.9$ <br> and/or $19.72 / 5.27$ | $\begin{aligned} & \text { 2-DE-MS, LC } \\ & \text { MS/MS } \end{aligned}$ | - | 66-67 | (31-37) | IINVIGEPIDER <br> LVLEVAQHLGENTVR | Catalyzes ATP synthesis | mitochondria 1 | P [12] |
| Ensangp00000018543 | Chromosome associated polypeptide C XCAP C homolog | 156.83/5.34 | LC MS/MS | - | - | - | LQTELIELKR | Structural maintenance of chromosome ABC transporter | nuclear | this work |


| Ensangp00000003518 | CoA carboxylase | 130.5/6.67 | 1-DE-MS | 15\% | - | - |  | related domain Key enzyme in | mitochondria | this work |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000003518 | mitochondrial precursor* | 130.5/6.67 | 1-DE-MS |  |  |  |  | the catabolic <br> pathway of odd- <br> chain amino <br> acids : <br> isoleucine, threonine, methionine and valine | 1 matrix | this work |
| Ensangp00000026391 | Cofilin | 16.93/7.28 | 2-DE-MS, LC MS/MS | - | 170 | 42 | LFLMSWCPDTAK | Binds actin and assists in translocation of actin from the cytoplasm to the nucleus essential for cytokinesis, endocytosis and other cell processes that require rapid turnover of actin filaments | cytoplasmic | T [9] |
| Ensangp00000022538 | Creatine kinase | 26.4/5.18 | 2-DE-MS | - | 90 | 30 | - | Phosphorylation | cytoplasmic | P [12] |
| Ensangp00000025174 and/or | D7 precursor allergen AED A2 | $\begin{aligned} & 35.57 / 5.7 \\ & \text { and/or } \end{aligned}$ | $\begin{aligned} & \text { 1-DE-MS, 2- } \\ & \text { DE-MS, LC } \end{aligned}$ | 42\% | 114-119, | $\begin{aligned} & (19-33) \\ & \text { PSD } \end{aligned}$ | ALDPEEAWYVYER BVLIGLQLYEEK | ? | secreted | P [12], [9] |
| Ensangp00000018280 ${ }^{\text {d }}$ |  | 32.7/5.1 | MS/MS |  | 121-125, |  | NYELSGSSQFK SADYAFLLR |  |  |  |
|  |  |  |  |  | 149-151, |  | SANYGYLAMGK |  |  |  |
|  |  |  |  |  |  |  | SDLEPEVR |  |  |  |
|  |  |  |  |  | 154, 169 |  | SVLASCTGTQAYDYYSCLLNS |  |  |  |
|  |  |  |  |  |  |  | PVK |  |  |  |
|  |  |  |  |  |  |  | DYELADSAEFR |  |  |  |
|  |  |  |  |  |  |  | IYHGTVDSVAK |  |  |  |
|  |  |  |  |  |  |  | NAFYFHELR |  |  |  |
|  |  |  |  |  |  |  | NAMDCVFR |  |  |  |


| Ensangp00000018340* | D7 related-1 protein precursor | 18.73/9.57 | $\begin{aligned} & \text { 1-DE-MS, LC } \\ & \text { MS/MS } \end{aligned}$ | 20\% | $\stackrel{-}{-}$ | ${ }^{-}$ | BLVESTSGEAFK <br> KLPALSQYSSVVDK <br> KVFDTVELVK <br> CLVESTSGEAFK | Antiinflammatory Scavenger of biogenic amines | secreted | P [12], [9] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000018371* | D7 related-2 protein precursor | 18.46/4.8 | 1-DE-MS, 2-DE-MS, LC MS/MS | 28\% | 181-183 | PSD | ANTFYTCFLGTSSLAGFK ESVLLELLQR HMQBVLEVVGFVDGNGEVK KANTFYTCFLGTSSLAGFK MQTSDPFDMNR NAVDYNELLK QYTPVSSDDMDK | Antiinflammatory | secreted | P [12], [9] |
| Ensangp00000018330 | D7 related-3 protein precursor | 19.7/4.38 | 1-DE-MS, 2-DE-MS, LC MS/MS | 33\% | 180-181 | PSD | ANTFYTCFLGTSSAQAFK AGKLDMGTTFNAGQVSALMK LDMGTTFNAGQVSALMK YAVDYVELLR | Antiinflammatory | secreted | P [12], [9] |
| Ensangp00000018328 | D7 related-4 protein precursor | 19.29/7.4 | $\begin{aligned} & \text { 2-DE-MS, LC } \\ & \text { MS/MS } \end{aligned}$ | 30\% | 171-176 | (22-40) | LYDPLNIIELDK CIGECVQVPTSER <br> RYEIIEGPEMDK <br> YTAEFVQIMK <br> VFDLMELK | Antiinflammatory | secreted | P [12] |
| Ensangp00000027211 | Disulfide isomerase precursor | 54.31/5.47 | 2-DE-MS | - | 52 | 15 | - | Catalyzes the rearrangement of -s-s- bonds in proteins | intracellular | P [12] |
| Ensangp00000014287 | Electron transfer flavoprotein alpha subunit mitochondrial pecursor* | 34.14/8.62 | 2-DE-MS | - | 113 | 33 |  | Participates in catalyzing the initial step of the mitochondrial fatty acid betaoxidation | mitochondria 1 | this work |
| Ensangp00000003806 | Facilitated glucose transporter | 16.83/8.48 | LC MS/MS | - | - | - | HISQIVPLVAKGFSSKPLVP | Sugar transporter | membranar | this work T [9] |
| Ensangp00000000937 | probable Fatty acid binding protein | 19.37/9.59 | LC MS/MS | - | - | - | LGGGFDEETVDGR | Fatty acid binding protein | cytoplasmic | this work |
| Ensangp00000016366 | Precursor | 45.95/9.43 | 2-DE-MS | - | 142 | 23 | - | Involved in energy pathways | cytoplasmic | this work |
| Ensangp00000011661 | Glutathion S transferase (class theta) | 23.78/6.51 | 2-DE-MS | - | 155 | 33 | - | Key role in cellular detoxification | cytoplasmic and nuclear | This work $\underline{P}[50]$ |
| Ensangp00000024808 | Glutathion S transferase | 23.44/6.26 | 2-DE-MS | - | 156 | 23 | - | « | « | this work |
| Ensangp00000010081 | Glycogen phosphorylase | 96.48/6.33 | 1-DE-MS | 18\% | - | - | - | Carbohydrate metabolism | cytoplasmic | this work |


| Ensangp00000009988 | GSG3 | 20.01/4.34 | 2-DE-MS |  | 75,76 | PSD | - | ? | secreted | $\begin{aligned} & \text { this work } \\ & \mathrm{T}[51] \end{aligned}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000019455 | GSG6 | 13.05/5.15 | 1-DE-MS, LC MS/MS | 36\% | - | - | EPLPYMYACPGTEPCQSSDR ETREPLPYMYACPGTEPCQSS DR | ? | secreted | P [12], [9] |
| Ensangp00000021970 | GSG7 | 16.29/8.46 | LC MS/MS, | - | - | - | SMHDVLCDRIDQAFLEQ <br> TLADETAQCMR <br> TLADETAQCLR <br> YGVQNQLR | ? | secreted | P [12] |
| Ensangp00000005326 | Guanine nucleotide releasing factor | 137.53/9.17 | LC MS/MS | - | - | - | LIEKALIYK | May play a role in intracellular signaling cascade | M embraneassociated | this work |
| Ensangp00000021028* | putative gVAG protein precursor | 28.9/8.96 | 1-DE-MS, LC MS/MS | 43\% |  | ${ }^{-}$ | DGQMDVYYFVBNYSFTNIMD R <br> FPYAGQNIAITQFFGYR FVSSWWSEYLDARPEHVR GGPHVGCNPPSSSGGPTCQGK KYPSSYSGKPIGHFTQIASDR MPTLTWDPELASLADANAR VGCSMWYWK | Allergen Belongs to the CAP family: protease inhibitors or proteolytic activity, probably inhibiting host coagulation or complement activity Defence-related protein | secreted | P [12], [9] |
| Ensangp00000017720 | 3 Hydroxyisobutyrate dehydrogenase mitochondrial | 34.31/9.27 | LC MS/MS | - | - | - | VFADIVNASTGR | Involved in amino acid catabolism pathway | mitochondria <br> 1 | this work |
| Ensangp00000016660 | Isocitrate dehydrogenase | 46.96/7.59 | 1-DE-MS | 32\% | - | - | - | Plays a key role in cellular defense against oxidative stressinduced damage | mitochondria <br> 1 | this work |
| Ensangp00000020184 | Malate dehydrogenase | 35.27/9.52 | LC MS/MS | - | - | ${ }^{-}$ | ANTFVGEAAGVDPQK | Metabolic enzymes which catalyse the last step in anaerobic glycolysis | mitochondria <br> 1 | P [12] |
| Ensangp00000011006 | Malate dehydrogenase | 35.37/6.95 | 2-DE-MS |  | 96 | PSD | DDLFNTNASIVR | Participates in the citric acid cycle | cytoplasmic | this work |
| Ensangp00000017682 | Maltase | 67.21/5.87 | 1-DE-MS, 2- | 27\% | 6-8, 12 | (17-43) | AMPSGAIANWVLGNHDNSR | Carbohydrate | secreted | P [12], |


|  |  |  | $\begin{aligned} & \text { DE-MS, LC } \\ & \text { MS/MS } \end{aligned}$ |  |  |  | DQPETYDMVHQWR ELNVAAQLAAPR GITQTIDYLK | digestion Converts sucrose in nectar to glucose and fructose |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000015067 <br> Ensangp00000011253 | Mitochondrial carrier Nucleoside diphosphate kinase | $\begin{aligned} & 35.74 / 10.4 \\ & 19.01 / 8.46 \end{aligned}$ | 2-DE-MS 1DE-MS, | $32 \%$ | $186$ | $16$ | GDLCVQVGR | Maintenance of cellular pool of nucleoside triphosphates | cytoplasmic and plasma membrane | T [9] |
| Ensangp00000012716* | putative 5' Nucleotidase precursor | 63.47/7.01 | 1-DE-MS, 2-DE-MS, LC MS/MS | 20\% | $\begin{aligned} & 10,13- \\ & 33,38- \\ & 51,54, \\ & 57,60, \\ & 64,65, \\ & 77-82, \\ & 85-87, \\ & 130-134, \\ & 140-14, \\ & 144-145 \end{aligned}$ | $\begin{aligned} & (15-30) \\ & \text { PSD } \end{aligned}$ | APFPLTLIHINDLHAR <br> DQIYYVVVPSYLADGKDGFA <br> MK <br> ECIAGIAR <br> GLAPYLAELEK <br> LGTQVIGTTEVFLDRESCR <br> LSGADLWSAIDHSFTLDDEFR <br> MKIPTVVANLEK <br> NVNIIVVLSHCGLDGDK <br> QLAEEAGDLIDVIVGAHSHSLL LNK | Anti-platelet | secreted | P [12] |
| Ensangp00000028058 | Peroxidase precursor | 24.99/8.23 | $\begin{aligned} & \text { 1-DE-MS, LC } \\ & \text { MS/MS } \end{aligned}$ | 16\% | - |  | AFAGAININDHMFNPTVLER CFAIPVRPDDPVLSAGGIQCLD LVR LLPAEYGDGVYVPR SNITPELTILHVAFLR TTLVNMQFGQLVAHDMGLR WEDFVELR | Vasodilatator | secreted | P [12], [50] |
| Ensangp00000012460 | Phosphoglycerate kinase | 43.84/7.54 | 2-DE-MS |  | 109 | 27 | - | Glycolysis | cytoplasmic | this work |
| Ensangp00000015800 | Phosphoglycerate mutase | 28.7/6.8 | $\begin{aligned} & \text { 2-DE-MS, LC } \\ & \text { MS/MS } \end{aligned}$ | - | 148 | 25 | YGEEQVLIWR | Involved in energy pathways | cytoplasmic | this work |
| Ensangp00000012492 | Precursor | 12.39/8.75 | 1-DE-MS | 22\% | - | - | - | EGF-like domain | ? | this work |
| Ensangp00000013568 | Precursor | 41.83/5.4 | 2-DE-MS | - | 75 | - | - | Aspartic protease A1 | secreted | this work |
| Ensangp00000016366 | Precursor | 45.95/9.43 | 2-DE-MS | - | 142 | 23 | - | Glucose-methanol-choline oxidoreductase Involved in energy pathways | cytoplasmic | this work |
| Ensangp00000019046 | Precursor | 28.47/5.04 | LC MS/MS | - | - | - | ANDRAMVK | EGF-like domain | ? | this work |


| Ensangp00000020734 | Pterin 4 alpha carbinol amine dehydratase | 21.20/10.23 | LC MS/MS | - | - | - | LAQFLDQAAAVAK | Transcriptional activator/pterin dehydratase |  | this work |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000027538 ${ }^{\text {c }}$ | Retrovirus related pol polyprotein | 9.51/11.28 | 2-DE-MS | - | 181, 183 | PSD | - | ? | nuclear | this work |
| Ensangp00000021077 | Ribonuclease | 14.41/8.04 | LC MS/MS |  |  |  | ALAPYNQAIVADR | Inhibits protein synthesis by cleavage of mRNA | ? | this work |
| Ensangp00000027418* | Salivary gland 1-like 3 | 44.51/6.04 | 1-DE-MS | 30\% | - | - | - | ? | secreted | P [12] |
| Ensangp00000018041 | Toll precursor | 16.69/4.51 | 2-DE-MS | - | 152 | 17 | - | Toll IA <br> Involved in signal transduction pathways in response to pathogens | plasma membrane | P [50] |
| Ensangp00000018152 | Triosephosphate isomerase | 26.3/6.2 | 1-DE-MS, 2- <br> DE-MS, LC <br> MS/MS | 30\% | - | - | AIFGETDELIAEK DWSNVVIAYEPVWAIGTGK SLLPETIGVAAQNCYK DLGLGWVILGHSER | Central enzyme in the glycolytic pathway Plays an important role in several metabolic pathways | cytoplasmic | this work P[52] |
| Ensangp00000012072 | Unknown | 29.21/4.43 | 2-DE-MS | - | 135 | 20 | DSTLIMQLLR | 14-3-3 protein. <br> Family of conserved regulatory molecules that bind a multitude of functionally diverse signaling proteins | cytoplasmic | P [12] |
| Ensangp00000015472 ${ }^{\text {c }}$ | Unknown | 15.64/10.38 | 1-DE-MS | 20\% | - | ${ }^{-}$ | - | InterPro <br> Zn-finger, C2H2 <br> type <br> nucleic acid- <br> binding protein | nuclear? | $\begin{aligned} & \text { this work } \\ & \text { P [50], T [9] } \end{aligned}$ |
| Ensangp00000019887 | Unknown | 70.9/5.1 | 2-DE-MS | - | 9 | 18 | - | Heat shock 70 region May be involved in response to | cytoplasmic and organelles | P [12] |


${ }^{\text {a) }}$ When several spots corresponded to the same protein, the percentage range of the sequence coverage is indicated in parenthesis. ${ }^{\text {b) }}$ Subcellular localization is inferred from sequence or structure similarity with orthologous proteins. ${ }^{\text {c) }}$ Identification was performed using Ensembl database v35 of november 2005 . ${ }^{\text {d) }}$ Cases where the same peptides match more than one genomic sequence. Shaded lines: proteins identified for the first time by a proteomic approach. $*$ means that the proteins were also identified in saliva. References underlined correspond to proteins found in human saliva.

Table 2 : Proteins identified by LC MS/MS in salivary glands of 21 day-old blood-fed Anopheles gambiae

| Ensembl <br> Identification <br> (Ensembl release 43) | Protein <br> Family/Description | Predicted $\mathrm{Mr} / \mathrm{pI}$ | Peptide sequence | Comments | Subcellular Localization a) | Found in other proteomic (P) or transcriptomic (T) studies |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\underset{\text { b), c), d) }}{\text { Ensangp00000028522 }}$ | 30 kDa protein | 26.90/3.8 | EQELSDCIVDKR IKECFSSLDK <br> ELDDGLIEREQELSDCIVDK LMNPTIDLVSTIEKYSK ECFSSLDKDVSAMVK | GE rich salivary gland | secreted | P [12] P [9] |
| Ensangp00000018525 | Aconitate hydratase mitochondrial precursor | 82.65/8.63 | FDQNVYLPYEK ISILGLNNFAPGK | Iron-sulphur proteins that function as electron carriers biosynthesis of aminoacid | mitochondrial | this work |
| Ensangp00000016546 | Ambiguous | 25.56/9.94 | KGIGTHLMITLEVLAR | GCN5-related N acetyltransferase Putative role in transcription and DNA repair | ? | this work |
| Ensangp00000026066 <br> b) | Ambiguous | 25.13/7.06 | MSDKVVSSFLR | ? | ? | this work |
| Ensangp00000027299 | Ambiguous | 339.53/6.98 | EILYDDIERPILQTK <br> LAGVFTPQEPLMNYVISCWVR QIVTFPDEER <br> TAYLYDPQDVQLSVDGIVFR <br> TFDETWATLAVR <br> YPFGAGGEPFR <br> LYFFASK | Subtilase serine protease domain ? proteasome | cytoplasmic | this work |
| Ensangp00000029258 | Apolipoprotein D precursor | 26.11/4.55 | QSDVGRAVVAFPDESPLEAK | Extracellular ligand-binding proteins displaying high specificity for small hydrophobic molecules response to pathogens | secreted | this work |
| Ensangp00000015382 ${ }^{\text {c }}$ | Apyrase | 61.79/8.6 | AAEEGDTCIAGIAR LNVAQVAGLR <br> GDITNEEAIGASPFSNTVDLLTLR | Anti-platelet | secreted | P [12] |
| $\underset{\text { b),c) }}{\text { Ensangp000000026391 }}$ | Cofilin | 16.93/7.28 | LFLMSWCPDTAK | Binds actin and assists in translocation of actin from the cytoplasm to the nucleus essential for cytokinesis, endocytosis and other cell processes that require rapid | cytoplasmic | T [9] |


|  |  |  |  | turnover of actin filaments |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000025174 ${ }^{\text {c }}$ | D7 precursor allergen AED A2 | $\begin{aligned} & 35.57 / 5.7 \\ & \text { ou } \\ & 32.7 / 5.1 \end{aligned}$ | ALDPEEAWYVYER <br> BVLIGLQLYEEK <br> NYELSGSSQFK <br> SADYAFLLR <br> SANYGYLAMGK <br> SDLEPEVR <br> SVLASCTGTQAYDYYSCLLNSP <br> VK <br> DYELADSAEFR <br> IYHGTVDSVAK <br> NAFYFHELR <br> NAMDCVFR | ? | secreted | P [12], P [9] |
| ${ }_{\text {) }}$ Ensangp00000018340 ${ }^{\text {C }}$ | D7 related-1 protein precursor | 18.73/9.57 | BLVESTSGEAFK <br> KLPALSQYSSVVDK <br> KVFDTVELVK <br> CLVESTSGEAFK | Anti-inflammatory Scavenger of biogenic amines | secreted | P [12], P [9] |
| Ensangp00000018371 ${ }^{\text {c }}$ | D7 related-2 protein precursor | 18.46/4.8 | ANTFYTCFLGTSSLAGFK ESVLLELLQR HMQBVLEVVGFVDGNGEVK KANTFYTCFLGTSSLAGFK MQTSDPFDMNR NAVDYNELLK QYTPVSSDDMDK | Anti-inflammatory Scavenger of biogenic amines | secreted | P [12], P [9] |
| ${ }_{5}$ Ensangp00000018330 ${ }^{\text {c }}$ | D7 related-3 protein precursor | 19.66/4.46 | ANTFYTCFLGTSSAQAFK AGKLDMGTTFNAGQVSALMK <br> LDMGTTFNAGQVSALMK <br> YAVDYVELLR | Anti-inflammatory Scavenger of biogenic amines | secreted | P [12], P [9] |
| Ensangp00000018328 ${ }^{\text {c }}$ | D7 related-4 protein precursor | 19.29/7.4 | LYDPLNIIELDK CIGECVQVPTSER RYEIIEGPEMDK YTAEFVQIMK VFDLMELK | Anti-inflammatory Scavenger of biogenic amines | secreted | P [12] |
| Ensangp00000018321 | D7 related-5 protein precursor | 18.79/5.82 | SGSFFSCMLR | $?$ | secreted | P [12] |
| Ensangp00000003578 | GSG5 precursor | 38.2/6.42 | TYFQNEFVEYR | ? | secreted | T [51] |
| Ensangp00000019455 ${ }^{\text {c }}$ | GSG6 | 13.05/5.15 | EPLPYMYACPGTEPCQSSDR ETREPLPYMYACPGTEPCQSSDR SMHDVLCDRIDQAFLEQ | ? | secreted | P [12], P [9] |
| ${ }_{\text {) }}$ Ensangp00000021970 ${ }^{\text {c }}$ | GSG7 | 16.29/8.46 | TLADETAQCMR TLADETAQCLR | ? | secreted | P [12] |


| Ensangp00000021028 ${ }^{\text {c }}$ | putative gVAG protein precursor | 28.9/8.96 | $\begin{aligned} & \text { YGVQNQLR } \\ & \text { DGQMDVYYFVBNYSFTNIMDR } \end{aligned}$ | Allergen <br> Belongs to the CAP family: <br> protease inhibitors or proteolytic activity, probably inhibiting host coagulation or complement activity Defence-related protein | secreted | P [12], P [9] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | FPYAGQNIAITQFFGYR |  |  |  |
| ) |  |  | FVSSWWSEYLDARPEHVR |  |  |  |
|  |  |  | GGPHVGCNPPSSSGGPTCQGK |  |  |  |
|  |  |  | KYPSSYSGKPIGHFTQIASDR |  |  |  |
|  |  |  | MPTLTWDPELASLADANAR |  |  |  |
|  |  |  | VGCSMWYWK |  |  |  |
| Ensangp00000009655 | Homolog | 118.45/6.27 | DGKELDLVCMQK | C2 domain (cellular proteins involved in signal transduction or membrane trafficking) Cytochrome c heme-binding site (electron-transfer proteins) | ? | this work |
|  |  |  |  |  |  |  |
| Ensangp00000018375 | Hypothetical 10 kD protein | 10/6.22 | LSLQLEEFAVCK <br> AISDLQQGLFDLNHCTK | ? | secreted | this work |
| Ensangp00000018379 | Hypothetical 10.2 kD protein | 10.13/4.52 | LQQMVEDFTACR | ? | secreted | P [12] |
| Ensangp00000004315 | Hypothetical 8.8 kDa | 8.82/4.05 | DKPDIDPVDFLVDVIK | ? | secreted | P [12] |
| Ensangp00000020384 | Low density lipoprotein receptor | 17.3/5.04 | CISRAGICDGK | Lipid metabolism | membranar | P [50] |
| Ensangp00000022875 | Lysozyme precursor | 15.33/8.56 | NGSTDYGIFQINNK | Immunity related Antibacterial enzyme | secreted | P [12], P [50] |
|  |  |  | YWBDSGYGSNDCK |  |  |  |
|  |  |  | NLLNDDITDDIK |  |  |  |
|  |  |  | KLPNVSSCF |  |  |  |
| Ensangp00000020184 ${ }^{\text {c }}$ | Malate dehydrogenase | 35.27/9.52 | ANTFVGEAAGVDPQK | Metabolic enzymes which catalyse the last step in anaerobic glycolysis | mitochondrial | P [12] |
| Ensangp00000017682 ${ }^{\text {c }}$ | Maltase | 67.21/5.87 | AMPSGAIANWVLGNHDNSR DQPETYDMVHQWR ELNVAAQLAAPR GITQTIDYLK | Sugar digestion Converts sucrose in nectar to glucose and fructose | secreted | P [12] |
| Ensangp00000004215 | Mitogen activated kinase kinase kinase kinase | 159.64/10.04 | NIATYYGAFIK | Protein kinase ATP binding | cytoplasmic | this work |
| Ensangp00000003978 | N acylneuraminate cytidyltransferase | 21.1/5.67 | HLTLARILLGME | Forms CMP-NeuAc, the nucleotide sugar donor used by sialyltransferases (modification may be important in pathogenesis) | cytoplasmic | this work |
| Ensangp00000021120 | NADPH dependent carbonyl reductase | 27.05/7.73 | MDFTGKVVLITGASSGIGASTAK | Sugar metabolism | cytoplasmic | this work |


| $\underset{)}{\text { Ensangp000000012716c }}$ | Putative 5' nucleotidase precursor | 63.47/7.01 | APFPLTLIHINDLHAR <br> DQIYYVVVPSYLADGKDGFAM <br> K <br> ECIAGIAR <br> GLAPYLAELEK <br> LGTQVIGTTEVFLDRESCR <br> LSGADLWSAIDHSFTLDDEFR <br> MKIPTVVANLEK <br> NVNIIVVLSHCGLDGDK <br> QLAEEAGDLIDVIVGAHSHSLLL <br> NK | Anti-platelet | secreted | P [12] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000020778 | Peptidyl prolyl cis trans isomerase | 18.29/8.97 | $\begin{aligned} & \text { FFDMTVDNQPLGR } \\ & \text { IVIELRPDVVPK } \\ & \text { HVVFGSVVEGMDVVR } \end{aligned}$ | Accelerates protein folding | cytopasmic | this work |
| Ensangp00000028058c | Peroxidase precursor | 24.99/8.23 | AFAGAININDHMFNPTVLER CFAIPVRPDDPVLSAGGIQCLDL VR <br> LLPAEYGDGVYVPR <br> SNITPELTILHVAFLR <br> TTLVNMQFGQLVAHDMGLR WEDFVELR | Vasodilatator | secreted | P [12], P [50] |
| Ensangp00000015800́ㅗ | Phosphoglycerate mutase | 28.7/6.8 | YGEEQVLIWR | Involved in energy pathways | cytoplasmic | this work Table 1 |
| Ensangp00000029324 | Precursor | 25.94/4.8 | TLTFVLKPTK | Alpha 2 macroglobulin domain | intracellular | this work |
| ${ }_{\text {E }}^{\text {Ensangp0000002 }}$ | Ribonuclease | 14.41/8.04 | ALAPYNQAIVADR | Inhibits protein synthesis by cleavage of mRNA | ? | this work Table 1 |
| Ensangp00000019607 | Ryanodine receptor 1 | 577.53/5.18 | YFDMFLKLK | $\mathrm{Ca} 2+$ release channels involved in secretory pathways? | membranar | this work |
| Ensangp00000008103 | Stromal interaction molecule precursor | 54.49/6.36 | DVEGLLKAEVALK | ? | membranar | this work |
| Ensangp00000028309 | Trans enoyl COA isomerase mitochondrial precursor | 30.18/7.13 | ALEQAVAFLNR | Fatty acid metabolism | mitochondrial | this work |
| Ensangp00000018152c | Triosephosphate isomerase | 22.52/5.09 | AIFGETDELIAEK DWSNVVIAYEPVWAIGTGK SLLPETIGVAAQNCYK DLGLGWVILGHSER | Central enzyme in the glycolytic pathway Plays an important role in several metabolic pathways | cytoplasmic | this work Table 1 |
| Ensangp00000000334 ${ }^{\text {b }}$ | Unknown | 39.57/7.29 | SPILLLDDIFDK | ATP/GTP-binding site motif A | intracellular | this work |


| Ensangp00000011593 | Wilm's tumor 1 <br> associating WT1 <br> associated splicing <br> regulator female <br> lethal 2-D homolog | $32.55 / 4.78$ | FTPDSNTGKR | (P-loop) <br> Potential role in transcriptional <br> regulation |
| :--- | :--- | :--- | :--- | :--- |
| Involves in alternative splicing |  |  |  |  |
| regulation |  |  |  |  |$\quad$ this work |  |
| :--- |

${ }^{\text {a) }}$ Subcellular localization is inferred from sequence or structure similarity with orthologous proteins. ${ }^{\text {b) }}$ Identification was performed using
Ensembl database v35 of november 2005. ${ }^{\text {c) }}$ proteins identified from salivary gland extracts of young blood-fed females. Shaded lines: Proteins identified for the first time by a proteomic approach.

Table 3 : List of proteins identified in salivary gland extract of 21day-old blood-fed Anopheles gambiae using iTRAQ

| Ensembl <br> Identification <br> (Ensembl release 43) | Protein <br> Family/Description | Predicted Mr/pI | Ratio 117/114 ${ }^{\text {a }}$ ) | Peptide sequence | Comments | Subcellular Localization b) | Found in other proteomic (P) or transcriptomic (T) studies |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\begin{aligned} & \text { Ensangp000000028522 } \\ & \text { c),d) } \end{aligned}$ | 30 kDa protein | 26.90/3.8 | $1.3 \pm 0.5$ | EQELSDCIVDKR IKECFSSLDK <br> ELDDGLIEREQELSDCIVDK <br> EGEEGAGSDDAVSGADDETEES <br> KDDAEEDSEEGGEEGGDGASGG <br> EGGEKESPR <br> LMNPTIDLVSTIEKYSK <br> ECFSSLDKDVSAMVK | GE rich salivary gland | secreted | P [12], P [9] |
| Ensangp00000022344 | 30 kDa protein | 18.7/3.7 |  | EGEEGAGSDDAVSGADDETEES KDDAEEDSEEGGEEGGDGASGG EGGEKESPR | GE rich salivary gland |  |  |
| Ensangp00000018525 | Aconitate hydratase mitochondrial precursor | $\begin{aligned} & 82.65 / 8.6 \\ & 3 \end{aligned}$ | - | FDQNVYLPYEK ISILGLNNFAPGK | Iron-sulphur proteins that function as electron carriers biosynthesis of amino acid | mitochondrial | this work Table 2 |
| Ensangp00000019171 | Acyl-coA-binding protein | 9.85/9.45 | - | RPSDAELLELYALFK | May act as an intra-cellular carrier of acyl-CoA esters | intracellular | this work |
| Ensangp00000031876 | Acyl-coA-binding protein | 9.65/7.35 | - | NLNATPADADLLEIYGLFJ | « | « | this work |
| Ensangp00000017843 | Alanine amino transferase 2 | $\begin{aligned} & 52.54 / 7.7 \\ & 9 \end{aligned}$ | - | ANIGDCHAMGQPPITFIR | Metabolism of amino acid | cytoplasmic | this work |
| Ensangp00000026558 ${ }^{\text {c }}$ <br> ) | Ambiguous* | $\begin{aligned} & 124.54 / 8 \\ & 43 \end{aligned}$ | - | STTAALLISVLVR | ? | ? | this work |
| Ensangp00000027299 | Ambiguous | $\begin{aligned} & 339.53 / 6 . \\ & 98 \end{aligned}$ | - | EILYDDIERPILQTK <br> LAGVFTPQEPLMNYVISCWVR QIVTFPDEER <br> TAYLYDPQDVQLSVDGIVFR <br> TFDETWATLAVR <br> YPFGAGGEPFR <br> LYFFASK | Subtilase serine protease ? proteasome | cytoplasmic | this work Table 2 |
| Ensangp00000015145 <br> and/or <br> Ensangp00000012963 ${ }^{\text {e }}$ | Annexin | $\begin{aligned} & 35.57 / 4.3 \\ & 1 \\ & \text { and/or } \\ & 27.25 / 4.1 \\ & 1 \end{aligned}$ | - | LLTMIIVGAR | Inhibit PLA2 activity, involved in exocytosis calcium-dependent phospholipid-binding proteins | intracellular | this work |


| Ensangp00000015382 | Apyrase | 61.79/8.6 | $0.71 \pm 0.11$ | AAEEGDTCIAGIAR <br> LNVAQVAGLR <br> GDITNEEAIGASPFSNTVDLLTLR | Anti-platelet | secreted | P [12] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000024604 | ATP synthase subunit alpha mitochondrial precursor | $\begin{aligned} & 59.45 / 9.5 \\ & 2 \end{aligned}$ | - | GAEISAILEER | Catalyzes ATP synthesis | mitochondrial | P [12] |
| Ensangp00000024137 <br> and/or <br> Ensangp00000016868 <br> e) | ATP synthase subunit beta mitochondrial precursor | $\begin{aligned} & \text { 22.69/4.9 } \\ & \text { and/or } \\ & 19.72 / 5.2 \\ & 7 \end{aligned}$ | * | IINVIGEPIDER <br> LVLEVAQHLGENTVR | Catalyzes ATP synthesis | mitochondrial | P [12] |
| Ensangp00000012700 | Calmodulin | $\begin{aligned} & 17.25 / 3.9 \\ & 9 \end{aligned}$ | - | EAFSLFDKDGDGTITTK <br> VFDKDGNGFISAAELR <br> GQNPTEAELQDMINEVDADGNG <br> T <br> TTKELGT <br> IDFPEFLTM <br> ADGNGTIDFP <br> GTITTKELGTV <br> EEVDEMIREAD <br> IDFPEFLTMMAR <br> ADQLTEEQIAEFK <br> DMINEVDADGNGT <br> QVNYEARILHLIK <br> FSLFDKDGDGTITT <br> DADGNGTIDFPEFL <br> AFSLFDKDGDGTITTK | Calcium binding protein | intracellular | T [53] |
| Ensangp00000026391 | Cofilin | $\begin{aligned} & 16.93 / 7.2 \\ & 8 \end{aligned}$ | - | LFLMSWCPDTAK | Binds actin and assists in translocation of actin from the cytoplasm to the nucleus essential for cytokinesis, endocytosis and other cell processes that require rapid turnover of actin filaments | cytoplasmic | This work Tables 1 and 2 T [9] |
| Ensangp00000022538 | Creatine kinase | 26.4/5.18 | * | AVQQQLIDDHFLFK TFLVWCNEEDHLR | Phosphorylation | cytoplasmic | P [12] |
| Ensangp00000020091 | Cytochrome c | $\begin{aligned} & 11.78 / 10 . \\ & 17 \end{aligned}$ | - | GDLIAYLK | Electron tranporter | Mitochondrial membrane | this work |


| Ensangp00000025174 and/or | D7 precursor allergen AED A2 | $\begin{aligned} & 35.57 / 5.7 \\ & \text { and/or } \end{aligned}$ | $0.77 \pm 0.05$ | ALDPEEAWYVYER BVLIGLQLYEEK | ? | secreted | P [12], P [9] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000018280 |  | 32.7/5.1 |  | NYELSGSSQFK |  |  |  |
|  |  |  |  | SADYAFLLR |  |  |  |
|  |  |  |  | SANYGYLAMGK |  |  |  |
|  |  |  |  | SDLEPEVR |  |  |  |
|  |  |  |  | SVLASCTGTQAYDYYSCLLNSP |  |  |  |
|  |  |  |  | VK |  |  |  |
|  |  |  |  | DYELADSAEFR |  |  |  |
|  |  |  |  | IYHGTVDSVAK |  |  |  |
|  |  |  |  | NAFYFHELR |  |  |  |
|  |  |  |  | NAMDCVFR |  |  |  |
| Ensangp00000018340 | D7 related-1 protein | 18.73/9.5 | $0.67 \pm 0.07$ | BLVESTSGEAFK | Anti-inflammatory | secreted | P [12], P [9] |
|  | precursor | 7 |  | KLPALSQYSSVVDK | Scavenger of biogenic amines |  |  |
|  |  |  |  | KVFDTVELVK |  |  |  |
|  |  |  |  | CLVESTSGEAFK |  |  |  |
| Ensangp00000018371 | D7 related-2 protein | 18.46/4.8 | $0.92 \pm 0.08$ | ANTFYTCFLGTSSLAGFK | Anti-inflammatory | secreted | P [12], P [9] |
|  | precursor |  |  | ESVLLELLQR | Scavenger of biogenic amines |  |  |
|  |  |  |  | HMQBVLEVVGFVDGNGEVK |  |  |  |
|  |  |  |  | KANTFYTCFLGTSSLAGFK |  |  |  |
|  |  |  |  | MQTSDPFDMNR |  |  |  |
|  |  |  |  | NAVDYNELLK |  |  |  |
|  |  |  |  | QYTPVSSDDMDK |  |  |  |
| Ensangp00000018330 | D7 related-3 protein | 19.66/4.4 | $0.95 \pm 0.15$ | ANTFYTCFLGTSSAQAFK | Anti-inflammatory | secreted | P [12], P [9] |
| and/or | precursor | 6 |  | AGKLDMGTTFNAGQVSALMK | Scavenger of biogenic amines |  |  |
| Ensangp00000025580 ${ }^{\text {d }}$ |  | and/or |  | LDMGTTFNAGQVSALMK |  |  |  |
| ), e) |  | 18.6/4.5 |  | YAVDYVELLR |  |  |  |
| Ensangp00000018328 ${ }^{\text {d }}$ | D7 related-4 protein | 19.29/7.4 | $0.9 \pm 0.05$ | LYDPLNIIELDK | Anti-inflammatory | secreted | P [12] |
|  | precursor |  |  | CIGECVQVPTSER | Scavenger of biogenic amines |  |  |
| ) |  |  |  | RYEIIEGPEMDK |  |  |  |
|  |  |  |  | YTAEFVQIMK |  |  |  |
|  |  |  |  | VFDLMELK |  |  |  |
| Ensangp00000018321 | D7 related-5 protein | 18.79/5.8 | - | SGSFFSCMLR | ? | secreted | P [12] |
|  | precursor |  |  |  |  |  |  |


| Ensangp00000018385 ${ }^{\text {c }}$ | Disulfide isomerase precursor | $\begin{aligned} & 54.31 / 5.4 \\ & 7 \end{aligned}$ | - | QGETDAVFLFR | Catalyzes the rearrangement of -s-s- bonds in proteins | intracellular | P [12] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000026077 | Disulfide isomerase precursor | $\begin{aligned} & 55.41 / 4.6 \\ & 1 \end{aligned}$ | * | ELETVEAAEEFLK ILEFFGMK ILEFVQSFLDGK | Catalyzes the rearrangement of -s-s- bonds in proteins | endoplasmic reticulum lumen | P [12] |
| Ensangp00000002028 | DNA2 helicase | $\begin{aligned} & 117.99 / 8 . \\ & 45 \end{aligned}$ | - | EKLIIIGDR | ATP binding | ? | this work |
| Ensangp00000014287 ${ }^{\text {d }}$ | Electron transfer flavoprotein subunit alpha mitochondrial pecursor | $\begin{aligned} & 34.14 / 8.6 \\ & 2 \end{aligned}$ | * | FTHIVAGATAFGK | Participates in catalyzing the initial step of the mitochondrial fatty acid beta-oxidation | mitochondrial | this work |
| Ensangp00000018531 | Enolase | 46.62/6.9 | * | AAVPSGASTGVHEALELR EALNLIQDAIAK GNPTVEVDLVTDLGLFR | Glycolytic enzyme | cytoplasmic* | P [12] |
| Ensangp00000010297 | Enzyme | $\begin{aligned} & \text { 79.47/9.6 } \\ & 7 \end{aligned}$ | - | LTSIPTALDLALTGK | Includes enoyl coA hydratase involved in fatty-acid metabolism | mitochondrial | this work |
| Ensangp00000024159 | Fructose biphosphate aldolase | $\begin{aligned} & 39.18 / 7.7 \\ & 2 \end{aligned}$ | * | KPTAQEIALATVTALR IVPIVEPEILPDGDHDLER | Glycolytic enzyme | ? | P [12], P [50] |
| Ensangp00000020828 | Fumarase mitochondrial precursor | $\begin{aligned} & 50.22 / 7.5 \\ & 5 \end{aligned}$ | - | IADAIALAADDVISGK | Amino acid metabolism ? | mitochondrial | this work |
| Ensangp00000017396 | Fumaryl aceto acetase | 45.64/6 | - | GTKQVSLAGGETR | Last enzyme of the tyrosine catabolic pathway | cytoplasmic | this work |
| Ensangp00000029040 | Glutathion S transferase | 19.16/7.5 | * | LYFDMGTLYQR | * | " | this work |
| Ensangp00000010360 | Glyceraldehyde phosphate dehydrogenase | $\begin{aligned} & 35.46 / 8.5 \\ & 5 \end{aligned}$ | - | AGAEYVVESTGVFTTTEK WRDG <br> KLTGM <br> GCLVVN <br> ASVVAI <br> IIPAATG <br> HATTATQKT <br> AFRVPTPNVS <br> LSKPATYDQI <br> GAAKAVGKVIP | Plays an important role in glycolysis and gluconeogenesis | cytoplasmic | P [12], P [50] |


| Ensangp00000003578 | GSG5 | 38.2/6.42 | - | TYFQNEFVEYR | ? | secreted | This work Table 2 T [51] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000019455 | GSG6 | $\begin{aligned} & 13.05 / 5.1 \\ & 5 \end{aligned}$ | $0.65 \pm 0.05$ | EPLPYMYACPGTEPCQSSDR ETREPLPYMYACPGTEPCQSSDR SMHDVLCDRIDQAFLEQ | ? | secreted | P [12], P [9] |
| Ensangp00000021970 | GSG7 | $\begin{aligned} & 16.29 / 8.4 \\ & 6 \end{aligned}$ | - | TLADETAQCMR <br> TLADETAQCLR <br> YGVQNQLR | ? | secreted | P [12] |
| Ensangp00000021028 | putative gVAG protein precursor | 28.9/8.96 | $1.97 \pm 0.6$ | DGQMDVYYFVBNYSFTNIMDR FPYAGQNIAITQFFGYR <br> FVSSWWSEYLDARPEHVR GGPHVGCNPPSSSGGPTCQGK KYPSSYSGKPIGHFTQIASDR MPTLTWDPELASLADANAR VGCSMWYWK | Allergen <br> Belongs to the CAP family: protease inhibitors or proteolytic activity. probably inhibiting host coagulation or complement activity | secreted | P [12], P [9] |
| Ensangp00000014839 | 60 kDa heat shock protein mitochondrial precursor | $\begin{aligned} & 60.77 / 5.2 \\ & 8 \end{aligned}$ | - | VEFQDALVLFSEK | Protein refolding | mitochondrial | this work |
| Ensangp00000003808 | Histone acetyltransferase GCN5 | 85.65/8.9 | - | SIPIESIPGLR | Control of amino acid synthesis | nuclear | this work |
| Ensangp00000009655 | Homolog | $\begin{aligned} & 118.45 / 6 . \\ & 27 \end{aligned}$ | - | DGKELDLVCMQK | C2 domain (cellular proteins involved in signal transduction or membrane trafficking) Cytochrome c heme-binding site (electron-transfer proteins) | ? | this work Table 2 |
| Ensangp00000004315 | Hypothetical 8.8 kDa protein | 8.82/4.05 | * | DKPDIDPVDFLVDVIK | ? | secreted | P [12] |
| Ensangp00000018375 | Hypothetical 10 kDa protein | 10/6.22 | - | LSLQLEEFAVCK <br> AISDLQQGLFDLNHCTK | ? | secreted | this work Table 2 |
| Ensangp00000018379 | Hypothetical 10.2 kDa protein | $\begin{aligned} & 10.13 / 4.5 \\ & 2 \end{aligned}$ | - | LQQMVEDFTACR | ? | secreted | P [12] |
| Ensangp00000013285 | 3 Ketoacyl coA thiolase | $\begin{aligned} & 41.67 / 8.4 \\ & 7 \end{aligned}$ | * | AALDAAGLKPDQVDSVNIGQVL <br> VLSSTDGAFLPR <br> LACAGELGLDINKLNL <br> NGAQDILVGAAH <br> TAGTASGI <br> ASGSRITG | Involved in biosynthetic pathways such as poly betahydroxybutyrate synthesis or steroid biogenesis | intracellular | this work |
| Ensangp00000010689 | Kinase | $\begin{aligned} & 74.81 / 9.3 \\ & 3 \end{aligned}$ | - | SLDLLDSMLVLDP <br> PGSEDLSGEEDIGSPLLPSNRDTI <br> QNLTPSG <br> REIKILRQ | Protein phosphorylation | cytoplasmic | this work |


| Ensangp00000020132 | Low density lipoprotein receptor | $\begin{aligned} & 179.24 / 6 \text {. } \\ & 29 \end{aligned}$ | - | AGINMM <br> GGAGAPAG <br> DGTERVLIVSQNL <br> GSQRVELITK <br> IVTAEIQAPDG <br> SPDDAPADHVCACPQGLMLLK <br> GRTN | Lipid metabolism | membranar | this work |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000022875 | Lysozyme precursor | $\begin{aligned} & 15.33 / 8.5 \\ & 6 \end{aligned}$ | * | NGSTDYGIFQINNK <br> YWBDSGYGSNDCK <br> NLLNDDITDDIK <br> KLPNVSSCF | Immunity related Antibacterial enzyme | secreted | P [12], P [50] |
| Ensangp00000020184 | Malate dehydrogenase | $\begin{aligned} & 35.27 / 9.5 \\ & 2 \end{aligned}$ | - | ANTFVGEAAGVDPQK | Metabolic enzymes which catalyse the last step in anaerobic glycolysis | mitochondrial | P [12] |
| Ensangp00000017682 | Maltase | $\begin{aligned} & 67.21 / 5.8 \\ & 7 \end{aligned}$ | $1.3 \pm 0.2$ | AMPSGAIANWVLGNHDNSR DQPETYDMVHQWR ELNVAAQLAAPR GITQTIDYLK | Sugar digestion Converts sucrose in nectar to glucose and fructose | secreted | P [12] |
| Ensangp00000003748 | Myosin | $121.89 / 10$ | - |  | Contractile protein | cytoplasmic | this work $\mathrm{P}[52]$ |
| Ensangp00000026137 | Nucleolar RNA associated protein | $\begin{aligned} & 117.56 / 7 \text {. } \\ & 05 \end{aligned}$ | - | LSSETIDELEK | Appears to be associated with ribosome biogenesis | cytoplasmic | this work |
| Ensangp00000011253 | Nucleoside diphosphate kinase | $\begin{aligned} & 19.01 / 8.4 \\ & 6 \end{aligned}$ | - | GDLCVQVGR | Maintenance of cellular pool of nucleoside triphosphates | cytoplasmic and plasma membrane | this work Table 1 T [9] |
| Ensangp00000012716 | Putative 5, nucleotidase precursor | $\begin{aligned} & 63.47 / 7.0 \\ & 1 \end{aligned}$ | $0.92 \pm 0.24$ | APFPLTLIHINDLHAR <br> DQIYYVVVPSYLADGKDGFAM <br> K <br> ECIAGIAR <br> GLAPYLAELEK <br> LGTQVIGTTEVFLDRESCR <br> LSGADLWSAIDHSFTLDDEFR <br> MKIPTVVANLEK <br> NVNIIVVLSHCGLDGDK <br> QLAEEAGDLIDVIVGAHSHSLLL NK | Anti-platelet | secreted | P [12] |
| Ensangp00000020778 | Peptidyl prolyl cis trans isomerase | $\begin{aligned} & 18.29 / 8.9 \\ & 7 \end{aligned}$ | - | FFDMTVDNQPLGR IVIELRPDVVPK HVVFGSVVEGMDVVR | Accelerates protein folding | cytopasmic | this work |
| Ensangp00000028058 | Peroxidase precursor | $\begin{aligned} & 24.99 / 8.2 \\ & 3 \end{aligned}$ | $0.95 \pm 0.15$ | AFAGAININDHMFNPTVLER CFAIPVRPDDPVLSAGGIQCLDL VR LLPAEYGDGVYVPR | Vasodilatator | secreted | P [12], P[50] |


|  |  |  |  | SNITPELTILHVAFLR TTLVNMQFGQLVAHDMGLR WEDFVELR |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000020634 | Peroxysomal targeting signal 2 receeptor | $\begin{aligned} & 36.41 / 6.2 \\ & 2 \end{aligned}$ | - | VSGSGDGSVQLWNT <br> FTTNR <br> TNLAS <br> SVQLWNTNLASN <br> SQFYGLAGGGT | Family of potein implicated in a variety of functions ranging from signal transduction and transcription regulation to cell cycle control and apoptosis | ? | this work |
| Ensangp00000024749 | Pheromone/general odorant binding protein OBP56 | $\begin{aligned} & 27.15 / 5.5 \\ & 2 \end{aligned}$ | - | SASEVQDDKCK | ? | ? | this work |
| Ensangp00000013993 | Phosphatidylethanola mine-binding protein | $\begin{aligned} & \text { 24.17/6.6 } \\ & 7 \end{aligned}$ | - | YVFLVYK | Proteinase inhibitor | $?$ | $\begin{aligned} & \text { this work } \\ & \mathrm{P}[52] \end{aligned}$ |
| Ensangp00000020531 | Precursor | 200.9/4.5 | - | ERTGEIMLLQR AGTIVGNVSALDEDVGPNG TRDARLDRDTNPESYAI GTIFVNSTLNYNYAAVI VERQLDYEE VSGVLDRFTVEMQERLANANLE LS | Cadherin | membranar | this work |
| Ensangp00000031578 | Precursor | $\begin{aligned} & 58.96 / 9.6 \\ & 8 \end{aligned}$ | - | DMPNITLLNLDGNQLSR <br> NLLQNLDLALFVAMPQLLNLN ASSPV <br> ANNLT <br> SAPIA <br> PVTGR <br> PNITLLN <br> VSAPIGL <br> NKITTFNIT | Leucine rich repeat <br> Putatively involved in proteinprotein interaction | ? | this work |
| Ensangp00000021077 | Ribonuclease | $\begin{aligned} & 14.41 / 8.0 \\ & 4 \end{aligned}$ | - | ALAPYNQAIVADR | Inhibits protein synthesis by cleavage of mRNA | ? | this work |
| Ensangp00000006850 | DNA directed RNA polymerase | $\begin{aligned} & 68.25 / 8.1 \\ & 8 \end{aligned}$ | - | LSYISALGMMTR | Transcription | nuclear | this work |
| Ensangp00000017327 | Putative salivary protein GSG1b | 46.6/7.37 | - | DYESYLGAMFAADAFHVVYEA D GK | ? | secreted | P [12] |
| Ensangp00000032098 ${ }^{\text {d }}$ | Salivary D3 protein | ? | - | AAAGPAPDPSSQFCQQLLDDAQ R | Saglin | secreted | P [12] |
| Ensangp00000020530 | Serine protease precursor | 25.2/4.57 | - | NGQNDIALLQLDRK VITSAQCTTDEGNGIPSVVRLGG TK | Involved in immunity or in coagulation cascade | secreted | this work |


| Ensangp00000016680 | Serpin 9 | $\begin{aligned} & 46.36 / 7.0 \\ & \end{aligned}$ | - | SVLFAVL | Serine protease inhibitor Involved in immunity | secreted | T [11] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | LIWDSVV |  |  |  |
|  |  |  |  | ALLQLDRKIIIN |  |  |  |
|  |  |  |  | TTDEGNGIPSVVR |  |  |  |
|  |  |  |  | LAAETDILHEVVNEGISR |  |  |  |
| Ensangp00000009988 | SG3 | $\begin{aligned} & 4.3 / 20.01 \\ & 3 \end{aligned}$ | - | ATGPLFLPHFGQGPR | Mucin | secreted | T [51] |
|  |  |  |  | RGQQ |  |  |  |
|  |  |  |  | LIFLAA |  |  |  |
|  |  |  |  | SVERNPA |  |  |  |
|  |  |  |  | ATIAVASAAT |  |  |  |
|  |  |  |  | ASPTTAEA |  |  |  |
|  |  |  |  | QQQRQQVQR |  |  |  |
| Ensangp00000009009 | Fact complex subunit facilitates chromatin trancription | $\begin{aligned} & 71.65 / 6.2 \\ & 8 \end{aligned}$ | - | RPLSAYMLWLNSAR | Recombination signal sequence recognition T160 | nuclear | this work |
| Ensangp00000016164 | Superoxyde dismutase | $\begin{aligned} & 15.67 / 5.4 \\ & 5 \end{aligned}$ | - | SLVVHADPDDLGVGGHELSK | Metalloprotein that prevents damage by oxygen-mediated free radicals | intracellular | this work |
| Ensangp00000021085 | Translationally controlled tumor protein TCTP | $\begin{aligned} & 19.54 / 4.4 \\ & 2 \end{aligned}$ | - | LVDDVMYEVYGK | Histamine-releasing factor | ? | T [9], T [11] |
| Ensangp00000017522 | Trio protein | 43.78/6.4 | - | SMYDLIGQLVQSSK | ? | secreted? | P [12] |
| Ensangp00000025045 | Trypsin precusror | $\begin{aligned} & 28.65 / 6.4 \\ & 9 \end{aligned}$ | - | QIGIVSWGDTQCVGT | Proteolytic enzyme | secreted | this work |
|  |  |  |  | RGGSSTL |  |  |  |
|  |  |  |  | NETDLTVR |  |  |  |
|  |  |  |  | RLALTAGH |  |  |  |
|  |  |  |  | NGNFVPNL |  |  |  |
|  |  |  |  | PAPARATGRIV |  |  |  |
| Ensangp00000008105 | E3 Ubiquitin ligase | $\begin{aligned} & \text { 201.24/6. } \\ & 46 \end{aligned}$ | - | GLAMADLDRLEK QQLCIKP | Involved in protein degradation pathway | cytoplasmic | this work |
|  |  |  |  | NPDN |  |  |  |
|  |  |  |  | SEHRNHK |  |  |  |
|  |  |  |  | GTYHSVN |  |  |  |
|  |  |  |  | TQASQQQQAPL |  |  |  |
|  |  |  |  | LRDGSRVMMMG |  |  |  |
| Ensangp00000012822 | Unknown | 74.9/7.88 | * | DVQASHISRLGTSSIVSYTP | Immunoglobulin-like | ? | this work |
|  |  |  |  | TLRNGTPQASNSI |  |  |  |
|  |  |  |  | YCTLRNGT |  |  |  |
|  |  |  |  | NVSMC |  |  |  |
|  |  |  |  | PDTIDSD |  |  |  |
| Ensangp00000012893 | Unknown | 72.74/4.9 | - | ELEDIVQPIIAK | Hsp70 and tropomyosin | ER ? | this work |


|  |  |  |  |  | domains |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000016832 | Unknown | $\begin{aligned} & 19.42 / 4.8 \\ & 8 \end{aligned}$ | - | QQAAAAAETTSQAAGTLMDHA K | Anti-freeze protein | ? | this work |
| Ensangp00000017135 | Unknown | $\begin{aligned} & 85.43 / 8.6 \\ & 4 \end{aligned}$ | - | IKCGLLLEGVR | ? | ? | this work |
| Ensangp00000019537 | Unknown | $\begin{aligned} & 90.81 / 7.4 \\ & 1 \end{aligned}$ | * | KLMSDYYSSVVASTN EMQSLFLPSS QREH MAHSQ TGSTT | ? | ? | this work |
| Ensangp00000028177 | Unknown | $\begin{aligned} & 36.81 / 10 . \\ & 03 \end{aligned}$ | - | LGIGSSSINGSGAVVRK | Basic helix-loop-helix dimerisation region |  | this work |
| Ensangp00000029447 | Unknown | $\begin{aligned} & 20.35 / 6.2 \\ & 4 \end{aligned}$ | - | EQQQLALDVR | ? | secreted | this work |

${ }^{\text {a) }}$ Ratios indicated in bold correspond to a significant increase or decrease of protein expression in the presence of Plasmodium. ${ }^{\text {b) }}$ Subcellular localization is inferred from sequence or structure similarity with orthologous proteins. ${ }^{\text {c) }}$ Identification was performed using Ensembl database v35 of november 2005. ${ }^{\text {d) }}$ The part of the sequence in bold is that described in Ensembl 43. ${ }^{\text {a }}$ Cases where the same peptides match more than one genomic sequence. * means that the protein was quantified one time. Shaded lines : Proteins newly identified by iTRAQ. References underlined correspond to proteins found in human saliva.

Supplementary Table 1 : List of proteins identified by 1-DE-MS according to their slice number

| Slice number | Ensembl identification | Protein family /description |
| :---: | :---: | :---: |
| 1 | No signal |  |
| 2 | Ensangp00000003518 | CoA carboxylase |
| 3 | Ensangp00000010081 and Ensangp00000012716 | Glycogen phosphorylase and <br> putative $5^{\prime}$ nucleotidase precursor |
| 4 | Ensangp000000012716 and Ensangp000000017682 | putative $5^{\prime}$ nucleotidase precursor and Maltase |
| 5 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 6 | Ensangp00000017682 | Maltase |
| 7 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 8 | Ensangp00000012716 and Ensangp00000015382 | putative $5^{\prime}$ nucleotidase precursor and Apyrase |
| 9 | Ensangp00000016660 | Isocitrate dehydrogenase |
| 10 | NI |  |
| 11-12 | Ensangp00000025174 | D7 precursor allergen AED A2 |
| 13 | Ensangp00000027418 | Salivary gland 1-like 3 |
| 14-15 | Ensangp00000018280/25174 | D7 precursor allergen AED A2 |
| 16 | Ensangp00000018280/25174 and Ensangp000000021028 | D7 precursor allergen AED A2 and <br> Putative gVAG protein precursor |
| 17 | Ensangp00000018280 | D7 precursor allergen AED A2 |
| 18 | NI |  |
| 19 | Ensangp00000018152 | Triosephosphate isomerase |
| 20-22 | Ensangp00000011253 <br> and <br> Ensangp00000021028 <br> and <br> Ensangp00000018328 | Nucleoside diphosphate kinase and <br> putative gVAG protein precursor and <br> D7 related-4 protein precursor |
| 23 | Ensangp00000018328 and <br> Ensangp00000018330 <br> And <br> Ensangp000000018340 | D7 related-4 protein precursor and <br> D7 related-3 protein precursor and <br> D7 related-1 protein precursor |
| 24-25 | Ensangp00000018371 and Ensangp00000018330 | D7 related-2 protein precursor and <br> D7-related-3 protein precursor |
| 26-27 | Ensangp000000018371 <br> and <br> Ensangp00000018330 <br> and <br> Ensangp000000019455 | D7 related-2 protein precursor and <br> D7 related-3 protein precursor and GSG6 |


| 28-29 | Ensangp00000018371 <br> and <br> Ensangp00000018330 | D7 related-2 protein precursor <br> and <br> D7 related-3 protein precursor |
| :--- | :--- | :--- |
| $30-31$ | Ensangp00000018371 <br> and <br> Ensangp00000012492 | D7 related-2 protein precursor <br> and <br> precursor |

NI : non-identified protein

Supplementary Table 2 : List of proteins identified by 2-DE-MS according to their spot number

| Spot number | Ensembl identification | Protein family /description |
| :---: | :---: | :---: |
| 1-5 | NI |  |
| 6-8 | Ensangp00000017682 | Maltase |
| 9 | Ensangp00000019887 | unknown |
| 10 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 11 | NI |  |
| 12 | Ensangp00000017682 | Maltase |
| 13-33 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 34-37 | NI |  |
| 38-51 | Ensangp00000012716 | putative $5^{\prime}$ nucleotidase precursor |
| 52 | Ensangp00000027211 | Disulfide isomerase precursor |
| 53 | NI |  |
| 54 | Ensangp00000012716 | putative 5 ' nucleotidase precursor |
| 55-56 | NI |  |
| 57 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 58-59 | NI |  |
| 60 | Ensangp00000012716 | putative 5 ' nucleotidase precursor |
| 61-63 | NI |  |
| 64-65 | Ensangp00000012716 | putative $5^{\prime}$ nucleotidase precursor |
| 66-67 | Ensangp00000024137 <br> and/or <br> Ensangp000000016868 | ATP synthase subunit beta mitochondrial precursor |
| 68-74 | NI |  |
| 75 | Ensangp00000013568 and | Precursor and |
|  | Ensangp00000009988 | GSG3 |
| 76 | Ensangp00000009988 | GSG3 |
| 77-82 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 83 | NI |  |
| 84 | Ensangp00000028522 | 30 kDa |
| 85-87 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 88-89 | NI |  |
| 90 | Ensangp00000022538 | Creatine kinase |
| 91-94 | NI |  |
| 95 | Ensangp00000011707 | Aspartate amino transferase |
| 96 | Ensangp00000011006 | Malate dehydrogenase |
| 97-108 | NI |  |
| 109 | Ensangp00000012460 | Phosphoglycerate kinase |
| 110 | Ensangp00000018590 | 5 aminolevulinate synthase |
| 111-112 | NI |  |
| 113 | Ensangp00000014287 and | Electron transfer flavoprotein alpha subunit and |
|  | Ensangp00000025174 | D7 precursor allergen AED A2 |
| 114-119 | Ensangp00000025174 | D7 precursor allergen AED A2 |
| 120 | NI |  |
| 121-125 | Ensangp00000025174 | D7 precursor allergen AED A2 |


| 126-129 | NI |  |
| :---: | :---: | :---: |
| 130-134 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 135 | Ensangp00000012702 | Unknown |
| 136-139 | NI |  |
| 140-141 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 142 | Ensangp00000016366 | Glucose dehydrogenase precursor |
| 143 | NI |  |
| 144-145 | Ensangp00000012716 | putative 5' nucleotidase precursor |
| 146 | NI |  |
| 147 | NI |  |
| 148 | Ensangp00000015800 | Phosphoglycerate mutase |
| 149-151 | Ensangp00000025174 | D7 precursor allergen AED A2 |
| 152 | Ensangp00000018152 and | Triose phosphate isomerase and |
|  | Ensangp00000018041 | Toll precursor |
| 153 | NI |  |
| 154 | Ensangp00000025174 | D7 precursor allergen AED A2 |
| 155 | Ensangp00000011661 | Glutathion S transferase |
| 156 | Ensangp00000024808 | Glutathion S transferase |
| 157-168 | NI |  |
| 169 | Ensangp000000026134 and | Ambiguous and |
|  | Ensangp00000025174 | D7 precursor allergen AED A2 cofilin |
| 170 | Ensangp00000026391 |  |
| 171-176 | Ensangp00000018328 | D7 related-4 protein precursor |
| 177-179 | NI |  |
| 180 | Ensangp00000018330 | D7 related-3 protein precursor |
| 181 | Ensangp00000018371 and | D7-related-2 protein precursor and |
|  | Ensangp00000027538 and | Retrovirus related pol polyprotein and |
|  | Ensangp00000018330 | D7 related-3 protein precursor |
| 182-183 | Ensangp00000018371 and | D7 related-2 protein precursor and |
|  | Ensangp00000027538 | Retrovirus related pol polyprotein |
| 184-185 | Ensangp00000028522 | 30 kDa |
| 186 | Ensangp00000015067 and | Mitochondrial carrier and |
|  | Ensangp00000028522 | 30 kDa |
| 187-202 | Ensangp00000028522 | 30 kDa |
| 203-205 | NI |  |

NI : non-identified protein

Supplementary Table 3: Proteins identified in salivary gland extracts of A. gambiae blood-fed females

| Ensembl | Protein | Predicted | Identification | 1DE-MS |  | E-MS | MS/MS | Comments | Subcell |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Identification <br> (Ensembl release 43) | Family/Description | $\mathrm{M}_{\mathrm{r}} / \mathrm{pI}$ |  | $\%$ <br> coverage | spot numb | $\begin{gathered} \% \\ \text { coverage }^{\text {a }} \end{gathered}$ | Peptide sequence |  | Localization b) | other <br> proteomic (P) or <br> transcriptomic (T) <br> studies |
| Ensangp00000028522 | 30 kDa protein | $\begin{aligned} & \hline 26.90 / 3.8 \\ & \mathbf{1 8 . 7 / 3 . 7} \end{aligned}$ | $\begin{aligned} & \hline \text { 2-DE-MS } \\ & \text { LC MS/MS } \end{aligned}$ | - | $\begin{aligned} & 84, \\ & 184- \end{aligned}$ | PSD | EQELSDCIVDKR IKECFSSLDK | GE rich salivary | secreted | P [12], P [9] |
| Ensangp00000022344 <br> d) |  |  | iTRAQ |  | 188, <br> 190, <br> 192,- <br> 193, <br> 196- <br> 199, <br> 201- <br> 202 |  | ELDDGLIEREQELSDCIVDK LMNPTIDLVSTIEKYSK ECFSSLDKDVSAMVK EGEEGAGSDDAVSGADDETE ESKDDAEEDSEEGGEEGGDG ASGGEGGEKESPR | gland |  |  |
| Ensangp00000018525e | Aconitate hydratase mitochondrial precursor | $\begin{aligned} & 82.65 / 8.6 \\ & 3 \end{aligned}$ | LC MS/MS iTRAQ | - | - | - | FDQNVYLPYEK ISILGLNNFAPGK | Iron-sulphur proteins that function as electron carriers biosynthesis of amino acid | mitochondrial | this work |
| $\underset{)}{\text { Ensangp000000019171 }}$ | Acyl-coA -binding protein | 9.85/9.45 | iTRAQ | - | - | - | RPSDAELLELYALFK | May act as an intra-cellular carrier of acyl-CoA esters | intracellular | this work |
| Ensangp00000031876 | Acyl-coA -binding protein | 9.65/7.35 | iTRAQ | - | - | - | NLNATPADADLLEIYGLFJ | « | « | this work |
| Ensangp00000017843 | Alanine aminotransferase 2 | $\begin{aligned} & 52.54 / 7.7 \\ & 9 \end{aligned}$ | iTRAQ | - | - | - | ANIGDCHAMGQPPITFIR | Metabolism of amino acid | cytoplasmic | this work |
| Ensangp00000016546 | Ambiguous | $\begin{aligned} & 25.56 / 9.9 \\ & 4 \end{aligned}$ | LC MS/MS | - | - | - | KGIGTHLMITLEVLAR | GCN5- <br> related N acetyltransfer ase | ? | this work |
| $\text { Ensangp000000022917 }{ }^{\text {d }}$ | Ambiguous | $\begin{aligned} & 72.38 / 10 . \\ & 16 \end{aligned}$ | LC MS/MS | - | - | - | GRPILPLLKTVQSYK | Tropomyosin domain | intracellular | this work |
| $\text { Ensangp00000024702 }{ }^{\mathrm{d}}$ | Ambiguous | $\begin{aligned} & 30.31 / 9.5 \\ & 8 \end{aligned}$ | LC MS/MS | - | - | - | IHDGVTHAAK | ? | ? | this work |
| Ensangp00000026066 | Ambiguous | $\begin{aligned} & 25.13 / 7.0 \\ & 6 \end{aligned}$ | LC MS/MS | - | - | - | MSDKVVSSFLR | ? | ? | this work |


|  | Ambiguous | 23.01/10 | 2-DE-MS | - | 169 | PSD | - | ? | ? | this work |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000026558 ${ }^{\text {c }}$ | Ambiguous | $\begin{aligned} & 124.54 / 8 . \\ & 43 \end{aligned}$ | LC MS/MS | - | - | - | STTAALLISVLVR | ? | ? | this work |
| Ensangp00000027299 ${ }^{\text {e }}$ | Ambiguous | $\begin{aligned} & 339.53 / 6 \text {. } \\ & 98 \end{aligned}$ | LC MS/MS, ITRAQ | - | - | - | EILYDDIERPILQTK <br> LAGVFTPQEPLMNYVISCWVR QIVTFPDEER <br> TAYLYDPQDVQLSVDGIVFR <br> TFDETWATLAVR <br> YPFGAGGEPFR <br> LYFFASK | Subtilase serine protease | cytoplasmic | this work |
| ${ }_{\text {) }}$ Ensangp00000018590 ${ }^{\text {d }}$ | 5 Aminolevulinate synthase erythroid specific mitochondrial precursor | $\begin{aligned} & 46.31 / 7.5 \\ & 4 \end{aligned}$ | 2-DE-MS | - | 110 | 25 | - | Metabolism of amino acid | mitochondrial matrix | this work |
| Ensangp00000015145 <br> and/or <br> $\underset{\text { e, }, \mathrm{f})}{\text { Ensangp00000012963 }}$ | Annexin | $\begin{aligned} & 35.57 / 4.3 \\ & 1 \\ & \text { and/or } \\ & 27.25 / 4.1 \\ & 1 \end{aligned}$ | iTRAQ | - | - | - | LLTMIIVGAR | Inhibit PLA2 activity, involved in exocytosis calciumdependent phospholipid -binding proteins | intracellular | this work |
| ${ }^{\text {Ensangp00000029258 }}$ | Apolipoprotein D precursor | $\begin{aligned} & 26.11 / 4.5 \\ & 5 \end{aligned}$ | LC MS/MS | - | - | - | QSDVGRAVVAFPDESPLEAK | Extracellular <br> ligandbinding proteins displaying high specificity for small hydrophobic molecules | secreted | this work |
| $\underset{\text { ),e) }}{\text { Ensangp00000015382 }}$ | Apyrase | 61.79/8.6 | 1-DE-MS LC MS/MS iTRAQ | 20\% | - | - | AAEEGDTCIAGIAR <br> LNVAQVAGLR <br> GDITNEEAIGASPFSNTVDLLT <br> LR | Anti-platelet | secreted | P [12] |
| $\text { Ensangp00000011707 }{ }^{\text {d }}$ | Aspartate aminotransferase | $\begin{aligned} & 44.71 / 6.7 \\ & 8 \end{aligned}$ | 2-DE-MS | - | 95 | 17 | - | Metabolism of amino acid | cytoplasmic | this work |
| Ensangp00000024604 | ATP synthase subunit alpha | $\begin{aligned} & 59.45 / 9.5 \\ & 2 \end{aligned}$ | iTRAQ | - | - | - | GAEISAILEER | $\begin{aligned} & \text { Catalyzes } \\ & \text { ATP } \end{aligned}$ | mitochondrial | P [12] |


| Ensangp00000024137 <br> and/or <br> $\underset{\text { d,e,f) }}{\text { Ensangp000000 }} 0$ | mitochondrial precursor ATP synthase subunit beta mitochondrial precursor | $\begin{aligned} & 22.69 / 4.9 \\ & \text { and/or } \\ & 19.72 / 5.2 \\ & 7 \end{aligned}$ | LC MS/MS iTRAQ | - | 66-67 | (31-37) | IINVIGEPIDER <br> LVLEVAQHLGENTVR | synthesis <br> Catalyzes <br> ATP <br> synthesis | mitochondrial | P [12] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000012700 | Calmodulin | $\begin{aligned} & 17.25 / 3.9 \\ & 9 \end{aligned}$ | iTRAQ | - | - | - | EAFSLFDKDGDGTITTK VFDKDGNGFISAAELR GQNPTEAELQDMINEVDADG NGT <br> TTKELGT <br> IDFPEFLTM <br> ADGNGTIDFP <br> GTITTKELGTV <br> EEVDEMIREAD <br> IDFPEFLTMMAR <br> ADQLTEEQIAEFK <br> DMINEVDADGNGT <br> QVNYEARILHLIK <br> FSLFDKDGDGTITT <br> DADGNGTIDFPEFL <br> AFSLFDKDGDGTITTK | Calcium binding protein | intracellular | this work |
| Ensangp00000018543 | Chromosome associated polypeptide C XCAP C homolog | $\begin{aligned} & 156.83 / 5 . \\ & 34 \end{aligned}$ | LC MS/MS | - | - | - | LQTELIELKR | Structural maintenance of chromosome ABC transporter related domain | nuclear | this work |
| $\text { Ensangp000000003518 }{ }^{\text {d }}$ | CoA carboxylase mitochondrial precursor | $\begin{aligned} & 130.5 / 6.6 \\ & 7 \end{aligned}$ | 1-DE-MS | 15\% | - | - | - | Key enzyme in the catabolic pathway of odd-chain fatty acids : isoleucine, threonine, methionine and valine | mitochondrial matrix | this work |
| Ensangp00000026391 | Cofilin | $\begin{aligned} & 16.93 / 7.2 \\ & 8 \end{aligned}$ | 2-DE-MS <br> LC MS/MS | - | 170 | 42 | LFLMSWCPDTAK | Binds actin and assists in | cytoplasmic | T [9] |





|  | aldolase |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000020828 | Fumarase mitochondrial precursor | $\begin{aligned} & 50.22 / 7.5 \\ & 5 \end{aligned}$ | iTRAQ | - | - | - | IADAIALAADDVISGK | Generation of precursor metabolites and energy | mitochondrial | this work |
| Ensangp000000017396 | Fumaryl aceto acetase | 45.64/6 | iTRAQ | - | - | - | GTKQVSLAGGETR | Last enzyme of the tyrosine catabolic pathway | cytoplasmic | this work |
| Ensangp00000029040 | Glutathion S transferase | 19.16/7.5 | iTRAQ | - | - | - | LYFDMGTLYQR | Key role in cellular detoxificatio n | cytoplasmic and nuclear | this work |
| ${ }_{\text {E }}^{\text {Ensangp00000011661 }}$ | Glutathion S transferase (class theta) | $\begin{aligned} & 23.78 / 6.5 \\ & 1 \end{aligned}$ | 2-DE-MS | - | 155 | 33 | - |  |  | this work $\mathrm{P}[50]$ |
| Ensangp00000010360 ${ }^{\text {e }}$ | Glyceraldehyde phosphate dehydrogenase | $\begin{aligned} & 35.46 / 8.5 \\ & 5 \end{aligned}$ | iTRAQ | - | - | - | AGAEYVVESTGVFTTTEK WRDG KLTGM GCLVVN <br> ASVVAI <br> IIPAATG <br> HATTATQKT <br> AFRVPTPNVS <br> LSKPATYDQI <br> GAAKAVGKVIP | Plays an important role in glycolysis and gluconeogen esis | cytoplasmic | $\mathrm{P}[12], \mathrm{P}[50]$ |
| Ensangp000000024265 | Glycin cleavage system H protein mitochondrial precursor | 13.52/4.2 | iTRAQ | - | - | - | LMSEEQYTEFLK | Catalyses the catabolism of glycine in eukaryotes | mitochondrial | this work |
| $\text { Ensangp00000010081 }{ }^{\text {d }}$ | Glycogen phosphorylase | 96.4/6.33 | 1-DE-MS | 18\% | - | - | - | Carbohydrate metabolism | cytoplasmic | this work |


| Ensangp0000000998 ${ }^{\text {e }}$ | GSG3 | $\begin{aligned} & 20.01 / 4.3 \\ & 4 \end{aligned}$ | 2-DE-MS | - | 75,76 | PSD | - | ? | secreted | T [51] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000003578e | GSG5 precursor | 38.2/6.42 | LC MS/MS iTRAQ | - | - | - | TYFQNEFVEYR | ? | secreted | T [51] |
| $\underset{\text { ),e) }}{\text { Ensangp000000019455 }}$ | GSG6 | $\begin{aligned} & 13.05 / 5.1 \\ & 5 \end{aligned}$ | 1-DE-MS <br> LC MS/MS <br> iTRAQ | 36\% | - | - | EPLPYMYACPGTEPCQSSDR <br> ETREPLPYMYACPGTEPCQSS <br> DR <br> SMHDVLCDRIDQAFLEQ | ? | secreted | P [12], P [9] |
| $\underset{\text { ),e) }}{\text { Ensangp000000021970 }}$ | GSG7 | $\begin{aligned} & 16.29 / 8.4 \\ & 6 \end{aligned}$ | LC MS/MS iTRAQ | - | - | - | TLADETAQCMR TLADETAQCLR YGVQNQLR | ? | secreted | P [12] |
| $\text { Ensangp000000005326 }{ }^{\text {d }}$ | Guanine nucleotide releasing factor | $\begin{aligned} & 137.53 / 9 . \\ & 17 \end{aligned}$ | LC MS/MS | - | - | - | LIEKALIYK | May play a role in intracellular signaling cascade | membraneassociated | this work |
| $\underset{* \mathrm{~d}), \mathrm{e})}{\text { Ensangp00000021028 }}$ | putative gVAG protein precursor | 28.9/8.96 | 1-DE-MS <br> LC MS/MS <br> iTRAQ | 43\% | - |  | DGQMDVYYFVBNYSFTNIMD R <br> FPYAGQNIAITQFFGYR <br> FVSSWWSEYLDARPEHVR <br> GGPHVGCNPPSSSGGPTCQGK <br> KYPSSYSGKPIGHFTQIASDR <br> MPTLTWDPELASLADANAR <br> VGCSMWYWK | Allergen. Belongs to the CAP family | secreted | P [12], P [9] |
| Ensangp00000014839 | 60 kDa heat shock protein mitochondrial precursor | $\begin{aligned} & 60.77 / 5.2 \\ & 8 \end{aligned}$ | iTRAQ | - | - | - | VEFQDALVLFSEK | Protein refolding | mitochondrial | this work |
| Ensangp00000003808 | Histone acetyltransferase GCN5 | 85.65/8.9 | iTRAQ | - | - | - | SIPIESIPGLR | Control of amino acid synthesis | nuclear* | this work |
| Ensangp00000009655 ${ }^{\text {e }}$ | Homolog | $\begin{aligned} & 118.45 / 6 . \\ & 27 \end{aligned}$ | iTRAQ | - | - | - | DGKELDLVCMQK | C2 domain (cellular proteins involved in signal transduction or membrane trafficking) Cytochrome c heme- | ? | this work |


| $\underset{y}{\text { Ensangp00000017720 }}$ | 3 <br> Hydroxyisobutyrat e dehydrogenase mitochondrial | $\begin{aligned} & 34.31 / 9.2 \\ & 7 \end{aligned}$ | LC MS/MS | - | - | - | VFADIVNASTGR | binding site (electrontransfer proteins) Involved in amino acid catabolism pathway | mitochondrial | this work |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000018375e | Hypothetical 10 kDa protein | 10/6.22 | $\begin{aligned} & \text { LC MS/MS } \\ & \text { iTRAQ } \end{aligned}$ | - | - | - | LSLQLEEFAVCK <br> AISDLQQGLFDLNHCTK | ? | secreted | this work |
| Ensangp00000018379e | Hypothetical 10.2 kDa protein | $\begin{aligned} & 10.13 / 4.5 \\ & 2 \end{aligned}$ | LC MS/MS iTRAQ | - | - | - | LQQMVEDFTACR | ? | secreted | P [12] |
| Ensangp00000004315e | Hypothetical 8.8 kDa protein | 8.82/4.05 | LC MS/MS iTRAQ | - | - | - | DKPDIDPVDFLVDVIK | ? | secreted | P [12] |
| $\text { Ensangp00000016660 }{ }^{\text {d }}$ | Isocitrate dehydrogenase | $\begin{aligned} & 46.96 / 7.5 \\ & 9 \end{aligned}$ | 1-DE-MS | $32 \%$ | - | - | - | Plays a key role in cellular defense against oxidative stressinduced damage | mitochondrial | this work |
| Ensangp00000013285 ${ }^{\text {e }}$ | 3 Ketoacyl coA thiolase | $\begin{aligned} & 41.67 / 8.4 \\ & 7 \end{aligned}$ | iTRAQ | - | - | - | AALDAAGLKPDQVDSVNIGQ <br> VLVLSSTDGAFLPR <br> LACAGELGLDINKLNL <br> NGAQDILVGAAH <br> TAGTASGI <br> ASGSRITG | Involved in biosynthetic pathways such as poly betahydroxybutyr ate synthesis or steroid biogenesis |  | this work |
| ${ }_{\text {, }}$ Ensangp000000010689 ${ }^{\text {e }}$ | cell division Kinase | $\begin{aligned} & 74.82 / 9.3 \\ & 3 \end{aligned}$ | iTRAQ | - | - | - | SLDLLDSMLVLDP <br> PGSEDLSGEEDIGSPLLPSNRD <br> TIQNLTPSG <br> REIKILRQ <br> AGINMM <br> GGAGAPAG | Protein phosphorylati on | cytoplasmic | this work |
| Ensangp00000020132 | Low density lipoprotein receptor | $\begin{aligned} & 179.24 / 6 . \\ & 29 \end{aligned}$ | iTRAQ | - | - | - | DGTERVLIVSQNL <br> GSQRVELITK <br> IVTAEIQAPDG <br> SPDDAPADHVCACPQGLMLL <br> K | Lipid metabolism | membranar | this work |


| Ensangp00000020384 ${ }^{\text {e }}$ | Low density lipoprotein receptor | 17.3/5.04 | LC MS/MS | - | - | - | GRTN <br> BISRAGICDGK | Lipid metabolism | membranar | this work P [50] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000022875 | Lysozyme precursor | $\begin{aligned} & 15.33 / 8.5 \\ & 6 \end{aligned}$ | LC MS/MS iTRAQ | - | - | - | NGSTDYGIFQINNK YWBDSGYGSNDCK NLLNDDITDDIK KLPNVSSCF | Immunity related Antibacterial enzyme | secreted | P [12], P [50] |
| ${ }_{5}^{\text {Ensangp } 00000011006}{ }^{\text {e }}$ | Malate dehydrogenase | $\begin{aligned} & 35.37 / 6.9 \\ & 5 \end{aligned}$ | 2-DE-MS |  | 96 | PSD | DDLFNTNASIVR | Participates in the citric acid cycle | cytoplasmic | this work |
| $\underset{,, \mathrm{e})}{\text { Ensangp00000020184 }}$ | Malate dehydrogenase | $\begin{aligned} & 35.27 / 9.5 \\ & 2 \end{aligned}$ | LC MS/MS | ${ }^{-}$ | ${ }^{-}$ | ${ }^{-}$ | ANTFVGEAAGVDPQK | Metabolic enzymes which catalyse the last step in anaerobic glycolysis | mitochondrial | P [12] |
|  | Maltase | $\begin{aligned} & 67.21 / 5.8 \\ & 7 \end{aligned}$ | 1-DE-MS <br> 2-DE-MS <br> LC MS/MS <br> iTRAQ | 27\% | 6-8, 12 | $(17-43)$ | AMPSGAIANWVLGNHDNSR DQPETYDMVHQWR ELNVAAQLAAPR GITQTIDYLK | Sugar digestion Converts sucrose in nectar to glucose and fructose | secreted | P [12], T [9] |
| Ensangp00000015067 | Mitochondrial carrier |  | 2-DE-MS | 16 |  | 186 |  |  |  |  |
| ${ }_{\text {, }}^{\text {Ensangp }} 00000004215^{\text {e }}$ | Mitogen activated kinase kinase kinase kinase | $\begin{aligned} & 159.64 / 10 \\ & .04 \end{aligned}$ | LC MS/MS | - | - | - | NIATYYGAFIK | Protein kinase ATP binding | cytoplasmic | this work |
| ${ }_{\text {) }}$ Ensangp00000003748 ${ }^{\text {e }}$ | Myosin | $\begin{aligned} & 121.89 / 10 \\ & .23 \end{aligned}$ | iTRAQ | - | - | - |  | Contractile protein | cytoplasmic | $\begin{aligned} & \text { this work } \\ & \mathrm{P}[52] \end{aligned}$ |
| , Ensangp00000003978 ${ }^{\text {e }}$ | N acylneuraminate cytidyltransferase | 21.1/5.67 | LC MS/MS | - | - | - | HLTLARILLGME | Forms CMP- <br> NeuAc, the nucleotide sugar donor used by sialyltransfer ases (modification may be important in pathogenesis) | cytoplasmic | this work |


| Ensangp00000021120e | NADPH dependent carbonyl reductase | $\begin{aligned} & 27.05 / 7.7 \\ & 3 \end{aligned}$ | LC MS/MS | - | - | - | MDFTGKVVLITGASSGIGAST AK | Carbohydrate metabolism | cytoplasmic | this work |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp0000002613 ${ }^{\text {e) }}$ | Nucleolar RNA associated protein | $\begin{aligned} & 117.56 / 7 . \\ & 05 \end{aligned}$ | iTRAQ | - | - | - | LSSETIDELEK | Appears to be associated with | cytoplasmic | this work |
| $\underset{\mathrm{l}, \mathrm{e})}{\text { Ensangp00000011253 }}$ | Nucleoside diphosphate kinase | $\begin{aligned} & 19.01 / 8.4 \\ & 6 \end{aligned}$ | $\begin{aligned} & \text { 1-DE-MS } \\ & \text { iTRAQ } \end{aligned}$ | 32\% | - | - | GDLCVQVGR | Maintenance of cellular pool of nucleoside triphosphates | cytoplasmic and plasma membrane | this work T [9] |
| $\underset{*, \mathbf{d}), \mathbf{e})}{\text { Ensangp00000012716 }}$ | Putative 5' nucleotidase precursor | $\begin{aligned} & 63.47 / 7.0 \\ & 1 \end{aligned}$ | 1-DE-MS <br> 2-DE-MS <br> LC MS/MS <br> iTRAQ | 20\% | $\begin{aligned} & 10,13- \\ & 33,38- \\ & 51,54, \\ & 57,60, \\ & 64,65 \\ & 77-82, \\ & 85-87 \\ & 130- \\ & 134, \\ & 140- \\ & 141, \\ & 144- \\ & 145 \end{aligned}$ | $\begin{aligned} & (15-30) \\ & \text { PSD } \end{aligned}$ | APFPLTLIHINDLHAR <br> DQIYYVVVPSYLADGKDGFA <br> MK <br> ECIAGIAR <br> GLAPYLAELEK <br> LGTQVIGTTEVFLDRESCR <br> LSGADLWSAIDHSFTLDDEFR <br> MKIPTVVANLEK <br> NVNIIVVLSHCGLDGDK <br> QLAEEAGDLIDVIVGAHSHSLL <br> LNK <br> YDTIEGDYPLVVKK <br> VVIENHTNGTCSWDLDSQR <br> NPIEKGDITNGLAIEAAPYGSS <br> VDMIK | Anti-platelet | secreted | P [12] |
| Ensangp00000020778 | Peptidyl prolyl cis trans isomerase | $\begin{aligned} & 18.29 / 8.9 \\ & 7 \end{aligned}$ | LC MS/MS iTRAQ | - | - | - | FFDMTVDNQPLGR IVIELRPDVVPK HVVFGSVVEGMDVVR | Accelerates protein folding | cytopasmic | this work |
| $\underset{\mathrm{l}, \mathrm{e})}{\text { Ensangp00000028058 }}$ | Peroxidase precursor | $\begin{aligned} & 24.99 / 8.2 \\ & 3 \end{aligned}$ | 1-DE-MS <br> LC MS/MS <br> iTRAQ | 16\% | - | - | AFAGAININDHMFNPTVLER CFAIPVRPDDPVLSAGGIQCLD LVR <br> LLPAEYGDGVYVPR SNITPELTILHVAFLR TTLVNMQFGQLVAHDMGLR WEDFVELR | Vasodilatator | secreted | P [12], P [50] |
| Ensangp00000020634 | Peroxysomal targeting signal 2 receptor | $\begin{aligned} & 36.41 / 6.2 \\ & 2 \end{aligned}$ | iTRAQ | - | - | - | VSGSGDGSVQLWNT <br> FTTNR <br> TNLAS <br> SVQLWNTNLASN | Family of potein implicated in a variety of | ? | this work |


|  |  |  |  |  |  |  | SQFYGLAGGGT | functions ranging from signal transduction and transcription regulation to cell cycle control and apoptosis |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000024749 ${ }^{\text {e }}$ | Pheromone/general odorant binding protein OBP56 | $\begin{aligned} & 27.15 / 5.5 \\ & 2 \end{aligned}$ | iTRAQ | - | - | - | SASEVQDDKCK | ? | ? | this work |
| $\underset{\text { E }}{\text { Ensangp00000013993e }}$ | Phosphatidylethano lamine-binding protein | $\begin{aligned} & 24.17 / 6.6 \\ & 7 \end{aligned}$ | iTRAQ | - | - | - | YVFLVYK | Proteinase inhibitor | ? | this work P[52] |
| Ensangp00000012460 | Phosphoglycerate kinase | $\begin{aligned} & 43.84 / 7.5 \\ & 4 \end{aligned}$ | 2-DE-MS | - | 109 | 27 | - | Glycolysis | cytoplasmic | this work |
| $\underset{\text { ),e) }}{\text { Ensangp00000015800 }}$ | Phosphoglycerate mutase | 28.7/6.8 | LC MS/MS | - | 148 | 25 | YGEEQVLIWR | Involved in energy pathways | cytoplasmic | this work |
| Ensangp000000020531 | Precursor | 200.9/4.5 | iTRAQ | - | - | - | ERTGEIMLLQR <br> AGTIVGNVSALDEDVGPNG <br> TRDARLDRDTNPESYAI <br> GTIFVNSTLNYNYAAVI <br> VERQLDYEE <br> VSGVLDRFTVEMQERLANAN <br> LELS | Cadherin | membranar | this work |
| Ensangp000000012492 | Precursor | $\begin{aligned} & 28.47 / 5.0 \\ & 4 \end{aligned}$ | 1-DE-MS | 22\% | - | - | - | EGF-like domain | ? | this work |
| $\text { Ensangp000000013568 }{ }^{\text {d }}$ | Precursor | 41.83/5.4 | 2-DE-MS | - | 75 | - | - | Aspartic protease A1 | secreted | this work |
| Ensangp000000016366 | Precursor | 45.95/9.4 | 2-DE-MS | - | 142 | 23 | - | Glucose-methanolcholine oxidoreducta se Involved in energy pathways | cytoplasmic | this work |
| Ensangp00000019046 | Precursor | $\begin{aligned} & 12.39 / 8.7 \\ & 5 \end{aligned}$ | LC MS/MS | - | - | - | ANDRAMVK | EGF-like domain | ? | this work |
| Ensangp00000029324 ${ }^{\text {e }}$ | Precursor | 25.94/4.8 | LC MS/MS | - | - | - | TLTFVLKPTK | Alpha 2 | intracellular | this work |


| Ensangp00000031578 ${ }^{\text {e }}$ | Precursor | 58.96/9.6 | iTRAQ | - | - | - | DMPNITLLNLDGNQLSR | macroglobuli n domain Leucine rich | ? | this work |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | 8 |  |  |  |  | NLLQNLDLALFVAMPQLLNLN ASSPV <br> ANNLT <br> SAPIA <br> PVTGR <br> PNITLLN <br> VSAPIGL <br> NKITTFNIT | repeat Putatively involved in proteinprotein interaction |  |  |
| Ensangp000000020734 | Pterin 4 alpha carbinol amine dehydratase | $\begin{aligned} & 21.20 / 10 . \\ & 23 \end{aligned}$ | LC MS/MS | - | - | - | LAQFLDQAAAVAK | Transcription al activator/pter in dehydratase | ? | this work |
| Ensangp000000027538c | Retrovirus related pol polyprotein | $\begin{aligned} & 9.51 / 11.2 \\ & 8 \end{aligned}$ | 2-DE-MS | - | $\begin{aligned} & 181, \\ & 183 \end{aligned}$ | PSD | - | ? | nuclear | this work |
| $\underset{\mathrm{l}, \mathrm{e})}{\text { Ensangp00000021077 }}$ | Ribonuclease | $\begin{aligned} & 14.41 / 8.0 \\ & 4 \end{aligned}$ | LC MS/MS iTRAQ |  |  |  | ALAPYNQAIVADR | Inhibits protein synthesis by cleavage of mRNA | ? | this work |
| Ensangp000000006850 | DNA directed RNA polymerase | $\begin{aligned} & 68.25 / 8.1 \\ & 8 \end{aligned}$ | iTRAQ | - | - | - | LSYISALGMMTR | Transcription | nuclear | this work |
| Ensangp00000019607 ${ }^{\text {e }}$ | Ryanodine receptor 1 | $\begin{aligned} & 577.53 / 5 \text {. } \\ & 18 \end{aligned}$ | LC MS/MS | - | - | - | YFDMFLKLK | $\mathrm{Ca} 2+$ release channels involved in secretory pathways? | membranar | this work |
| Ensangp00000020530 ${ }^{\text {e }}$ | Serine protease precursor | 25.2/4.57 | iTRAQ |  |  |  | NGQNDIALLQLDRK <br> VITSAQCTTDEGNGIPSVVRLG <br> GTK <br> SVLFAVL <br> LIWDSVV <br> ALLQLDRKIIIN <br> TTDEGNGIPSVVR | Involved in immunity or in coagulation cascade | secreted | this work |
| Ensangp00000016680 | Serpin 9 | 46.36/7 | iTRAQ | - | - | - | LAAETDILHEVVNEGISR | Serine protease inhibitor Involved in immunity | secreted | T [11] |


| Ensangp00000017327 | putative Salivary protein SG1B | 46.6/7.37 | iTRAQ | - | - | - | DYESYLGAMFAADAFHVVYE AD <br> GK | ? | secreted | P [12] |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\underset{\text { ),f) }}{\text { Ensangp00000032098é }}$ | Salivary D3 protein |  | iTRAQ | - | - | - | AAAGPAPDPSSQFCQQLLDDA QR | Saglin | ? | P [12] |
| $\underset{)_{*}}{\text { Ensangp00000027418 }{ }^{\mathrm{d}}}$ | Salivary gland 1like 3 protein | $\begin{aligned} & 44.51 / 6.0 \\ & 4 \end{aligned}$ | 1-DE-MS | 30\% | - | - | QR | ? | secreted | P [12] |
| Ensangp00000009988 ${ }^{\text {e }}$ | SG3 | 20.01/4.3 | iTRAQ |  |  |  | ATGPLFLPHFGQGPR RGQQ <br> LIFLAA <br> SVERNPA <br> ATIAVASAAT <br> ASPTTAEA <br> QQQRQQVQR | Mucin | secreted | T [51] |
| $\text { Ensangp00000008103 }{ }^{\text {e }}$ | Stromal interaction molecule precursor | $\begin{aligned} & 54.49 / 6.3 \\ & 6 \end{aligned}$ | LC MS/MS | - | - | - | DVEGLLKAEVALK | Role in RNA binding | membranar | this work |
| Ensangp00000009009 ${ }^{\text {e }}$ <br> ) | Fact complex subunit facilitates chromatin transcription | $\begin{aligned} & 71.65 / 6.2 \\ & 8 \end{aligned}$ | iTRAQ | - | - | - | RPLSAYMLWLNSAR | Recombinati <br> on signal <br> sequence <br> recognition <br> T160 | nuclear | this work |
| Ensangp00000016164 | Superoxyde dismutase | $\begin{aligned} & 15.67 / 5.4 \\ & 5 \end{aligned}$ | iTRAQ | - | - | - | SLVVHADPDDLGVGGHELSK | Metalloprotei n that prevents damage by oxygenmediated free radicals | intracellular | this work |
| ${ }_{\text {E }}^{\text {Ensangp000000018041 }}$ | Toll precursor | $\begin{aligned} & 16.69 / 4.5 \\ & 1 \end{aligned}$ | 2-DE-MS | - | 152 | 17 | - | Toll IA Involved in signal transduction pathways in response to pathogens | plasma membrane | this work $\mathrm{P}[50]$ |



|  |  |  |  |  |  |  |  | that bind a multitude of functionally diverse signaling proteins |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ensangp00000012822e | Unknown | 74.9/7.88 | iTRAQ |  |  |  | DVQASHISRLGTSSIVSYTP <br> TLRNGTPQASNSI <br> YCTLRNGT <br> NVSMC <br> PDTIDSD | Immunoglob ulin-like domain Involved in cell adhesion | membrane | this work |
| $\underset{\text { 権 }}{\text { Ensangp } 00000015472^{\text {c }}}$ | Unknown | $\begin{aligned} & 15.64 / 10 . \\ & 38 \end{aligned}$ | 1-DE-MS | 20\% |  | - | - | InterPro Zn -finger, C2H2 type nucleic acidbinding protein | nuclear? | this work $\mathrm{P}[50], \mathrm{T}[9]$ |
| Ensangp00000016832 ${ }^{\text {e }}$ | Unknown | $\begin{aligned} & 19.42 / 4.8 \\ & 8 \end{aligned}$ | iTRAQ |  |  |  | QQAAAAAETTSQAAGTLMDH AK | Anti-freeze protein | ? | this work |
| , Ensangp00000017135 ${ }^{\text {e }}$ | Unknown | $\begin{aligned} & 85.43 / 8.6 \\ & 4 \end{aligned}$ | LC MS/MS iTRAQ | - | - | - | IKCGLLLEGVR | ? | ? | this work |
| Ensangp00000019537 ${ }^{\text {e }}$ | Unknown | $90.81 / 7.4$ $1$ | iTRAQ | - | - | - |  | $?$ | $?$ | this work |
| $\underset{,}{\text { Ensangp00000019887 }}$ | Unknown | 70.9/5.1 | 2-DE-MS | - | 9 | 18 |  | Heat-shock 70 domain May be involved in response to stress | cytoplasmic and organelles | P [12] |
| Ensangp00000028177 ${ }^{\text {e }}$ | Unknown | $\begin{aligned} & 36.81 / 10 \text {. } \\ & 03 \end{aligned}$ | iTRAQ | - | - | - | LGIGSSSINGSGAVVRK | Basic helix-loop-helix dimerisation region |  | this work |
| ${ }_{3}{ }^{\text {Ensangp00000028294 }}$ | Unknown | $\begin{aligned} & 15.18 / 4.5 \\ & 7 \end{aligned}$ | LC MS/MS | - | - | - | GSTINLTBAVK | Immunoglob ulin-like domain Involved in cell adhesion | membrane ? | this work |
| , Ensangp00000029447 ${ }^{\text {e }}$ | Unknown | $\begin{aligned} & 20.35 / 6.2 \\ & 4 \end{aligned}$ | iTRAQ | - | - | - | EQQQLALDVR | ? | secreted | this work |
| Ensangp0000012893 ${ }^{\text {e }}$ | Unknown | $\begin{aligned} & 72.74 / 4.9 \\ & 2 \end{aligned}$ | iTRAQ | - | - | - | ELEDIVQPIIAK | Hsp70 and tropomyosin domains | ER ? | this work |


| Ensangp00000011593 ${ }^{\text {e }}$ | Wilm's tumor 1 associating WT1 associated pr splicing regulator female lethal 2-D homolog | $\begin{aligned} & 33.55 / 4.7 \\ & 8 \end{aligned}$ | LC MS/MS | - | - | - | FTPDSNTGKR | Potential role nuclear in <br> transcriptiona <br> 1 regulation <br> Involves in <br> alternative <br> splicing <br> regulation | this work |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |

${ }^{\text {a) }}$ When several spots corresponded to the same protein, the percentage range of the sequence coverage is indicated in parenthesis. ${ }^{\text {b) }}$ Subcellular localization is inferred from sequence or structure similarity with orthologous proteins. ${ }^{\text {c) }}$ Identification was performed using Ensembl database v35 of november 2005 . ${ }^{\text {d) }}$ proteins identified from salivary gland extracts of young blood-fed females. ${ }^{\text {e) }}$ Proteins identified from salivary gland extracts of olf blood-fed females. ${ }^{\text {f) }}$ Proteins allowing a correction of incorrect genome annotation (the part of the sequence in bold is that described in Ensembl v43. PSD : post source decay. Shaded lines : Proteins identified for the first time by a proteomic approach. * means that the proteins were also identified in saliva. References are underlined when they correspond to proteins identified in human saliva.


Salivary components were separated by a $12 \%$ NU-PAGE Bis-Tris gel under denaturating and reducing conditions. Molecular mass markers are shown on the left. After Coomassie staining, the gel was cut into millimeter slices as indicated by the numbers on the right side of the figure. The plugs obtained were analyzed by mass spectrometry as described in the Methods section.


Saliva was collected from 7200 females using artificial feeders. After lyophilisation, saliva components were re-suspended in water and aliquots were analyzed by SDS-PAGE. Following silver nitrate staining, the numbered protein bands were analyzed by mass spectrometry

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A


B


A


B


